# Fundamental Determinants of the Accuracy of Equilibrium Constants for Affinity Complexes 

Tong Ye Wang, Hongchen Ji, Daniel Everton, An T. H. Le, Svetlana M. Krylova, René Fournier, and Sergey N. Krylov*



Cite This: Anal. Chem. 2023, 95, 15826-15832


Read Online

| ACCESS | \|lll Metrics \& More | (国) Article Recommendations | (si) Supporting Information |
| :---: | :---: | :---: | :---: |


#### Abstract

The equilibrium constant of a chemical reaction is arguably the key thermodynamic parameter in chemistry; we naturally expect that equilibrium constants are determined accurately. The majority of equilibrium constants determined today are those of binding reactions that form affinity complexes, such as protein-protein, protein-DNA, and protein-small molecule. There is growing awareness that the determination of  equilibrium constants for highly stable affinity complexes may be very inaccurate. However, fundamental (i.e., method-independent) determinants of accuracy are poorly understood. Here, we present a study that explicitly shows what the accuracy of equilibrium constants of affinity complexes depends on. This study reveals the critical importance of the choice of concentration of interacting components and creates a theoretical foundation for improving the accuracy of the equilibrium constants. The predicted influence of concentrations on accuracy was confirmed experimentally. The results of this fundamental study provide instructive guidance for experimentalists independently on the method they use.


Equilibrium constants of chemical reactions are the cornerstone of chemical thermodynamics. We have seen them multiple times through the years of studies, starting with a freshmen-chemistry course. We learn how to calculate them using tabulated standard free energies of formation, typically presented with three to four significant figures. Accordingly, we naturally expect that equilibrium constants are accurately determined parameters. The real world, however, is very far from this utopia.

The vast majority of equilibrium constants that are published in the scientific literature today are those of binding reactions involving large biological molecules and forming highly stable affinity complexes. ${ }^{1-4}$ These equilibrium constants cannot be calculated because standard free energies of formation of typical interactants, such as proteins, DNA, drug leads, etc., are unknown. Therefore, equilibrium constants of affinity complexes are determined experimentally using a large scope of methods. ${ }^{5-9}$ Most of these methods are supported by commercial instruments in which a researcher essentially loads the solutions into the instrument, pushes a button, and reads the resulting value with multiple significant figures and a small standard deviation. While being enormously powerful and informative tools, user-friendly commercial instruments inadvertently create an illusion that the determination of equilibrium constants is a trivial technical step, which requires only following an instrument manual. Accordingly, equilibrium-constant values published in a single paper are typically determined with a single method and presumed to be accurate values. However, discrepancies between the values reported by different laboratories for the
same complex may reach orders of magnitude leading to misconceptions and wrong conclusions, e.g., about the potency of drug leads. ${ }^{10,11}$ Great variations are often observed even for the values determined with the same method, suggesting that fundamental (method-independent) sources of inaccuracy play a critical role in inconsistencies of equilibrium constants for highly stable affinity complexes. ${ }^{12,13}$ There is an understanding that the amount of inconsistent data grows rapidly due to the use of high-throughput methods. ${ }^{12,14,15}$ The described "crisis of inaccuracy" in the field of thermodynamics of affinity complexes calls for a systematic search for sources of inaccuracy and approaches that could improve the accuracy. This work focuses on fundamental sources of inaccuracy of the equilibrium constants for affinity complexes.

In this study, we will use notations that are typically used in the field of affinity interactions. The interacting species is called a target and a ligand. Their binding with the formation of the target-ligand complex will be characterized by the equilibrium dissociation constant $\left(K_{\mathrm{d}}\right)$

$$
\begin{equation*}
\text { Target }+ \text { Ligand } \underset{K_{\mathrm{d}}}{\rightleftharpoons} \text { Complex } \tag{1}
\end{equation*}
$$

[^0]
$K_{\mathrm{d}}$ is defined as
\[

$$
\begin{equation*}
K_{\mathrm{d}}=T L / C \tag{2}
\end{equation*}
$$

\]

where $T, L$, and $C$ are the equilibrium concentrations of the target, ligand, and complex, respectively, in the binding reaction (eq 1). Lower $K_{\mathrm{d}}$ values correspond to greater complex stability and are typically desired. ${ }^{16-18}$ The values of $K_{\mathrm{d}}$ define therapeutic concentrations of drugs and analytical parameters of diagnostic methods. ${ }^{18-21}$ Therefore, they are used not only for relative ranking of the ligands but also for assessment of their suitability for intended applications. ${ }^{19,20,22}$ The most straightforward approach to $K_{\mathrm{d}}$ determination is an equilibrium (i.e., nonkinetic) method. It involves preparing a series of equilibrium mixtures of targets and ligands in which the total concentration of the ligand $\left(L_{0}\right)$ is the same, while the total concentration of the target $\left(T_{0}\right)$ varies. These mixtures are used to build a binding isotherm (dots in Figure 1), i.e., the experimental dependence of a fraction $R$ of the


Figure 1. Example of determining $K_{d}$ with a binding isotherm. A binding isotherm is the experimental dependence of the fraction of the unbound ligand ( $R$ ) on the total target concentration $\left(T_{0}\right)$, which is represented by black dots. The value of $K_{d}$ is determined by the best fit (red line) of the isotherm with eq 3 .
unbound ligand ( $R=L / L_{0}$ ) on $T_{0}$ for constant $L_{0}$. The value of $K_{d}$ is then typically found by fitting the binding isotherm with the theoretical dependence of $R$ on $T_{0}{ }^{23-25}$

$$
\begin{equation*}
R=-\frac{K_{\mathrm{d}}+T_{0}-L_{0}}{2 L_{0}}+\sqrt{\left(\frac{K_{\mathrm{d}}+T_{0}-L_{0}}{2 L_{0}}\right)^{2}+\frac{K_{\mathrm{d}}}{L_{0}}} \tag{3}
\end{equation*}
$$

while varying $K_{d}$ until the best fit (red line in Figure 1) is obtained. It is important to emphasize that eq 3 is obtained by solving a quadratic obtained from an expression for $K_{d}$

$$
\begin{equation*}
K_{\mathrm{d}}=\frac{T_{0}-L_{0}(1-R)}{(1 / R-1)} \tag{4}
\end{equation*}
$$

which, in turn, is obtained from first principles: definitions of $K_{d}$ (eq 2) and $R$ as well as mass balance for the target and ligand ( $T_{0}=T+C$ and $L_{0}=L+C$ ). In other words, eqs 3 and 4 are fundamental, i.e., method-independent, and obtained without assumptions, which would restrict the generality.
There are two fundamental requirements for $K_{d}$ accuracy: (i) the binding reaction (eq 1) must approach the equilibrium, and (ii) the study must be done in a so-called binding regime, which is assured by satisfying a condition of $L_{0} / K_{\mathrm{d}} \ll 1 .{ }^{12,26-28}$ According to a recent review by Jarmoskaite et al., ${ }^{12}$ of 100 publications dealing with $K_{\mathrm{d}}$ determination, fewer than 10 and $5 \%$ of publications reported
the satisfying of these two requirements. Failure to satisfy fundamental requirements may deem the vast majority of the published $K_{d}$ values to be greatly inaccurate. The alarming level of researchers' ignoring the fundamental requirement is due to the lack of basic knowledge of how this can affect the accuracy of $K_{d}$. There are a few important contributions to the field, which assess $K_{d}$ accuracy for specific methods, ${ }^{29-34}$ e.g., isothermal titration calorimetry ${ }^{28,29}$ and capillary electrophoresis. ${ }^{30-32}$ However, there are no comprehensive studies that would explain on what errors of $K_{d}$ are fundamentally dependent in the way that would further guide theorists and instruct experimenters. Accordingly, the field of experimental determination of $K_{\mathrm{d}}$ is largely an art in which researchers rely mostly on mnemonic rules and intuition in designing experiments and interpreting their results. The goal of our work was to initiate a graduate maturation of this field into a solid quantitative science through the understanding of what fundamental errors of $K_{d}$ depend on and what it means for experimentalists.

## - MATERIALS AND METHODS

Chemicals, Materials, and Solutions. All chemicals were purchased from Sigma-Aldrich (Oakville, ON, Canada) unless otherwise stated. Fused silica capillaries with an inner diameter of $75 \mu \mathrm{~m}$ and an outer diameter of $360 \mu \mathrm{~m}$ were purchased from Molex Polymicro (Phoenix, AZ, USA) and used throughout this work. His-tagged recombinant Thermus aquaticus MutS protein ( $\mathrm{MW}=92.8 \mathrm{kDa}$ ) was purchased from Prospec (Ness-Ziona, Israel). An Alexa488-labeled MutS-binding aptamer was synthesized by Integrated DNA Technologies (Coralville, IA, USA) with the following sequence: $5^{\prime}$-Alexa488-CTT CTG CCC GCC TCC TTC CTG GTA AAG TCA TTA ATA GGT GTG GGG TGC CGG GCA TTT CGG AGA CGA GAT AGG CGG ACA CT-3'. ${ }^{35}$ The aptamer stock solution was subjected to annealing by incubating at $90{ }^{\circ} \mathrm{C}$ for 2 min before cooling it to $20^{\circ} \mathrm{C}$ at a rate of $0.5{ }^{\circ} \mathrm{C} / \mathrm{s}$ prior to dilution and preparation of equilibrium mixtures. One single sample buffer (SB) was used for all experimental procedures: 50 mM TrisAcetate at pH 8.2 was supplemented with $0.1 \%$ Tween 20 and $0.1 \%$ Tween 80 to reduce the adsorption of DNA and MutS protein to surfaces. All solutions were made using double-distilled water deionized with Milli-Q and filtered through a $0.22 \mu \mathrm{~m}$ filter (Millipore, Nepean, ON, Canada); we termed it $\mathrm{ddH}_{2} \mathrm{O}$.

Poly(vinyl alcohol) (PVA) Coating. To reduce sample adsorption, the inner wall of all capillaries used in capillary electrophoresis (CE) was coated with poly(vinyl alcohol) (PVA) using a previously described procedure. ${ }^{36}$ PVA (5\%, $\mathrm{w} / \mathrm{v}$ ) was prepared by dissolving the polymer in boiling $\mathrm{ddH}_{2} \mathrm{O}$. Each time, two 130 cm long capillaries were coated simultaneously. The two uncoated fused silica capillaries were sequentially flushed with 0.1 M NaOH and $\mathrm{ddH}_{2} \mathrm{O}$ for 1 h under a pressure of 12 psi . The pretreated capillaries were then flushed with the PVA solution for 10 min at 15 psi and emptied using a 10 psi nitrogen gas flow for 10 min . PVA was immobilized on the capillary surface by drying overnight in an oven set at $140{ }^{\circ} \mathrm{C}$ and continuously flushed with nitrogen at a pressure of 5 psi . The detection window on the capillary was made by removing the outer polyimide layer with a fuming solution of $\mathrm{H}_{2} \mathrm{SO}_{4}$.

Experimental Procedure. All CE experiments were performed with a P/ACE MDQ apparatus (SCIEX, Concord,

Ontario, Canada) equipped with a laser-induced fluorescence (LIF) detection system. Fluorescence was excited with a blue line $(488 \mathrm{~nm})$ of a solid-state laser and detected at 520 nm . All capillaries were PVA-coated and had a length of 50 cm (approximately 40 cm from the capillary inlet to the detector). Prior to each run, the capillary was rinsed with running buffer at a pressure of 20 psi for 3 min .
All dilutions of sample components were made using SB. All equilibrium mixtures were prepared to final volumes of 40 $\mu \mathrm{L}$ in SB and incubated at an ambient temperature of $23 \pm 1$ ${ }^{\circ} \mathrm{C}$ for a minimum of 1 h prior to injection. A sample plug was injected into a capillary by pressure at 1.0 psi for 10 s (sample plug's length $=1.3 \mathrm{~cm}$ and volume $=59 \mathrm{~nL}$ ) and then propagated at 0.9 psi for 30 s (propagation distance $=$ 3.6 cm ) with SB to pass the uncooled region of the capillary. The sample was then electrophoresed at 25 kV for 15 min with a negative electrode in the capillary inlet. The capillary coolant was set to $15{ }^{\circ} \mathrm{C}$.

