

LECTURES

I. GEOLOGICAL HISTORY OF PHOTOSYNTHESIS

- A. Macrofossil Evidence
 - 1. Stromatolites – structural and functional stratification of oxygenic and anoxygenic photosynthesizers, and aerobic and anaerobic heterotrophs
- B. ^{12}C / ^{13}C Discrimination
 - 1. Indirect evidence for carbon dioxide fixation due to ^{12}C preference
- C. Oxygen production and geochemistry
 - 1. Marine Fe(II) oxidation to Fe(III) as a major sink for early oxygen production

The range of evidence creates a cohesive picture of atmospheric oxygen, created from oxygenic photosynthesis, appearing in concert with life forms more complex than the prokaryotic groups.

The geological history of photosynthesis provides students with an overview of the profound impact of photosynthesis as terra-former of the world. An understanding of this vast scope of biological imprint fulfills one of the Learning Objectives of the course.

II. PHOTOSYNTHETIC DIVERSITY

- A. Introduction to Anoxygenic Prokaryotic Groups
 - 1. Bacteriochlorophylls
 - 2. H_2S and H_2 as electron donors
 - 3. Unique carbon fixation mechanisms including reverse Krebs cycle and the Hydroxypropionate pathway

The emphasis on prokaryotic diversity highlighted the major primordial contributors to photosynthesis (anoxygenic). In the context of bioengineering, these groups may be useful, for biomimetics and possibly as engineered microbes.

The ‘ancien regime’ of photosynthesis involved a multitude of biochemical pathways only rarely observed in present time. These underpin the geological time span of terra-forming, thereby fulfilling one of the Learning Objectives of the course.

III. LIGHT

- A. The Physics of Light
 - 1. Dipole origins
 - 2. Wave and particle: The photoelectric effect and the energy of a photon

The physical properties of light were introduced in detail, including an introduction to Maxwell’s field equations and a detailed presentation of the photoelectric effect and its use to determine the energy of absorption events.

- B. Light and Photosynthesis
 - 1. Absorption by chlorophyll and exciton fates
 - a. fluorescence, exciton transfer, radiationless loss, triplet formation (phosphorescence) and photochemistry
 - b. photooxidation

The properties of light are closely intertwined with multiple events during the light reactions of photosynthesis, including photochemistry, the desired outcome of an absorption event. Absorption events and outcomes were presented in the context of useful and useless outcomes, including the photooxidation damage caused by long-lived triplet states.

The physics of light (Planck’s Black Body, the Photoelectrical Effect, the fates of excitons) are introduced to provide students with the necessary foundations for understanding how photosynthesis uses light energy, fulfilling the Learning Objective of providing students with the basic knowledge needed to understand how photosynthesis works.

III. PIGMENTS OF PHOTOSYNTHESIS

- A. Carotenoids
 - 1. Synthesis and role in protection from photooxidation
- B. Bilins
 - 1. Introduction to the pyrrole motif of linear and cyclic tetrapyrroles
 - 2. Phylogenetic uniqueness of bilin distribution
- C. Chlorophylls
 - 1. Introduction to the diversity of cyclic tetrapyrroles
 - a. hemes (Fe insertion)
 - b. vitamins and vanadyl porphyrins
 - c. chlorophylls (Mg insertion)
 - 2. Synthesis
 - a. delta-aminolevulinic acid, the first committed precursor
 - i. succinyl CoA and glycine pathway
 - ii. glutamyl-tRNA mediated pathway
 - b. deamination, decarboxylation and unsaturation to form the porphyrin foundation of the chlorophyll molecule

The central role of tetrapyrroles in multiple biochemical processes in organisms was emphasized, with reference to the evolution of diverse synthesis pathways that occur in tandem in photosynthetic eukaryotic cells.

Students are introduced to the diverse mechanisms of trapping light energy that have evolved through geological eons, reinforcing the Learning Objective of illuminating the geological span of evolution and terra-forming.

IV. LIGHT HARVESTING

- A. Emerson and Arnold's Classic Experiments on Light and O₂ production
 - 1. Dark interval required to complete O₂ production
 - 2. Chlorophyll antenna
 - a. light-saturating conditions
 - b. chlorophyll molecules per oxygen molecule produced
- B. Light-Harvesting Complexes
 - 1. SDS-PAGE characterization
 - 2. X-ray crystallographic structures
 - 3. Regulation of the architecture of photosynthesis
 - a. PQH₂/PQ ratio and granal stacking to effect modulation of electron fate through cyclic (ATP synthesis) and non-cyclic (NADPH production) photosynthesis

The classic basis for light harvesting complexes was presented in detail, including reading of the original papers by Emerson and Arnold. This was followed by an in depth description of the composition of light-harvesting complexes from major phylogenetic lineages and finally their role in regulation of the light reactions of photosynthesis in higher plants.