## - RESULTS AND DISCUSSION

Theoretical Problem. $K_{\mathrm{d}}$ values are not measured; they are determined using eqs 3 or 4 from known values of $T_{0}, L_{0}$, and $R$ (in the case of eq 3 , we also need to use nonlinear regression). Accordingly, a deviation of the determined $K_{d}$ value ( $K_{\mathrm{d}, \mathrm{det}}$ ) from the true $K_{\mathrm{d}}$ value ( $K_{\mathrm{d}}$ ) is a result of errors of $T_{0}, L_{0}$, and $R$. So, the answer to the question of what the accuracy of $K_{d}$ depends on is simple: it depends on the accuracy of $T_{0}, L_{0}$, and $R$. However, there is a second important question: how does the error of $K_{\mathrm{d}}$ depend on the errors of $T_{0}, L_{0}$, and $R$ ? If the errors of $T_{0}, L_{0}$, and $R$ are not zero, which is always the case, then the error of $K_{d}$ is the result of propagation of errors of $T_{0}, L_{0}$, and $R$. Therefore, the answer to the second question is also conceptually simple: the dependence of the accuracy of $K_{d}$ on the errors of $T_{0}, L_{0}$, and $R$ is defined by the rules of error propagation. Accordingly, we will apply error propagation rules to eq 4 to find how the deviation of $K_{\mathrm{d}, \text { det }}$ from $K_{\mathrm{d}}\left(\Delta K_{\mathrm{d}}=K_{\mathrm{d}, \text { det }}-K_{\mathrm{d}}\right)$ depends on deviations of $T_{0}, L_{0}$, and $R\left(\Delta T_{0}, \Delta L_{0}\right.$, and $\Delta R$, respectively, defined similarly to $\Delta K_{\mathrm{d}}$ ) from their true values.
Note that deviations $\Delta T_{0}, \Delta L_{0}$, and $\Delta R$ are not random errors with a given error distribution; they can be both positive and negative. $\Delta K_{\mathrm{d}}$ is a systematic error, which defines the accuracy of $K_{\mathrm{d}, \mathrm{det}} \Delta K_{\mathrm{d}}$ may be much greater than the random error of $K_{\mathrm{d}, \mathrm{det}}$. Furthermore, $\Delta K_{\mathrm{d}}$ is virtually impossible to determine since the true $K_{d}$ value is fundamentally unknown owing to the absence of standard reference instruments for determination of $K_{d}$ or standard reference $K_{d}$ values.

Propagation of Correlated Errors. Since the sample preparation processes for both targets and ligands are similar and $R$ values are determined from the signals of bound and unbound ligands, the error sources for $T_{0}, L_{0}$, and $R$ are linearly correlated in $K_{\mathrm{d}}$ determination experiments. Therefore, to investigate the error of $K_{\mathrm{d}}$ propagated from the errors of $T_{0}, L_{0}$, and $R$, we can write the general dependence of $\Delta K_{\mathrm{d}}$ on $\Delta T_{0}, \Delta L_{0}$, and $\Delta R$ as

$$
\begin{equation*}
\left|\Delta K_{\mathrm{d}}\right|=\left|\left(\frac{\partial K_{\mathrm{d}}}{\partial T_{0}}\right) \Delta T_{0}\right|+\left|\left(\frac{\partial K_{\mathrm{d}}}{\partial L_{0}}\right) \Delta L_{0}\right|+\left|\left(\frac{\partial K_{\mathrm{d}}}{\partial R}\right) \Delta R\right| \tag{5}
\end{equation*}
$$

By applying the error propagation rule (eq 5) to eq 4, and with a series of mathematical transformations (Note S2), we finally obtain

$$
\begin{align*}
& \left|\frac{\Delta K_{\mathrm{d}}}{K_{\mathrm{d}}}\right|=a+b \frac{L_{0}}{K_{\mathrm{d}}}, \\
& \text { For } 0 \leq \Delta R<0.5: \\
& a=t+\frac{r(1+r)}{0.5+r}, b=\frac{0.5(t+l-r)}{1+r}+\frac{0.5 r}{0.5+r} \\
& \text { For }-0.5<\Delta R<0: \\
& a=t+\frac{r(1-r)}{0.5-r}, b=\frac{0.5(t+l-r)}{1-r}+\frac{0.5 r}{0.5-r} \tag{6}
\end{align*}
$$

where $t=\left|\Delta T_{0} / T_{0}\right|, l=\left|\Delta L_{0} / L_{0}\right|$, and $r=|\Delta R / R|$. Equation 6 reveals that, when the error sources for $T_{0}, L_{0}$, and $R$ are strongly correlated, the relative error of $K_{d}$ is a linear function of $L_{0} / K_{\mathrm{d}}$ with an intercept dependent only on relative errors of $T_{0}$ and $R$ and slope-dependent on relative errors of all three variables: $T_{0}, L_{0}$, and $R$. In the mathematical analysis (Note S2), we made a single simplifying assumption that the determined (from measured signals) value of $R$ is equal to 0.5 , which leads to the least erroneous $K_{d} ;{ }^{25}$ thus, we are considering the best-case scenario and are finding the lower limit for the error of $K_{d}$.

Equation 6 explicitly shows the role of $L_{0} / K_{d}$ in the relative error of $K_{\mathrm{d}}$. The value of $L_{0} / K_{\mathrm{d}}$ is important only when the second term is greater than the first, i.e., when $L_{0} / K_{d}>a / b$. In other words, in contrast to a commonly used requirement of $L_{0} / K_{\mathrm{d}} \ll 1$, it is unnecessary to decrease $L_{0} / K_{d}$ much below $a / b$. Rather, using $L_{0} / K_{\mathrm{d}} \ll a / b$ will likely cause the opposite effect, namely, it will lead to an increase in $\left|\Delta K_{d} / K_{d}\right|$ due to the increase in $r$ when $L_{0}$ is too low, causing an unacceptable signal-to-noise ratio $(S / N)$, which will be briefly discussed later. On the other hand, when $L_{0} / K_{d}>a / b$, the increase in $L_{0} / K_{\mathrm{d}}$ plays a crucial role in increasing the relative error of $K_{\mathrm{d}}$; an order of magnitude increase in $L_{0}$ will lead to an order of magnitude increase in $\left|\Delta K_{d} / K_{d}\right|$. One should appreciate that if $L_{0} / K_{\mathrm{d}} \gg a / b$, then $K_{\mathrm{d}, \text { det }}$ may differ from true $K_{d}$ by orders of magnitude.

For a large range of $L_{0}$ values, which is typically the case, it is more convenient to present the dependence of $\left|\Delta K_{d} / K_{d}\right|$ on $L_{0} / K_{\mathrm{d}}$ on a double-log scale. A graph of $\log \left(\left|\Delta K_{\mathrm{d}} / K_{\mathrm{d}}\right|\right)$ versus $\log \left(L_{0} / K_{\mathrm{d}}\right)$ is triphasic: two asymptotically linear ranges flank a nonlinear transition range (Figure 2a and Figure S1a). For small values of $L_{0} / K_{\mathrm{d}}$, i.e., for $L_{0} / K_{\mathrm{d}} \ll a / b$, the dependence is a linear function with no dependence on $L_{0} / K_{\mathrm{d}}: \log \left(\left|\Delta K_{\mathrm{d}} / K_{\mathrm{d}}\right|\right)=\log (a)=$ const. For a large value of $L_{0} / K_{d}$, i.e., for $L_{0} / K_{d} \gg a / b$, the dependence is a linear function $\log \left(\left|\Delta K_{\mathrm{d}} / K_{\mathrm{d}}\right|\right)=\log (b)+\log \left(L_{0} / K_{\mathrm{d}}\right)$ with an intercept with the ordinate equal to $\log (b)$ and a slope equal to unity. The abscissa of the intersection of the two asymptotic lines is defined by $\log \left(L_{0} / K_{\mathrm{d}}\right)=\log (a / b)$.

Using eq 6 and $\Delta K_{\mathrm{d}}=K_{\mathrm{d}, \text { det }}-K_{\mathrm{d}}$, we can show that (for strongly correlated error sources) not only $\left|\Delta K_{d} / K_{d}\right|$ depends linearly on $L_{0} / K_{d}$ but $K_{\mathrm{d} \text {,det }}$ is also a linear function of $L_{0}$ (Note S3)


Figure 2. General trends in dependencies of $\log \left(\left|\Delta K_{d} / K_{d}\right|\right)$ on $\log \left(L_{0} / K_{\mathrm{d}}\right)(\mathrm{a}, \mathrm{c})$ and $\log \left(K_{\mathrm{d}, \mathrm{det}}\right)$ on $\log \left(L_{0}\right)(\mathrm{b}, \mathrm{d})$. In all panels, reasonable relative errors of $L_{0}, T_{0}$, and $R$ were used: $\left|\Delta T_{0} / T_{0}\right|=$ $\left|\Delta L_{0} / L_{0}\right|=0.05$ and $\Delta R / R=0.02$ (the results corresponding to $\Delta R / R=-0.02$ are shown in Figure S1a,c). In panels (b, d), we set $K_{\mathrm{d}}=1$ (unitless) and $\Delta K_{\mathrm{d}} \geq 0$ (the results corresponding to $\Delta K_{\mathrm{d}}<$ 0 are shown in Figure $S 1 b, d)$. With these conditions, parameters $a$, $b, c$, and $d$ were calculated to be $0.089,0.058,1.1$, and 0.058 , respectively; $\alpha, \beta$, and $\gamma$ were calculated to be $0.064,0.036$, and 1.1, respectively.

$$
\begin{align*}
& K_{\mathrm{d}, \mathrm{det}}=c+d L_{0} \\
& \text { For } \Delta K_{\mathrm{d}} \geq 0: c=(1+a) K_{\mathrm{d}}, d=b \\
& \text { For } \Delta K_{\mathrm{d}}<0: c=(1-a) K_{\mathrm{d}}, \quad d=-b \tag{7}
\end{align*}
$$

If presented in a double-log scale, then this dependence is expectedly triphasic (Figure 2b) and has features similar to the dependence of $\log \left(\left|\Delta K_{d} / K_{d}\right|\right)$ on $\log \left(L_{0} / K_{d}\right)$ (Figure 2a).

Propagation of Uncorrelated Errors. In some experimental cases, the error sources of $L_{0}, T_{0}$, and $R$ may be weekly correlated. For example, stock concentrations of the ligand and target may largely differ from the labeled values in the opposite direction. In these cases, the errors from the stock reagents are uncorrelated (independent of each other) and might be much larger than the errors caused by the instruments or methods used in sample preparation. In addition, using an uncalibrated instrument to detect ligand signals can cause the errors in determined $R$ values to be uncorrelated to the errors of $T_{0}$ and $L_{0}$. To study the error propagation with uncorrelated (orthogonal) error sources, we should write the dependence of $\Delta K_{\mathrm{d}}$ on $\Delta T_{0}, \Delta L_{0}$, and $\Delta R$ as

$$
\begin{equation*}
\left|\Delta K_{\mathrm{d}}\right|=\sqrt{\left(\frac{\partial K_{\mathrm{d}}}{\partial T_{0}}\right)^{2} \Delta T_{0}^{2}+\left(\frac{\partial K_{\mathrm{d}}}{\partial L_{0}}\right)^{2} \Delta L_{0}{ }^{2}+\left(\frac{\partial K_{\mathrm{d}}}{\partial R}\right)^{2} \Delta R^{2}} \tag{8}
\end{equation*}
$$

Then, we apply the error propagation rule of eqs 8-4 and obtain (Note S4)

$$
\left|\frac{\Delta K_{\mathrm{d}}}{K_{\mathrm{d}}}\right|=\sqrt{\alpha^{2}+\lambda \frac{L_{0}}{K_{\mathrm{d}}}+\beta^{2}\left(\frac{L_{0}}{K_{\mathrm{d}}}\right)^{2}}
$$

For $0 \leq \Delta R<0.5$ :

$$
\begin{align*}
& \alpha=\sqrt{t^{2}+\left(\frac{r+r^{2}}{0.5+r}\right)^{2}} \\
& \beta=\frac{0.5}{1+r} \sqrt{t^{2}+l^{2}+\left(\frac{0.5 r}{0.5+r}\right)^{2}} \\
& \lambda=\frac{t^{2}}{1+r}+2\left(\frac{0.5 r}{0.5+r}\right)^{2} \\
& \text { For-0.5< } \Delta R<0: \\
& \alpha=\sqrt{t^{2}+\left(\frac{r-r^{2}}{r-0.5}\right)^{2}} \\
& \beta=\frac{0.5}{1-r} \sqrt{t^{2}+l^{2}+\left(\frac{0.5 r}{r-0.5}\right)^{2}} \\
& \lambda=\frac{t^{2}}{1-r}+2\left(\frac{0.5 r}{r-0.5}\right)^{2} \tag{9}
\end{align*}
$$

Here, as explained in the previous section, $t=\left|\Delta T_{0} / T_{0}\right|, l=$ $\left|\Delta L_{0} / L_{0}\right|$, and $r=|\Delta R / R|$ and we only investigate the case with the determined $R$ being equal to 0.5 to simplify the study.

Although the forms of eqs 6 and 9 are very different, eq 9 shows a significant similarity to eq 6 regarding the triphasic dependence of $\left|\Delta K_{\mathrm{d}} / K_{\mathrm{d}}\right|$ on $L_{0} / K_{\mathrm{d}}$. For small $L_{0} / K_{\mathrm{d}}$ values, eq 9 approaches a linear function of $\left|\Delta K_{\mathrm{d}} / K_{\mathrm{d}}\right|=\alpha=$ const that shows no dependence on $L_{0} / K_{\mathrm{d}}$. For large $L_{0} / K_{\mathrm{d}}$ values, eq 9 approaches a linear function of $\mid \Delta K_{\mathrm{d}} / K_{\mathrm{d}} \mathrm{I}=\beta\left(L_{0} / K_{\mathrm{d}}\right)$ that shows a high sensitivity of $\left|\Delta K_{\mathrm{d}} / K_{\mathrm{d}}\right|$ to $L_{0} / K_{\mathrm{d}}$. There is a nonlinear transition range between these two linear lines. Moreover, both eqs 6 and 9 suggest that the minimum $\mid \Delta K_{\mathrm{d}} /$ $K_{\mathrm{d}} \mathrm{l}$ value depends on a parameter ( $a$ in eq 6 or $\alpha$ in eq 9) that is defined only by $t$ and $r$, while the sensitivity of $\mid \Delta K_{\mathrm{d}} /$ $K_{\mathrm{d}} \mathrm{l}$ to $L_{0} / K_{\mathrm{d}}$ mainly depends on a parameter ( $b$ in eq 6 or $\beta$ in eq 9) that is defined by $t, l$, and $r$.

To show the dependence of $\left|\Delta K_{\mathrm{d}} / K_{\mathrm{d}}\right|$ on $L_{0} / K_{\mathrm{d}}$ in a large range of $L_{0} / K_{\mathrm{d}}$ for uncorrelated error sources, we demonstrate $" \log \left(\left|\Delta K_{\mathrm{d}} / K_{\mathrm{d}}\right|\right)$ versus $\log \left(L_{0} / K_{\mathrm{d}}\right)$ " in Figure 2c. In the double-log scale (Figure 2c), the asymptotic lines for small and large $L_{0} / K_{\mathrm{d}}$ values are approximated with functions of $\log \left(\left|\Delta K_{\mathrm{d}} / K_{\mathrm{d}}\right|\right)=\log (\alpha)$ and $\log \left(\left|\Delta K_{\mathrm{d}} / K_{\mathrm{d}}\right|\right)=$ $\log (\beta)+\log \left(L_{0} / K_{d}\right)$, respectively. Figure 2c suggests that, to avoid a significant increase in $r$ caused by an unacceptably low signal-to-noise ratio, it is unnecessary to decrease $L_{0} / K_{\mathrm{d}}$ much below a certain value $\alpha / \beta$ that is defined by the abscissa of intersection of the two asymptotic lines (i.e., $\left.\log \left(L_{0} / K_{\mathrm{d}}\right)=\log (\alpha / \beta)\right)$. According to the results in Figure 2 and Figure S1, with the same $t, l$, and $r$, the value of $\alpha / \beta$ is slightly greater than $a / b$ because $\left|\Delta K_{\mathrm{d}}\right|$ obtained with eq 8 is smaller than that determined with eq 5 .

Now, let us discuss how $K_{\mathrm{d}, \text { det }}$ depends on $L_{0}$ when the error sources are uncorrelated. With eq 9 and the definition of $\Delta K_{\mathrm{d}}$ (i.e., $\Delta K_{\mathrm{d}}=K_{\mathrm{d}, \text { det }}-K_{\mathrm{d}}$ ), we obtain (Note S5)

$$
\begin{align*}
K_{\mathrm{d}, \text { det }} & =K_{\mathrm{d}}+\sqrt{\alpha^{2} K_{\mathrm{d}}^{2}+\lambda K_{\mathrm{d}} L_{0}+\beta^{2} L_{0}^{2}} \text { when } \Delta K_{\mathrm{d}} \geq 0 \\
& =K_{\mathrm{d}}-\sqrt{\alpha^{2} K_{\mathrm{d}}^{2}+\lambda K_{\mathrm{d}} L_{0}+\beta^{2} L_{0}^{2}} \text { when } \Delta K_{\mathrm{d}}<0 \tag{10}
\end{align*}
$$

Equation 10 shows that the dependence of $K_{\mathrm{d}, \text { det }}$ on $L_{0}$ is also triphasic. For small $L_{0}$ values, $K_{\mathrm{d} \text {,det }}$ is insensitive to the change of $L_{0}$ and eq 10 approaches a linear function of $K_{\mathrm{d}, \text { det }}$ $=\gamma\left(\gamma=(1+\alpha) K_{d}\right.$ when $\Delta K_{d} \geq 0$ and $\gamma=(1-\alpha) K_{d}$ when $\left.\Delta K_{\mathrm{d}}<0\right)$ (Note S5). For large $L_{0}$ values, eq 10 approaches another linear function of $K_{\mathrm{d}, \text { det }}=\beta L_{0}$ that shows high sensitivity of $K_{\mathrm{d} \text { det }}$ to $L_{0}$ (only applicable for $\Delta K_{\mathrm{d}} \geq 0$ since $K_{\mathrm{d} \text { det }}$ has to be greater than zero) (Note S5). A nonlinear transition range exists between the two linear lines. Figure 2d shows an example of the dependence of $K_{\mathrm{d} d \mathrm{det}}$ on $L_{0}$ for uncorrelated error sources of $T_{0}, L_{0}$, and $R$ on a double-log scale, which has the expected triphasic feature.

Experimental Validation. The dependence shown in Figure $2 \mathrm{~b}, \mathrm{~d}$ has two known variables, $L_{0}$ and $K_{\mathrm{d}, \mathrm{det}}$ and therefore, it can be tested experimentally, which we have undertaken in this work. Experimentally, we used a wellestablished solution-based method of nonequilibrium capillary electrophoresis of equilibrium mixtures (NECEEM) as the $K_{\mathrm{d}}$-determination method and MutS-aptamer as the binding pair. ${ }^{25,37-39}$ In NECEEM, the unbound ligand and complex (bound ligand) are separated by an electric field in a capillary due to their different charge-to-size ratios. ${ }^{37-39}$ In six NECEEM experiments, the aptamer concentration $\left(L_{0}\right)$ was kept constant at $0.02,0.05,0.1,0.5,2$, and 10 nM ; the MutS concentration $\left(T_{0}\right)$ varied from 0 to 312.5 nM (all concentrations are given as nominal values with no significance to the number of digits). The lowest value of $L_{0}$ (i.e., 0.02 nM ) was chosen to be equal to the limit of quantitation (LOQ) (Figure S2) linked to $S / N$ as discussed below. The influence of $L_{0}$ on binding isotherms and the dependence of $K_{\mathrm{d}, \mathrm{det}}$ on $L_{0}$ are shown in Figure 3. Figure 3b shows that (in a double-log scale) $K_{\mathrm{d}, \text { det }}$ is insensitive to the change of $L_{0}$ when $L_{0}$ is at low concentrations (e.g., $\ll \sim 0.1$ nM ), and $K_{\mathrm{d}, \mathrm{det}}$ is sensitively dependent on $L_{0}$ when $L_{0}$ is at high concentrations. The results in Figure 3b are consistent with the theoretical predictions shown in Figure 2b,d, which confirm the validity of our theoretical analysis.

Instructive Guidance for Experimentalists. Although Figure 2 suggests that, with any type of experimental error sources (correlated or uncorrelated), we can always use an as-low-as-possible $L_{0}$ to avoid the error of $K_{\mathrm{d}, \text { det }}$ being greatly magnified, the choice of the lowest experimentally suitable $L_{0}$ is dictated by the limit of quantitation (LOQ) of an instrument used to measure the signal. The value of LOQ is, by definition, the analyte concentration (ligand concentration $L_{0}$ in our case) for which $S / N$ is equal to a certain value $X \gg$ 1, which guarantees that the noise does not affect quantitation significantly. If the noise is independent of the signal and the value of LOQ is known, then $S / N$ can be calculated for any given concentration of the ligand as $S / N=$ $X L_{0} /$ LOQ (Note S6). The minimum acceptable ligand concentration that satisfies this equality is $L_{0}=$ LOQ. Since $R$ values are determined based on the measured signals $S$, using $L_{0}<\mathrm{LOQ}$ is counterproductive as it will cause a significant increase in the relative error of $R(r)$, and, accordingly, in the relative error of $K_{\mathrm{d}, \text { det }}$.


Figure 3. Influence of $L_{0}$ on binding isotherms (a) and the dependence of $K_{\text {d,det }}$ on $L_{0}$ (b). In panel (a), all nonlinear regressions were conducted using OriginPro software with the iteration algorithm of Levenberg-Marquardt. See the Supporting Information for other details.

Based on the above analysis, to achieve good accuracy of $K_{\mathrm{d}, \mathrm{det}}$ in an experiment, if $\mathrm{LOQ} / K_{\mathrm{d}}<a / b$ (or $\alpha / \beta$ for uncorrelated error sources), then $L_{0} / K_{\mathrm{d}}$ should satisfy

$$
\begin{equation*}
\frac{\mathrm{LOQ}}{K_{\mathrm{d}}} \leq \frac{L_{0}}{K_{\mathrm{d}}}<\frac{a}{b}\left(\text { or } \frac{\alpha}{\beta}\right) \tag{11}
\end{equation*}
$$

where $a, b, \alpha$, and $\beta$ are defined by the absolute values of relative systematic errors of $T_{0}, L_{0}$, and $R$ (i.e., $t, l$, and $r$ ) as shown in eqs 6 and 9. Because $t, l$, and $r$ are relatively difficult to determine, practically, the LOQ can be determined and used as the sole determinant of the ligand concentration to minimize the error of $K_{\mathrm{d}, \mathrm{det}}$ in a $K_{\mathrm{d}}$ determination experiment. It is important to emphasize that using $L_{0}=$ LOQ does not imply that $L_{0} / K_{\mathrm{d}}<a / b($ or $\alpha / \beta$ ) and thus does not guarantee the accuracy of $K_{\mathrm{d}}$. Therefore, with the current state of affairs, the only way to confirm (indirectly) that $L_{0} / K_{\mathrm{d}}<a / b$ (or $\alpha / \beta$ ) is to conduct experiments not only at $L_{0}=$ LOQ but also at $L_{0}>$ LOQ (e.g., the experiments corresponding to $L_{0}=0.02$ and 0.05 nM shown in Figure 3). If $K_{d}$ is not affected by the value of $L_{0}$, then one can assume that $L_{0} / K_{\mathrm{d}}<a / b$ (or $\alpha / \beta$ ) and deem $K_{d}$ accurate.

## ■ CONCLUDING REMARKS

To conclude, in this study, we investigated the theoretical dependence of the relative deviation of $K_{\mathrm{d}, \mathrm{det}}$ from $K_{\mathrm{d}}\left(I \Delta K_{\mathrm{d}} /\right.$ $K_{\mathrm{d}} \mathrm{l}$ ) on the ratio of $L_{0} / K_{\mathrm{d}}$ with error propagations. Our analysis shows that with fixed correlated or uncorrelated
errors of $T_{0}, L_{0}$, and $R,\left|\Delta K_{\mathrm{d}} / K_{\mathrm{d}}\right|$ always presents a triphasic dependence on $L_{0} / K_{\mathrm{d}}$ in a double-log scale: When $L_{0} / K_{\mathrm{d}}$ is small, $\left|\Delta K_{\mathrm{d}} / K_{\mathrm{d}}\right|$ is insensitive to the change of $L_{0} / K_{\mathrm{d}}$ and approaches a constant that is defined by the errors of $T_{0}$ and $R$. When $L_{0} / K_{\mathrm{d}}$ is large, $\left|\Delta K_{\mathrm{d}} / K_{\mathrm{d}}\right|$ is sensitive to the change of $L_{0} / K_{\mathrm{d}}$, i.e., an order of magnitude increase in $L_{0}$ will lead to an order of magnitude increase in $\left|\Delta K_{\mathrm{d}} / K_{\mathrm{d}}\right|$. There is a nonlinear transition range between the two linear phases. We also theoretically demonstrated that the dependence of $K_{\mathrm{d}, \mathrm{det}}$ on $L_{0}$ should also show the triphasic features on a double-log scale, which was experimentally confirmed with NECEEM experiments using MutS-aptamer as the binding pair. The results of this work suggest that, without knowing the errors of $T_{0}, L_{0}$, and $R$, experimentalists should use $L_{0}=$ LOQ (i.e., the smallest ligand concentration that does not sacrifice the accuracy of $R$ ) in an equilibrium $K_{d}$-determination method and conduct an extra experiment with $L_{0}>$ LOQ to confirm the accuracy of the determined $K_{\mathrm{d}}$. To emphasize, we foresee that our findings (e.g., eqs 6 and 9 and the triphasic dependence of $\left|\Delta K_{\mathrm{d}} / K_{\mathrm{d}}\right|$ on $L_{0} / K_{\mathrm{d}}$ in Figure 2) can help to create an approach of assessing $K_{\mathrm{d}}$ accuracy from a singlebinding isotherm if the ranges of relative errors in $T_{0}, L_{0}$, and $R$ can be estimated.

## ■ ASSOCIATED CONTENT

## (s) Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acs.analchem.3c03557.

Derivation of the theoretical dependence of the fraction of the unbound ligand on the total target concentration (Note S1); derivation of the dependence of relative systematic error of $K_{\mathrm{d}}$ on the $L_{0} / K_{\mathrm{d}}$ ratio for correlated error sources (Note S2); derivation of the theoretical dependence of determined $K_{d}$ on $L_{0}$ for correlated error sources (Note S3); derivation of the dependence of relative systematic error of $K_{d}$ on the $L_{0} / K_{d}$ ratio for uncorrelated error sources (Note S4); derivation of the theoretical dependence of determined $K_{\mathrm{d}}$ on $L_{0}$ for uncorrelated error sources (Note S5); general trends in dependencies of $\log \left(\left|\Delta K_{\mathrm{d}} / K_{\mathrm{d}}\right|\right)$ on $\log \left(L_{0} / K_{\mathrm{d}}\right)$ and $\log \left(K_{\mathrm{d}, \text { det }}\right)$ on $\log \left(L_{0}\right)$ (Figure S 1$)$; derivation of the relationship between the signal-to-noise ratio and the limit of quantitation (Note S6); determination of limit of quantitation (Figure S2); electropherograms of the NECEEM experiments (Figure S3); results of $R$ values at different $T_{0}$ for the NECEEM experiments with different $L_{0}$ (Table S1) (PDF)

## - AUTHOR INFORMATION

## Corresponding Author

Sergey N. Krylov - Department of Chemistry and Centre for Research on Biomolecular Interactions, York University, Toronto, Ontario M3J 1P3, Canada; © orcid.org/0000-0003-3270-2130; Email: skrylov@yorku.ca

## Authors

Tong Ye Wang - Department of Chemistry and Centre for Research on Biomolecular Interactions, York University, Toronto, Ontario M3J 1P3, Canada; © orcid.org/0000-0001-9462-7194
Hongchen Ji - Department of Chemistry and Centre for Research on Biomolecular Interactions, York University,

Toronto, Ontario M3J 1P3, Canada; © orcid.org/0000-0001-5835-3843
Daniel Everton - Department of Chemistry and Centre for Research on Biomolecular Interactions, York University, Toronto, Ontario M3J 1P3, Canada
An T. H. Le - Department of Chemistry and Centre for Research on Biomolecular Interactions, York University, Toronto, Ontario M3J 1P3, Canada; © orcid.org/0000-0002-3659-9938
Svetlana M. Krylova - Department of Chemistry and Centre for Research on Biomolecular Interactions, York University, Toronto, Ontario M3J 1P3, Canada; © orcid.org/0000-0002-3291-6721
René Fournier - Department of Chemistry and Centre for Research on Biomolecular Interactions, York University, Toronto, Ontario M3J 1P3, Canada
Complete contact information is available at:
https://pubs.acs.org/10.1021/acs.analchem.3c03557

## Notes

The authors declare no competing financial interest.

## ACKNOWLEDGMENTS

This work was supported by the Natural Sciences and Engineering Research Council of Canada (grant SPG-P 521331-2018).

## ABBREVIATIONS

NECEEM, nonequilibrium capillary electrophoresis of equilibrium mixtures; LOQ, limit of quantitation

## REFERENCES

(1) Almehmadi, L. M.; Valsangkar, V. A.; Halvorsen, K.; Zhang, Q.; Sheng, J.; Lednev, I. K. Anal. Bioanal. Chem. 2022, 414, 60096016.
(2) Fu, H.; Qian, C.; Tong, W.; Li, H.; Chen, D. D. Y. J. Chromatogr. A 2019, 1589, 182-190.
(3) Ramsay, R. R.; Popovic-Nikolic, M. R.; Nikolic, K.; Uliassi, E.; Bolognesi, M. L. Clin. Trans. Med. 2018, 7, 3.
(4) Arshavsky-Graham, S.; Massad-Ivanir, N.; Paratore, F.; Scheper, T.; Bercovici, M.; Segal, E. ACS Sens. 2017, 2, 1767-1773.
(5) Friguet, B.; Chaffotte, A. F.; Djavadi-Ohaniance, L.; Goldberg, M. E. J. Immunol. Methods 1985, 77, 305-319.
(6) Avila, L. Z.; Chu, Y.-H.; Blossey, E. C.; Whitesides, G. M. J. Med. Chem. 1993, 36, 126-133.
(7) Mwakibete, H.; Cristantino, R.; Bloor, D. M.; Wyn-Jones, E.; Holzwarth, J. F. Langmuir 1995, 11, 57-60.
(8) Okhonin, V.; Berezovski, M. V.; Krylov, S. N. J. Am. Chem. Soc. 2010, 132, 7062-7068.
(9) Du, X.; Li, Y.; Xia, Y.-L.; Ai, S.-M.; Liang, J.; Sang, P.; Ji, X.-L.; Liu, S.-Q. Int. J. Mol. Sci. 2016, 17, 144.
(10) Bottari, F.; Daems, E.; de Vries, A.-M.; Wielendaele, P. V.; Trashin, S.; Blust, R.; Sobott, F.; Madder, A.; Martins, J. C.; De Wael, K. J. Am. Chem. Soc. 2020, 142, 19622-19630.
(11) Fielding, L. Curr. Top. Med. Chem. 2003, 3, 39-53.
(12) Jarmoskaite, I.; AlSadhan, I.; Vaidyanathan, P. P.; Herschlag, D. eLife 2020, 9, No. e57264.
(13) Wätzig, H.; Oltmann-Norden, I.; Steinicke, F.; Alhazmi, H. A.; Nachbar, M.; El-Hady, D. A.; Albishri, H. M.; Baumann, K.; Exner, T.; Böckler, F. M.; El Deeb, S. J. Comput.- Aided Mol. Des. 2015, 29, 847-865.
(14) Fuchs, H.; Gessner, R. Biochem. 2001, 359, 411-418.
(15) Shalaeva, M.; Kenseth, J.; Lombardo, F.; Bastin, A. J. Pharm. Sci. 2008, 97, 2395-2854.
(16) Zhang, H.; Li, X.-F.; Le, X. C. Anal. Chem. 2012, 84, 877884.
(17) Nakane, R.; Kurihara, O.; Natsubori, A. J. Phys. Chem. 1964, 68, 2876-2882.
(18) Han, B. J. Biol. Chem. 2020, 295, 15280-15291.
(19) Qian, C.; Fu, H.; Kovalchik, K. A.; Li, H.; Chen, D. D. Y. Anal. Chem. 2017, 89, 9483-9490.
(20) Tang, Z.; Shangguan, D.; Wang, K.; Shi, H.; Sefah, K.; Mallikratchy, P.; Chen, H. W.; Li, Y.; Tan, W. Anal. Chem. 2007, 79, 4900-4907.
(21) Cooper, M. A. Nat. Rev. Drug Discovery 2002, 1, 515-528.
(22) Shortridge, M. D.; Hage, D. S.; Harbison, G. S.; Powers, R. J. Comb. Chem. 2008, 10, 948-958.
(23) Sisavath, N.; Rukundo, J.-L.; Le Blanc, J. C. Y.; Galievsky, V. A.; Bao, J.; Kochmann, S.; Stasheuski, A. S.; Krylov, S. N. Angew. Chem. 2019, 131, 6707-6711.
(24) Jusko, W. J.; Molins, E. A. G.; Ayyar, V. S. Drug Metab. Dispos. 2020, 48, 894-902.
(25) Kanoatov, M.; Galievsky, V. A.; Krylova, S. M.; Cherney, L. T.; Jankowski, H. K.; Krylov, S. N. Anal. Chem. 2015, 87, 30993106.
(26) Hulme, E. C.; Trevethick, M. A. Br. J. Pharmacol. 2010, 161, 1219-1237.
(27) Strohkendl, I.; Saifuddin, F. A.; Rybarski, J. R.; Finkelstein, I. J.; Russell, R. Mol. Cell 2018, 71, 816-824.
(28) Tellinghuisen, J. Anal. Biochem. 2012, 424, 211-220.
(29) Tellinghuisen, J. Anal. Biochem. 2003, 321, 79-88.
(30) Rundlett, K. L.; Armstrong, D. W. J. Chromatogr. A 1996, 721, 173-186.
(31) Bowser, M. T.; Chen, D. D. Y. J. Phys. Chem. A 1998, 102, 8063-8071.
(32) Bowser, M. T.; Chen, D. D. Y. J. Phys. Chem. A 1999, 103, 197-202.
(33) Jameson, D. M.; Mocz, G. Fluorescence Polarization/ Anisotropy Approaches to Study Protein-Ligand Interactions. In Protein-Ligand Interactions: Methods and Applications; Nienhaus, G. U., Eds.; Humana Press: Totowa, NJ, 2005; Vol. 305, pp 301-322, DOI: 10.1385/1-59259-912-5:301.
(34) Contreras-Martos, S.; Nguyen, H. H.; Nguyen, P. N.; Hristozova, N.; Macossay-Castillo, M.; Kovacs, D.; Bekesi, A.; Oemig, J. S.; Maes, D.; Pauwels, K.; Tompa, P.; Lebrun, P. Front. Mol. Biosci. 2018, 5, 83.
(35) Krylova, S. M.; Karkhanina, A. A.; Musheev, M. U.; Bagg, E. A. L.; Schofield, C. J.; Krylov, S. N. Anal. Biochem. 2011, 414, 261265.
(36) de Jong, S.; Krylov, S. N. Anal. Chem. 2012, 84, 453-458.
(37) Berezovski, M.; Nutiu, R.; Li, Y.; Krylov, S. N. Anal. Chem. 2003, 75, 1382-1386.
(38) Krylov, S. N. NECEEM for Development, Characterization and Analytical Utilization of Aptamer. LabPlus International, November 2005, 6-9.
(39) Krylov, S. N. J. Biomol. Screening 2006, 11, 115-122.

## SUPPORTING INFORMATION

Fundamental Determinants of the Accuracy of Equilibrium Constants for Affinity Complexes

Tong Ye Wang, Hongchen Ji, Daniel Everton, An T. H. Le, Svetlana M. Krylova, René Fournier, and Sergey N. Krylov*<br>Department of Chemistry and Centre for Research on Biomolecular Interactions, York University, Toronto, Ontario M3J 1P3, Canada<br>*Corresponding author's email: skrylov@yorku.ca

## Table of Contents

Section S1: Derivation of equations .....................................................................................................S2
Note S1: Derivation of the theoretical dependence of fraction of unbound ligand $(R)$ on total target
concentrat................................................................................................................................... $\left(T_{0}\right)$
Note S2: Derivation of the dependence of relative systematic error of $K_{\mathrm{d}}\left(\left|\Delta K_{\mathrm{d}} / K_{\mathrm{d}}\right|\right)$ on $L_{0} / K_{\mathrm{d}}$ ratio for correlated (parallel) error sources .S3
Note S3: Derivation of the theoretical dependence of determined $K_{\mathrm{d}}$ ( $K_{\mathrm{d}, \mathrm{det}}$ ) on $L_{0}$ for correlated (parallel) error sources. ..... S5
Note S4: Derivation of the dependence of relative systematic error of $K_{\mathrm{d}}\left(\left|\Delta K_{\mathrm{d}} / K_{\mathrm{d}}\right|\right)$ on $L_{0} / K_{\mathrm{d}}$ ratio for uncorrelated (orthogonal) error sources ..... S6
Note S5: Derivation of the theoretical dependence of determined $K_{\mathrm{d}}$ ( $K_{\mathrm{d}, \mathrm{det}}$ ) on $L_{0}$ for uncorrelated (orthogonal) error sources ..... S8
Figure S1: General trends in dependencies of $\log \left(\left|\Delta K_{\mathrm{d}} / K_{\mathrm{d}}\right|\right)$ on $\log \left(L_{0} / K_{\mathrm{d}}\right)$ and $\log \left(K_{\mathrm{d}, \mathrm{det}}\right)$ on $\log \left(L_{0}\right)$. ..... S9
Note S6: Derivation of the relationship between signal to noise ratio $(S / N)$ and the limit of quantitation (LOQ) ..... S10
Section S2: Experiments ..... S11
Figure S2: Determination of limit of quantitation (LOQ) ..... S11
Figure S3: Electropherograms of the NECEEM experiments ..... S12
Table S1: Results of $R$ values at different $T_{0}\left([\mathrm{MutS}]_{0}\right)$ for the NECEEM experiments with different $L_{0}$ ([aptamer] ${ }_{0}$ ) ..... S13
References ..... S14

## Additional supplementary files

The following supplementary files can be found on Figshare:
DOI: 10.6084/m9.figshare. 23624583

| File name | Description/Experiment |
| :--- | :--- |
| Figures.zip | Figures in main text and supporting information (Origin and png files). |

## Section S1: Derivation of equations

## Note S1: Derivation of the theoretical dependence of fraction of unbound ligand $(\boldsymbol{R})$ on total target concentration ( $\boldsymbol{T}_{0}$ )

The definition of equilibrium dissociation constant $K_{\mathrm{d}}$ and mass balance for the target and ligand show:

$$
\begin{gather*}
K_{\mathrm{d}}=\frac{T L}{C}  \tag{S1}\\
T_{0}=T+C \Rightarrow T=T_{0}-C  \tag{S2}\\
L_{0}=L+C \Rightarrow C=L_{0}-L \tag{S3}
\end{gather*}
$$

where $T, L$, and $C$ are equilibrium concentrations of the target, ligand and complex, respectively; $T_{0}$ and $L_{0}$ are the total concentrations of target and ligand, respectively. The definition of fraction of unbound ligand $R(0 \leq R \leq 1)$ is:

$$
\begin{equation*}
R=\frac{L}{L_{0}} \Rightarrow L=R L_{0} \tag{S4}
\end{equation*}
$$

By inserting Eq S4 into Eq S3, we have:

$$
\begin{equation*}
C=L_{0}-R L_{0}=L_{0}(1-R) \tag{S5}
\end{equation*}
$$

By replacing $C$ in Eq S2 with Eq S5, we obtain:

$$
\begin{equation*}
T=T_{0}-L_{0}(1-R) \tag{S6}
\end{equation*}
$$

With inserting Eqs S 4 , S 5 and S 6 into Eq S1, we obtain:

$$
\begin{align*}
K_{\mathrm{d}} & =\frac{\left[T_{0}-L_{0}(1-R)\right] R L_{0}}{(1-R) L_{0}} \\
& =\frac{\left[T_{0}-L_{0}(1-R)\right] R}{(1-R)}  \tag{S7}\\
& =\frac{T_{0}-L_{0}(1-R)}{(1 / R-1)}
\end{align*}
$$

By rearranging Eq S7, we obtain a quadratic equation for $R$ :

$$
\begin{equation*}
L_{0} R^{2}+\left(K_{\mathrm{d}}+T_{0}-L_{0}\right) R-K_{\mathrm{d}}=0 \tag{S8}
\end{equation*}
$$

With the quadratic formula, the expression of $R$ is solved as

$$
\begin{equation*}
R=\frac{-\left(K_{\mathrm{d}}+T_{0}-L_{0}\right) \pm \sqrt{\left(K_{\mathrm{d}}+T_{0}-L_{0}\right)^{2}+4 L_{0} K_{\mathrm{d}}}}{2 L_{0}} \tag{S9}
\end{equation*}
$$

with two solutions. Now, let's discuss which solution should be picked to satisfy $0 \leq R \leq 1$. When $\left(K_{\mathrm{d}}+T_{0}-L_{0}\right) \geq 0$, the only solution that satisfies $R \geq 0$ is:

$$
\begin{equation*}
R=\frac{-\left(K_{\mathrm{d}}+T_{0}-L_{0}\right)+\sqrt{\left(K_{\mathrm{d}}+T_{0}-L_{0}\right)^{2}+4 L_{0} K_{\mathrm{d}}}}{2 L_{0}} \tag{S10}
\end{equation*}
$$

since the other solution leads to negative values for $R$.
When $\left(K_{\mathrm{d}}+T_{0}-L_{0}\right)<0$,

$$
\begin{gather*}
-\left(K_{\mathrm{d}}+T_{0}-L_{0}\right)=\left|K_{\mathrm{d}}+T_{0}-L_{0}\right|  \tag{S11}\\
\sqrt{\left(K_{\mathrm{d}}+T_{0}-L_{0}\right)^{2}+4 L_{0} K_{\mathrm{d}}} \geq \sqrt{\left(K_{\mathrm{d}}+T_{0}-L_{0}\right)^{2}}=\left|K_{\mathrm{d}}+T_{0}-L_{0}\right| \text {, since } L_{0} \geq 0 \text { and } K_{\mathrm{d}}>0 \tag{S12}
\end{gather*}
$$

Therefore, the only solution that satisfies $R \geq 0$ is also Eq S10:

$$
R=\frac{-\left(K_{\mathrm{d}}+T_{0}-L_{0}\right)+\sqrt{\left(K_{\mathrm{d}}+T_{0}-L_{0}\right)^{2}+4 L_{0} K_{\mathrm{d}}}}{2 L_{0}}
$$

since the other solution leads to $R<0$.
To sum up, the theoretical dependence of $R$ on $T_{0}$ is Eq S10 or rearranged to:

$$
\begin{equation*}
R=-\frac{K_{\mathrm{d}}+T_{0}-L_{0}}{2 L_{0}}+\sqrt{\left(\frac{K_{\mathrm{d}}+T_{0}-L_{0}}{2 L_{0}}\right)^{2}+\frac{K_{\mathrm{d}}}{L_{0}}} \tag{S13}
\end{equation*}
$$

which is Eq 3 in the main text.

## Note S2: Derivation of the dependence of relative systematic error of $K_{d}\left(\left|\Delta K_{d} / K_{d}\right|\right)$ on $L_{0} / K_{d}$ ratio for correlated (parallel) error sources

Eq 57 shows:

$$
K_{\mathrm{d}}=\frac{T_{0}-L_{0}(1-R)}{(1 / R-1)}
$$

When the error sources of $T_{0}, L_{0}$ and $R$ are strongly correlated, the systematic error of $K_{\mathrm{d}}$ can be calculated with the error propagation rule:

$$
\begin{equation*}
\left|\Delta K_{\mathrm{d}}\right|=\left|\left(\frac{\partial K_{\mathrm{d}}}{\partial T_{0}}\right) \Delta T_{0}\right|+\left|\left(\frac{\partial K_{\mathrm{d}}}{\partial L_{0}}\right) \Delta L_{0}\right|+\left|\left(\frac{\partial K_{\mathrm{d}}}{\partial R}\right) \Delta R\right| \tag{S14}
\end{equation*}
$$

which gives,

$$
\begin{align*}
\left|\Delta K_{\mathrm{d}}\right| & =\left|\frac{R}{1-R} \Delta T_{0}\right|+\left|-R \Delta L_{0}\right|+\left|\left[-L_{0}+\frac{T_{0}}{(1-R)^{2}}\right] \Delta R\right| \\
& =\left|\frac{R}{1-R}\right|\left|\Delta T_{0}\right|+|R|\left|\Delta L_{0}\right|+\left|-L_{0}+\frac{T_{0}}{(1-R)^{2}}\right||\Delta R| \tag{S15}
\end{align*}
$$

Apparently, both $R /(1-R)$ and $R$ are greater than zero since $R \in(0,1)$ when we use Eq S 7 to determine $K_{\mathrm{d}}$. Now, to expand and rearrange Eq S15, let's determine the sign of " $-L_{0}+T_{0} /(1-R)^{2}$ ":
From Eq S7, we obtain,

$$
\begin{equation*}
T_{0}=(1 / R-1) K_{\mathrm{d}}+(1-R) L_{0} \tag{S16}
\end{equation*}
$$

Then,

$$
\begin{align*}
-L_{0}+\frac{T_{0}}{(1-R)^{2}} & =\left[\frac{1 / R-1}{(1-R)^{2}}\right] K_{\mathrm{d}}+\frac{(1-R)}{(1-R)^{2}} L_{0}-L_{0} \\
& =\left[\frac{1-R}{R(1-R)^{2}}\right] K_{\mathrm{d}}+\frac{1}{1-R} L_{0}-L_{0}  \tag{S17}\\
& =\frac{K_{\mathrm{d}}}{R(1-R)}+\frac{R}{1-R} L_{0}
\end{align*}
$$

Eq S 17 clearly shows that the term " $-L_{0}+T_{0} /(1-R)^{2}$ " is greater than zero since both $K_{\mathrm{d}}$ and $L_{0}$ are greater than zero and $R \in(0,1)$. Hence, we rewrite Eq S15 as:

$$
\begin{equation*}
\left|\Delta K_{\mathrm{d}}\right|=\frac{R}{1-R}\left|\Delta T_{0}\right|+R\left|\Delta L_{0}\right|+\left[-L_{0}+\frac{T_{0}}{(1-R)^{2}}\right]|\Delta R| \tag{S18}
\end{equation*}
$$

To simplify the analysis, we only focus on the scenario of determined $R$ equal 0.5 , which is close to the least erroneous case. ${ }^{1}$ Then, the actual $R$ is:

$$
\begin{equation*}
R=0.5-\Delta R,-0.5<\Delta R<0.5 \tag{S19}
\end{equation*}
$$

Insert Eq S19 into Eq S16, we can express $T_{0}$ with $L_{0}$ and $\Delta R$,

$$
\begin{equation*}
T_{0}=\frac{(0.5+\Delta R) K_{\mathrm{d}}}{0.5-\Delta R}+(0.5+\Delta R) L_{0} \tag{S20}
\end{equation*}
$$

Now, divide $K_{\mathrm{d}}$ for both sides of Eq S18 to analyze the relative systematic error of $K_{\mathrm{d}}$ :

$$
\begin{equation*}
\left|\frac{\Delta K_{\mathrm{d}}}{K_{\mathrm{d}}}\right|=\frac{R}{1-R}\left|\frac{\Delta T_{0}}{K_{\mathrm{d}}}\right|+R\left|\frac{\Delta L_{0}}{K_{\mathrm{d}}}\right|+\left[-\frac{L_{0}}{K_{\mathrm{d}}}+\frac{1}{(1-R)^{2}} \frac{T_{0}}{K_{\mathrm{d}}}\right]|\Delta R| \tag{S21}
\end{equation*}
$$

Or rewrite as:

$$
\begin{equation*}
\left|\frac{\Delta K_{\mathrm{d}}}{K_{\mathrm{d}}}\right|=\frac{R}{1-R}\left|\frac{\Delta T_{0}}{T_{0}}\right|\left(\frac{T_{0}}{K_{\mathrm{d}}}\right)+R\left|\frac{\Delta L_{0}}{L_{0}}\right|\left(\frac{L_{0}}{K_{\mathrm{d}}}\right)+\left[-\frac{L_{0}}{K_{\mathrm{d}}}+\frac{1}{(1-R)^{2}} \frac{T_{0}}{K_{\mathrm{d}}}\right]|\Delta R| \tag{S22}
\end{equation*}
$$

By inserting Eq S19 and Eq S20 into Eq S22, we have:

$$
\begin{align*}
\left|\frac{\Delta K_{\mathrm{d}}}{K_{\mathrm{d}}}\right|= & \frac{0.5-\Delta R}{0.5+\Delta R}\left|\frac{\Delta T_{0}}{T_{0}}\right|\left[\frac{0.5+\Delta R}{0.5-\Delta R}+\left(\frac{L_{0}}{K_{\mathrm{d}}}\right)(0.5+\Delta R)\right]+(0.5-\Delta R)\left|\frac{\Delta L_{0}}{L_{0}}\right|\left(\frac{L_{0}}{K_{\mathrm{d}}}\right) \\
& +\left[-\frac{L_{0}}{K_{\mathrm{d}}}+\frac{1}{(0.5+\Delta R)^{2}}\left(\frac{0.5+\Delta R}{0.5-\Delta R}+\left(\frac{L_{0}}{K_{\mathrm{d}}}\right)(0.5+\Delta R)\right)\right]|\Delta R| \tag{S23}
\end{align*}
$$

If we set the absolute value of relative error of $R,|\Delta R / R|=r$ at $R=0.5-\Delta R(\mathrm{Eq} \mathrm{S} 19)$, then:
When $0 \leq \Delta R<0.5$,

$$
\begin{align*}
\Delta R & =R r=(0.5-\Delta R) r \\
& \Rightarrow \Delta R=0.5 r-r \Delta R \\
& \Rightarrow(1+r) \Delta R=0.5 r  \tag{S24}\\
& \Rightarrow \Delta R=\frac{0.5 r}{1+r}
\end{align*}
$$

When $-0.5<\Delta R<0$,

$$
\begin{align*}
\Delta R & =-R r=(\Delta R-0.5) r \\
& \Rightarrow \Delta R=r \Delta R-0.5 r \\
& \Rightarrow(r-1) \Delta R=0.5 r  \tag{S25}\\
& \Rightarrow \Delta R=\frac{0.5 r}{r-1}
\end{align*}
$$

If we also set the absolute values of relative errors of $T_{0}$ and $L_{0}$ as $\left|\Delta T_{0} / T_{0}\right|=t$ and $\left|\Delta L_{0} / L_{0}\right|=l$, respectively, Eq S 23 is rearranged to (we omit some intermediate steps here):

When $0 \leq \Delta R<0.5$,

$$
\begin{align*}
&\left|\frac{\Delta K_{\mathrm{d}}}{K_{\mathrm{d}}}\right|=\frac{0.5 t}{0.5+r}\left[\frac{0.5+r}{0.5}+\frac{0.5+r}{1+r} \frac{L_{0}}{K_{\mathrm{d}}}\right]+\frac{0.5 l}{1+r} \frac{L_{0}}{K_{\mathrm{d}}} \\
&+\left[-\frac{L_{0}}{K_{\mathrm{d}}}+\left(\frac{1+r}{0.5+r}\right)^{2}\left(\frac{0.5+r}{0.5}+\frac{0.5+r}{1+r} \frac{L_{0}}{K_{\mathrm{d}}}\right)\right] \frac{0.5 r}{1+r}  \tag{S26}\\
& \Rightarrow\left|\frac{\Delta K_{\mathrm{d}}}{K_{\mathrm{d}}}\right|=\left[t+\frac{r(1+r)}{0.5+r}\right]+\left[\frac{0.5(t+l-r)}{1+r}+\frac{0.5 r}{0.5+r}\right] \frac{L_{0}}{K_{\mathrm{d}}} \tag{S27}
\end{align*}
$$

When $-0.5<\Delta R<0$,

$$
\begin{align*}
&\left|\frac{\Delta K_{\mathrm{d}}}{K_{\mathrm{d}}}\right|=\frac{0.5 t}{0.5-r}\left[\frac{0.5-r}{0.5}+\frac{r-0.5}{r-1} \frac{L_{0}}{K_{\mathrm{d}}}\right]+\frac{0.5 l}{1-r} \frac{L_{0}}{K_{\mathrm{d}}} \\
&+\left[-\frac{L_{0}}{K_{\mathrm{d}}}+\left(\frac{r-1}{r-0.5}\right)^{2}\left(\frac{0.5-r}{0.5}+\frac{r-0.5}{r-1} \frac{L_{0}}{K_{\mathrm{d}}}\right)\right] \frac{0.5 r}{1-r}  \tag{S28}\\
& \Rightarrow\left|\frac{\Delta K_{\mathrm{d}}}{K_{\mathrm{d}}}\right|=\left[t+\frac{r(1-r)}{0.5-r}\right]+\left[\frac{0.5(t+l-r)}{1-r}+\frac{0.5 r}{0.5-r}\right] \frac{L_{0}}{K_{\mathrm{d}}} \tag{S29}
\end{align*}
$$

Or we can write Eq S27 and Eq S29 as:

$$
\begin{align*}
& \left|\frac{\Delta K_{\mathrm{d}}}{K_{\mathrm{d}}}\right|=a+b \frac{L_{0}}{K_{\mathrm{d}}}, \\
& a=t+\frac{r(1+r)}{0.5+r}, b=\frac{0.5(t+l-r)}{1+r}+\frac{0.5 r}{0.5+r} \text { when } 0 \leq \Delta R<0.5  \tag{S30}\\
& a=t+\frac{r(1-r)}{0.5-r}, b=\frac{0.5(t+l-r)}{1-r}+\frac{0.5 r}{0.5-r} \text { when }-0.5<\Delta R<0
\end{align*}
$$

which is Eq 6 in the main text.

Note S3: Derivation of the theoretical dependence of determined $K_{d}$ ( $K_{d, d e t}$ ) on $L_{0}$ for correlated (parallel) error sources
The definition of $\Delta K_{\mathrm{d}}$ is (with $K_{\mathrm{d}, \mathrm{det}}$ representing the determined $K_{\mathrm{d}}$ value),

$$
\begin{equation*}
\Delta K_{\mathrm{d}}=K_{\mathrm{d}, \mathrm{det}}-K_{\mathrm{d}} \tag{S31}
\end{equation*}
$$

By inserting Eq S31 into Eq S30, we get:

$$
\begin{equation*}
\left|\frac{K_{\mathrm{d}, \mathrm{det}}-K_{\mathrm{d}}}{K_{\mathrm{d}}}\right|=a+b \frac{L_{0}}{K_{\mathrm{d}}} \tag{S32}
\end{equation*}
$$

By multiplying $K_{\mathrm{d}}$ on both sides of Eq S32, we have:

$$
\begin{equation*}
\left|K_{\mathrm{d}, \mathrm{det}}-K_{\mathrm{d}}\right|=a K_{\mathrm{d}}+b L_{0} \tag{S33}
\end{equation*}
$$

When $\Delta K_{\mathrm{d}} \geq 0$,

$$
\begin{align*}
& K_{\mathrm{d}, \operatorname{det}}-K_{\mathrm{d}}=a K_{\mathrm{d}}+b L_{0} \\
& \Rightarrow K_{\mathrm{d}, \mathrm{det}}=(1+a) K_{\mathrm{d}}+b L_{0} \tag{S34}
\end{align*}
$$

When $\Delta K_{\mathrm{d}}<0$,

$$
\begin{align*}
& K_{\mathrm{d}}-K_{\mathrm{d}, \mathrm{det}}=a K_{\mathrm{d}}+b L_{0} \\
& \Rightarrow K_{\mathrm{d}, \mathrm{det}}=(1-a) K_{\mathrm{d}}-b L_{0} \tag{S35}
\end{align*}
$$

Or we can combine Eqs S34 and S35 to be:

$$
\begin{align*}
K_{\mathrm{d}, \mathrm{det}}=c+d L_{0}, c & =(1+a) K_{\mathrm{d}}, d=b \text { when } \Delta K_{\mathrm{d}} \geq 0 \\
c & =(1-a) K_{\mathrm{d}}, d=-b \text { when } \Delta K_{\mathrm{d}}<0 \tag{S36}
\end{align*}
$$

Eq S36 (Eq 7 in the main text) suggests that the dependence of $K_{\mathrm{d}, \mathrm{det}}$ on $L_{0}$ is linear in the whole range of $L_{0}$.

Note S4: Derivation of the dependence of relative systematic error of $K_{d}\left(\left|\Delta K_{d} / K_{d}\right|\right)$ on $L_{0} / K_{d}$ ratio for uncorrelated (orthogonal) error sources
When the error sources of $T_{0}, L_{0}$ and $R$ are uncorrelated, the systematic error of $K_{\mathrm{d}}$ can be calculated with the error propagation rule:

$$
\begin{equation*}
\left|\Delta K_{\mathrm{d}}\right|=\sqrt{\left(\frac{\partial K_{\mathrm{d}}}{\partial T_{0}}\right)^{2} \Delta T_{0}^{2}+\left(\frac{\partial K_{\mathrm{d}}}{\partial L_{0}}\right)^{2} \Delta L_{0}^{2}+\left(\frac{\partial K_{\mathrm{d}}}{\partial R}\right)^{2} \Delta R^{2}} \tag{S37}
\end{equation*}
$$

By applying Eq S37 to Eq S7 and square both sides, we have,

$$
\begin{equation*}
\Delta K_{\mathrm{d}}^{2}=\left(\frac{R}{1-R}\right)^{2} \Delta T_{0}^{2}+R^{2} \Delta L_{0}^{2}+\left[-L_{0}+\frac{T_{0}}{(1-R)^{2}}\right]^{2} \Delta R^{2} \tag{S38}
\end{equation*}
$$

By replacing $T_{0}$ with Eq S16, we obtain:

$$
\begin{equation*}
\Delta K_{\mathrm{d}}^{2}=\left(\frac{R}{1-R}\right)^{2} \Delta T_{0}^{2}+R^{2} \Delta L_{0}^{2}+\left[\left(\frac{1}{1-R}\right)\left(\frac{K_{\mathrm{d}}}{R}+R L_{0}\right)\right]^{2} \Delta R^{2} \tag{S39}
\end{equation*}
$$

Now, divide $K_{\mathrm{d}}{ }^{2}$ for both sides of Eq S39 to analyze the relative systematic error of $K_{\mathrm{d}}$ :

$$
\begin{align*}
\left(\frac{\Delta K_{\mathrm{d}}}{K_{\mathrm{d}}}\right)^{2} & =\left(\frac{R}{1-R}\right)^{2}\left(\frac{\Delta T_{0}}{K_{\mathrm{d}}}\right)^{2}+R^{2}\left(\frac{\Delta L_{0}}{K_{\mathrm{d}}}\right)^{2}+\left[\left(\frac{1}{1-R}\right)\left(\frac{1}{R}+R \frac{L_{0}}{K_{\mathrm{d}}}\right)\right]^{2} \Delta R^{2} \\
& =\left(\frac{R}{1-R}\right)^{2}\left(\frac{\Delta T_{0}}{T_{0}}\right)^{2}\left(\frac{T_{0}}{K_{\mathrm{d}}}\right)^{2}+R^{2}\left(\frac{\Delta L_{0}}{L_{0}}\right)^{2}\left(\frac{L_{0}}{K_{\mathrm{d}}}\right)^{2}+\left[\left(\frac{1}{1-R}\right)\left(\frac{1}{R}+R \frac{L_{0}}{K_{\mathrm{d}}}\right)\right]^{2} \Delta R^{2} \\
& =\left(\frac{R}{1-R}\right)^{2}\left(\frac{\Delta T_{0}}{T_{0}}\right)^{2}\left[\frac{(1 / R-1) K_{\mathrm{d}}+(1-R) L_{0}}{K_{\mathrm{d}}}\right]^{2}+R^{2}\left(\frac{\Delta L_{0}}{L_{0}}\right)^{2}\left(\frac{L_{0}}{K_{\mathrm{d}}}\right)^{2}+\left[\left(\frac{1}{1-R}\right)\left(\frac{1}{R}+R \frac{L_{0}}{K_{\mathrm{d}}}\right)\right]^{2} \Delta R^{2} \\
& =\left(1+R \frac{L_{0}}{K_{\mathrm{d}}}\right)^{2}\left(\frac{\Delta T_{0}}{T_{0}}\right)^{2}+R^{2}\left(\frac{\Delta L_{0}}{L_{0}}\right)^{2}\left(\frac{L_{0}}{K_{\mathrm{d}}}\right)^{2}+\left[\left(\frac{1}{1-R}\right)\left(\frac{1}{R}+R \frac{L_{0}}{K_{\mathrm{d}}}\right)\right]^{2} \Delta R^{2} \tag{S40}
\end{align*}
$$

After setting $\left|\Delta T_{0} / T_{0}\right|=t$ and $\left|\Delta L_{0} / L_{0}\right|=l$, Eq S40 can be expanded and rearranged to be:

$$
\begin{equation*}
\left(\frac{\Delta K_{\mathrm{d}}}{K_{\mathrm{d}}}\right)^{2}=\left[t^{2}+\left(\frac{\Delta R}{R(1-R)}\right)^{2}\right]+\left[2 R t^{2}+2\left(\frac{\Delta R}{1-R}\right)^{2}\right] \frac{L_{0}}{K_{\mathrm{d}}}+\left[R^{2} t^{2}+R^{2} l^{2}+\left(\frac{R \Delta R}{1-R}\right)^{2}\right]\left(\frac{L_{0}}{K_{\mathrm{d}}}\right)^{2} \tag{S41}
\end{equation*}
$$

When $0 \leq \Delta R<0.5$, by inserting Eqs S19 and S24 into Eq S41, we obtain:

$$
\begin{equation*}
\left(\frac{\Delta K_{\mathrm{d}}}{K_{\mathrm{d}}}\right)^{2}=\left[t^{2}+\left(\frac{r(1+r)}{0.5+r}\right)^{2}\right]+\left[\frac{t^{2}}{1+r}+2\left(\frac{0.5 r}{0.5+r}\right)^{2}\right] \frac{L_{0}}{K_{\mathrm{d}}}+\left[\left(\frac{0.5}{1+r}\right)^{2}\left(t^{2}+l^{2}+\left(\frac{0.5 r}{0.5+r}\right)^{2}\right)\right]\left(\frac{L_{0}}{K_{\mathrm{d}}}\right)^{2} \tag{S42}
\end{equation*}
$$

or

$$
\begin{equation*}
\left|\frac{\Delta K_{\mathrm{d}}}{K_{\mathrm{d}}}\right|=\sqrt{\left[t^{2}+\left(\frac{r(1+r)}{0.5+r}\right)^{2}\right]+\left[\frac{t^{2}}{1+r}+2\left(\frac{0.5 r}{0.5+r}\right)^{2}\right] \frac{L_{0}}{K_{\mathrm{d}}}+\left[\left(\frac{0.5}{1+r}\right)^{2}\left(t^{2}+l^{2}+\left(\frac{0.5 r}{0.5+r}\right)^{2}\right)\right]\left(\frac{L_{0}}{K_{\mathrm{d}}}\right)^{2}} \tag{S43}
\end{equation*}
$$

When $-0.5<\Delta R<0$, by inserting Eqs S 19 and S25 into Eq S41, we obtain:

$$
\begin{equation*}
\left(\frac{\Delta K_{\mathrm{d}}}{K_{\mathrm{d}}}\right)^{2}=\left[t^{2}+\left(\frac{r(1-r)}{r-0.5}\right)^{2}\right]+\left(\frac{t^{2}}{1-r}+2\left(\frac{0.5 r}{r-0.5}\right)^{2}\right) \frac{L_{0}}{K_{\mathrm{d}}}+\left[\left(\frac{0.5}{1-r}\right)^{2}\left(t^{2}+l^{2}+\left(\frac{0.5 r}{r-0.5}\right)^{2}\right)\right]\left(\frac{L_{0}}{K_{\mathrm{d}}}\right)^{2} \tag{S44}
\end{equation*}
$$

or

$$
\begin{equation*}
\left|\frac{\Delta K_{\mathrm{d}}}{K_{\mathrm{d}}}\right|=\sqrt{\left[t^{2}+\left(\frac{r(1-r)}{r-0.5}\right)^{2}\right]+\left(\frac{t^{2}}{1-r}+2\left(\frac{0.5 r}{r-0.5}\right)^{2}\right) \frac{L_{0}}{K_{\mathrm{d}}}+\left[\left(\frac{0.5}{1-r}\right)^{2}\left(t^{2}+l^{2}+\left(\frac{0.5 r}{r-0.5}\right)^{2}\right)\right]\left(\frac{L_{0}}{K_{\mathrm{d}}}\right)^{2}} \tag{S45}
\end{equation*}
$$

We can combine Eqs S43 and S45 as:

$$
\begin{align*}
& \left|\frac{\Delta K_{\mathrm{d}}}{K_{\mathrm{d}}}\right|=\sqrt{\alpha^{2}+\lambda \frac{L_{0}}{K_{\mathrm{d}}}+\beta^{2}\left(\frac{L_{0}}{K_{\mathrm{d}}}\right)^{2},} \\
& \alpha=\sqrt{t^{2}+\left(\frac{r+r^{2}}{0.5+r}\right)^{2}}, \beta=\frac{0.5}{1+r} \sqrt{t^{2}+l^{2}+\left(\frac{0.5 r}{0.5+r}\right)^{2}}, \lambda=\frac{t^{2}}{1+r}+2\left(\frac{0.5 r}{0.5+r}\right)^{2} \text { when } 0 \leq \Delta R<0.5  \tag{S46}\\
& \alpha=\sqrt{t^{2}+\left(\frac{r-r^{2}}{r-0.5}\right)^{2}}, \beta=\frac{0.5}{1-r} \sqrt{t^{2}+l^{2}+\left(\frac{0.5 r}{r-0.5}\right)^{2}}, \lambda=\frac{t^{2}}{1-r}+2\left(\frac{0.5 r}{r-0.5}\right)^{2} \text { when }-0.5<\Delta R<0
\end{align*}
$$

## Note S5: Derivation of the theoretical dependence of determined $K_{d}$ ( $K_{\mathrm{d}, \mathrm{det}}$ ) on $L_{0}$ for uncorrelated (orthogonal) error sources

By inserting Eq S31 into Eq S46, we get:

$$
\begin{equation*}
\left|\frac{K_{\mathrm{d}, \mathrm{det}}-K_{\mathrm{d}}}{K_{\mathrm{d}}}\right|=\sqrt{\alpha^{2}+\lambda \frac{L_{0}}{K_{\mathrm{d}}}+\beta^{2}\left(\frac{L_{0}}{K_{\mathrm{d}}}\right)^{2}} \tag{S47}
\end{equation*}
$$

By multiplying $K_{\mathrm{d}}$ on both sides of Eq S47, we have:

$$
\begin{equation*}
\left|K_{\mathrm{d}, \mathrm{det}}-K_{\mathrm{d}}\right|=\sqrt{\alpha^{2} K_{\mathrm{d}}^{2}+\lambda K_{\mathrm{d}} L_{0}+\beta^{2} L_{0}^{2}} \tag{S48}
\end{equation*}
$$

When $\Delta K_{\mathrm{d}} \geq 0$,

$$
\begin{align*}
& K_{\mathrm{d}, \mathrm{det}}-K_{\mathrm{d}}=\sqrt{\alpha^{2} K_{\mathrm{d}}^{2}+\lambda K_{\mathrm{d}} L_{0}+\beta^{2} L_{0}^{2}} \\
& \Rightarrow K_{\mathrm{d}, \mathrm{det}}=K_{\mathrm{d}}+\sqrt{\alpha^{2} K_{\mathrm{d}}^{2}+\lambda K_{\mathrm{d}} L_{0}+\beta^{2} L_{0}^{2}} \tag{S49}
\end{align*}
$$

From Eq S49, we can infer that, at low $L_{0}$ range, $K_{\mathrm{d}, \mathrm{det}}$ approaches a constant:

$$
\begin{equation*}
K_{\mathrm{d}, \mathrm{det}}=K_{\mathrm{d}}+\alpha K_{\mathrm{d}}=(1+\alpha) K_{\mathrm{d}} \tag{S50}
\end{equation*}
$$

Or we can write,

$$
\begin{equation*}
K_{\mathrm{d}, \mathrm{det}}=\gamma, \gamma=(1+\alpha) K_{\mathrm{d}} \tag{S51}
\end{equation*}
$$

At high $L_{0}$ range, the dependency of $K_{\mathrm{d}, \text { det }}$ on $L_{0}$ approaches a linear equation:

$$
\begin{equation*}
K_{\mathrm{d}, \mathrm{det}}=\beta L_{0} \tag{S52}
\end{equation*}
$$

When $\Delta K_{\mathrm{d}}<0$,

$$
\begin{align*}
& K_{\mathrm{d}}-K_{\mathrm{d}, \mathrm{det}}=\sqrt{\alpha^{2} K_{\mathrm{d}}^{2}+\lambda K_{\mathrm{d}} L_{0}+\beta^{2} L_{0}^{2}} \\
& \Rightarrow K_{\mathrm{d}, \mathrm{det}}=K_{\mathrm{d}}-\sqrt{\alpha^{2} K_{\mathrm{d}}^{2}+\lambda K_{\mathrm{d}} L_{0}+\beta^{2} L_{0}^{2}} \tag{S53}
\end{align*}
$$

From Eq S53, we can infer that, at low $L_{0}$ range, $K_{\mathrm{d}, \mathrm{det}}$ approaches a constant:

$$
\begin{equation*}
K_{\mathrm{d}, \mathrm{det}}=K_{\mathrm{d}}-\alpha K_{\mathrm{d}}=(1-\alpha) K_{\mathrm{d}} \tag{S54}
\end{equation*}
$$

Or we use the same parameter $\gamma$ to express Eq S54 as,

$$
\begin{equation*}
K_{\mathrm{d}, \mathrm{det}}=\gamma, \gamma=(1-\alpha) K_{\mathrm{d}} \tag{S55}
\end{equation*}
$$

The dependency of $K_{\mathrm{d}, \mathrm{det}}$ on $L_{0}$ at high $L_{0}$ range is unavailable because $K_{\mathrm{d}, \mathrm{det}}$ is required to be greater than zero.

Figure S1: General trends in dependencies of $\log \left(\left|\Delta K_{d} / K_{d}\right|\right)$ on $\log \left(L_{0} / K_{d}\right)$ and $\log \left(K_{d, d e t}\right)$ on $\log \left(L_{0}\right)$
Panels (a) and (c) show the general trends in dependency of $\log \left(\left|\Delta K_{\mathrm{d}} / K_{\mathrm{d}}\right|\right)$ on $\log \left(L_{0} / K_{\mathrm{d}}\right)$; panels (b) and (d) show the general trends in dependency of $\log \left(K_{\mathrm{d}, \mathrm{det}}\right)$ on $\log \left(L_{0}\right)$. In all panels, reasonable relative errors of $L_{0}, T_{0}$ and $R$ were used: $\left|\Delta T_{0} / T_{0}\right|=\left|\Delta L_{0} / L_{0}\right|=0.05$ and $\Delta R / R=-0.02$. In panels (b) and (d), we set $K_{\mathrm{d}}=1$ (unitless) and $\Delta K_{\mathrm{d}}<0$. With these conditions, the parameters $a, b, c$ and $d$ were calculated to be 0.011 , $0.040,0.99$ and -0.040 , respectively; $\alpha, \beta$ and $\gamma$ were calculated to be $0.064,0.037$ and 0.94 , respectively.


For uncorrelated error sources of $T_{0}, L_{0}$ and $R$


With Eqs S30, S36, S46 and S51, we produced representative graphs to show the dependence of $\left|\Delta K_{\mathrm{d}} / K_{\mathrm{d}}\right|$ on $L_{0} / K_{\mathrm{d}}$ and the dependence of $K_{\mathrm{d}, \mathrm{det}}$ on $L_{0}$ in double-log scale. The graphs corresponding to $0 \leq \Delta R<0.5$ and $\Delta K_{\mathrm{d}} \geq 0$ are shown in Figure 2 in the main text, and the graphs corresponding to $-0.5<\Delta R<0$ and $\Delta K_{\mathrm{d}}$ $<0$ are shown in Figure S1. Figures 2a, 2c (in main text) and Figures S1a, S1c show the same general trends in dependency of $\log \left(\left|\Delta K_{\mathrm{d}} / K_{\mathrm{d}}\right|\right)$ on $\log \left(L_{0} / K_{\mathrm{d}}\right)$. Although Figures S1b and S1d cannot show the part at high $L_{0}$ range (where $K_{\mathrm{d}, \text { det }}$ equals physically unfeasible negative values), the results in Figure S1b and S1d suggest a dependency of $K_{\mathrm{d}, \mathrm{det}}$ on $L_{0}$ that is similar to the trend shown in Figures 2b and 2d (in main text):
$K_{\mathrm{d}, \mathrm{det}}$ is insensitive to the change of $L_{0}$ when $L_{0}$ is at low concentrations, while the sensitivity of $K_{\mathrm{d}, \mathrm{det}}$ to $L_{0}$ increases with the increase of $L_{0}$.

## Note S6: Derivation of the relationship between signal to noise ratio $(S / N)$ and the limit of quantitation (LOQ)

The limit of quantitation (LOQ) of an instrument is defined as the analyte concentration (ligand concentration in our case) for which the signal to noise ratio $(S / N)$ is equal to a certain value $X \gg 1$ (e.g., $X=10$ ) which guarantees that the noise does not affect quantitation significantly, which can be expressed as:

$$
\begin{equation*}
\frac{S_{\mathrm{LOQ}}}{N}=X \Rightarrow S_{\mathrm{LOQ}}=X N \tag{S56}
\end{equation*}
$$

where $S_{\text {LOQ }}$ is the signal measured at LOQ. For one instrument and a specific ligand, since the measured signal $S$ is proportional to the ligand concentration, we can assume:

$$
\begin{equation*}
S_{\mathrm{LOQ}}=k \cdot \mathrm{LOQ} \Rightarrow k=\frac{S_{\mathrm{LOQ}}}{\mathrm{LOQ}}=\frac{X N}{\mathrm{LOQ}} \tag{S57}
\end{equation*}
$$

where factor $k$ is the ratio of measured signal to ligand concentration. Now, if the noise $N$ is independent of the signal $S$, the signal to noise ratio for any total ligand concentration $L_{0}$ can be calculated as:

$$
\begin{align*}
& \frac{S}{N}=\frac{k \cdot L_{0}}{N}=\frac{X N}{\mathrm{LOQ}} \cdot \frac{L_{0}}{N} \\
& \Rightarrow \frac{S}{N}=X \frac{L_{0}}{\mathrm{LOQ}} \tag{S58}
\end{align*}
$$

The minimum acceptable ligand concentration which satisfies Eq S 58 is $L_{0, \text { min }}=\mathrm{LOQ}$.

## Section S2: Experiments

Figure S2: Determination of limit of quantitation (LOQ)
a) Electropherograms of the measurements for samples of MutS-binding aptamer with concentrations of $0.1,1,10,20,50$, and 100 pM . The instrumental setup and experimental procedure are described in "Materials and Methods" section in the main text. b) An example ([aptamer] $]_{0}=20 \mathrm{pM}$ ) of determining signal and noise for an electropherogram: the intensity difference between the highest noise peak and the lowest noise peak was determined as the noise height $h$, which equals 2 times noise $N$; the midline of the noise peaks was determined as the baseline; the intensity difference between the highest signal of the signal peak and the baseline was determined as signal height $H$, which equal the signal $S$. c) Determining LOQ based on the linear plotting of "signal versus [aptamer]o".


To determine the LOQ of our laser-induced fluorescence (LIF) detection system for the fluorescently labeled MutS-binding aptamer, we measured the aptamer samples with concentrations ( $L_{0}$ or [aptamer $]_{0}$ ) of $0.1,1,10,20,50$, and 100 pM . In the measurements, the instrumental setup and experimental procedure were identical to that used in the NECEEM (nonequilibrium capillary electrophoresis of equilibrium mixtures) experiments as explained in "Materials and Methods" section in the main text. The obtained electropherograms are shown in Figure S2a. Based on the electropherograms, the signals of aptamer at each concentration were determined to be $0.0020,0.0031,0.022,0.039,0.072$, and 0.20 RFU , respectively. By analyzing all six electropherograms, the average noise was determined to be 0.0025 RFU. One example of determining signal ( $S$ ), noise ( $N$ ), and $S / N$ is demonstrated in Figure S2b. Then, the results of "signal versus
[aptamer] $]_{0}$ " were plotted (Figure S2c), and a linear fitting was applied to the data points. Conventionally, the analyte concentration corresponding to signal to noise ratio $(S / N)$ of 10 (i.e., $X=10$ in Eq S56) is defined as LOQ. ${ }^{2-4}$ In this work, considering the signal of unbound aptamer decreases with the increase of protein concentration, we chose the [aptamer] $]_{0}$ corresponding to $S / N=15$ as LOQ. For different applications, one can choose different values for $X$ in Eq S 57 to achieve a desired accuracy and precision of measured signals at LOQ. By inserting the signal equal 15 times of average noise plus the intercept of the fitting line (for correcting the signal offset caused by background signal) into Figure S2c, LOQ for this work was determined to be 20 pM (i.e., 0.02 nM ).

## Figure S3: Electropherograms of the NECEEM experiments

Panels $\mathbf{a}-\mathbf{f}$ show the electropherograms of the NECEEM experiments with $L_{0}$ ([aptamer $]_{0}$ ) of $0.02,0.05$, $0.1,0.5,2$, and 10 nM , respectively. The instrumental setup and experimental procedure are explained in "Materials and Methods" section in the main text.



Based on the electropherograms shown in Figure S3, the areas of the peaks for unbound aptamer and complex were measured with NECEEM Area Analysis Program (NAAP) that can be downloaded from https://www.yorku.ca/skrylov/resources.html. In each set of NECEEM experiments, since the corresponding highest MutS concentration was much greater than the expected $K_{\mathrm{d}}$ value range of $\leq 0.1 \mathrm{nM}$, we presumed that the highest MutS concentration in each set of NECEEM experiments saturated the binding to the aptamer, and the remaining unbound aptamer was the impurity (i.e., non-binders) in the aptamer sample. Therefore, from each unbound aptamer area, we subtracted the unbound aptamer area in corresponding "aptamer + highest MutS concentration" electropherogram to obtain the actual unbound aptamer area. Then, the determined areas were corrected by dividing them by their migration times. ${ }^{1}$ Based on the corrected areas, the actual fractions of unbound aptamer ( $R$ values) at different $T_{0}$ ( $[\mathrm{MutS}]_{0}$ ) were calculated, and the results are shown in Table S1. The binding isotherms for the NECEEM experiments are shown in Figure 3a in the main text.

Table S1: Results of $R$ values at different $T_{0}\left([M u t S]_{0}\right)$ for the NECEEM experiments with different $L_{0}$ ([aptamer]0)

| $\boldsymbol{T}_{\mathbf{0}}$ <br> $(\mathbf{n M})$ | $\boldsymbol{R}$ <br> $\left(\boldsymbol{L}_{\mathbf{0}}=\mathbf{0 . 0 2} \mathbf{n M}\right)$ | $\boldsymbol{R}$ <br> $\left(\boldsymbol{L}_{\mathbf{0}}=\mathbf{0 . 0 5} \mathbf{n M}\right)$ | $\boldsymbol{R}$ <br> $\left(\boldsymbol{L}_{\mathbf{0}}=\mathbf{0 . 1} \mathbf{~ n M}\right)$ | $\boldsymbol{R}$ <br> $\left(\boldsymbol{L}_{\mathbf{0}}=\mathbf{0 . 5} \mathbf{n M}\right)$ | $\boldsymbol{R}$ <br> $\left(\boldsymbol{L}_{\mathbf{0}}=\mathbf{2} \mathbf{n M}\right)$ | $\boldsymbol{R}$ <br> $\left(\boldsymbol{L}_{\mathbf{0}}=\mathbf{1 0} \mathbf{n M}\right)$ |
| :--- | :---: | :---: | :---: | :---: | :---: | :---: |
| $\mathbf{0}$ | 1.0 | 1.0 | 1.0 | 1.0 | 1.0 | 1.0 |
| $\mathbf{0 . 0 0 4}$ | 0.96 | N/A | N/A | N/A | N/A | N/A |
| $\mathbf{0 . 0 2}$ | 0.81 | 0.86 | 1.0 | N/A | N/A | N/A |
| $\mathbf{0 . 1}$ | 0.42 | 0.45 | 0.89 | 0.94 | N/A | N/A |
| $\mathbf{0 . 5}$ | 0.17 | 0.11 | 0.52 | 0.73 | 0.92 | 0.99 |
| $\mathbf{2 . 5}$ | 0.0 | 0.066 | 0.073 | 0.15 | 0.65 | 0.97 |
| $\mathbf{1 2 . 5}$ | N/A | 0.0 | 0.0 | 0.053 | 0.10 | 0.84 |
| $\mathbf{6 2 . 5}$ | N/A | N/A | N/A | 0.0 | 0.041 | 0.098 |
| $\mathbf{3 1 2 . 5}$ | N/A | N/A | N/A | N/A | 0.0 | 0.0 |

## References

(1) Kanoatov, M.; Galievsky, V. A.; Krylova, S. M.; Cherney, L. T.; Jankowski, H. K.; Krylov, S. N. Using Non-Equilibrium Capillary Electrophoresis of Equilibrium Mixtures (NECEEM) for Simultaneous Determination of Concentration and Equilibrium Constant. Anal. Chem. 2015, 87, 3099-3106.
(2) Schejbal, J.; Slezáčková, L.; Řemínek, R.; Glatz, Z. A Capillary Electrophoresis-Mass Spectrometry based Method for the Screening of $\beta$-Secretase Inhibitors as Potential Alzheimer's Disease Therapeutics. J. Chromatogr. A 2017, 1487, 235-241.
(3) Galievsky, V. A.; Stasheuski, A. S.; Krylov, S. N. "Getting the Best Sensitivity from On-Capillary Fluorescence Detection in Capillary Electrophoresis" - A Tutorial. Anal. Chem. Acta 2016, 935, 5881.
(4) Nicolaou, I. N.; Kapnissi-Christodoulou, C. P. Development of A Capillary Electrophoresis-Mass Spectrometry Method for the Determination of Rivastigmine in Human Plasma - Optimization of the Limits of Detection and Quantitation. Electrophor. 2012, 33, 644-652.


[^0]:    Received: August 9, 2023
    Accepted: September 27, 2023
    Published: October 13, 2023