Our understanding of the mechanisms of photosynthesis relies upon diverse biochemical characterizations, providing students with an understanding of how photosynthesis was discovered, fulfilling a fundamental Learning Objective of the course, reinforced in the Laboratory Exercises.

V. REACTION CENTERS

- A. Evidence for Two Reaction Centers in Oxygenic Photosynthesis
 - 1. Red drop and red light enhancement
 - 2. Cytochrome oxidation
- B. The Structure of the *Rhodospseudomonas* (*Rhodobacter*) Reaction Center
 - 1. X-ray crystallography (Michel and Deisenhofer)
- C. The Structure of the PS II Reaction Center
 - 1. X-ray crystallography
 - 2. Water-splitting: Structure, kinetics and mechanism

The historical evolution of our understanding of the reaction center was presented, with emphasis on how structural elucidations have informed the present 'state-of-art' regarding the physical mechanisms of photochemistry and the charge separation after the photochemical event.

The in depth presentation of biochemical characterizations is continued, emphasizing recent research advances to provide students with an understanding of current research in photosynthesis, fulfilling a fundamental Learning Objective of the course, reinforced in the Laboratory Exercises.

VI. NADPH AND ATP PRODUCTION

- A. e^- and H^+ Transfer Mechanisms
 - 1. Mediators of redox reactions
 - a. NADPH: structure and function
 - b. flavins: structure and function
 - c. ubiquinone (plastoquinone): structure and function
 - d. iron-sulfur centers: structure and function
 - e. hemes
 - 2. H^+ shuttling
 - a. cytochrome b_6f complex
 - b. plastoquinone

Emphasis was placed on the mediators of redox reactions, rather than the linear sequences of redox transfers that occur in the electron transport chain, since these vary depending upon the organism (principally between anoxygenic and oxygenic organisms).

- B. Chemiosmotic Theory and the Coupling Factor
 - 1. ATP bioenergetics
 - 2. ATP synthetase
 - a. kinetic mechanism
 - b. mechanical mechanism

The relation between 'osmotic' gradients and chemical synthesis were presented, as well as the kinetic steps in ATP synthesis and the concept of a mechanical (rotatory) linkage between the proton motive force and ATP production.

The in depth presentation of the biochemistry of photosynthesis continues. An in depth knowledge of the complexity of the photosynthetic process is crucial for fundamental understanding, and the ability to bioengineer photosynthesis in support of continued human survival, fulfilling a fundamental Learning Objective of the course, reinforced in the Laboratory Exercises, in which students are taught how to measure electron flow and ATP synthesis.

VII. DARK REACTIONS

- A. Carbon Dioxide Fixation by RuBisCO
 - 1. Carboxylase reaction
 - 2. Structure of the enzyme, and its regulation

Details of the mechanism of carboxylation within the active site, including the mechanism of enzyme

activation by carbamylation were presented in detail.

- B. Reductive Pentose Phosphate Pathway (the Calvin Cycle)
 - 1. Enzymatic reactions and the central role of 3-phosphoglyceraldehyde
 - 2. Evidence in support of the Calvin Cycle
- C. Photorespiration
 - 1. The oxygenase reaction of RuBisCO
 - 2. Carboxylase/Oxygenase specificity
 - 3. Regeneration of 3-phosphoglycerate
- D. C4 Pathways of Carbon Dioxide Fixation
 - 1. PEPCase reaction
 - a. comparisons of C3 and C4 lifestyles
 - 2. Mechanisms of carbon dioxide recycling to RuBisCO
 - 3. C4 ecophysiology

There is a vast scope of carbon dioxide fixing strategies. All were presented at the level of transformation of chemical structures. With the exception of some of the anoxygenic prokaryotes, the Calvin Cycle is central. Any variations amount to biochemical and/or transport mechanisms designed to concentrate CO₂ at the site of RuBisCO.

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VIII. Metabolic Flux

- A. C3 Pathway Bioengineering
 - 1. Flux control coefficients
 - 2. Modifying activity of C3 pathway enzymes
 - a. sedoheptulose 1,7 bisphosphatase
 - 3. Systems Bioengineering
 - a. transformation to enhance carbon concentration mechanisms
 - b. transformation to introduce novel photorespiratory pathways

The integration of the multiple biochemical requirements for life, utilizing the ATP and reducing equivalents provided by photosynthesis, were presented. The results of experiments bioengineering plants by transformation to modulate expression of key enzymes of the Calvin Cycle and the impact on biomass production were discussed. The most recent development of transformation technologies and introduction of genes for novel pathways was highlighted as the next revolution in photosynthesis.

Finally, the remarkable mechanisms of photosynthesis are placed in the context of biological systems. Here, students are provided with deep insight into systems approaches to photosynthesis. The Learning Objective is synthesis and integration of the biochemical complexities of photosynthesis at the level of the whole organisms.

LABORATORY EXERCISES (3 hours per week)

Origins and Fate of Light

Spectral Properties of Intact Leaves (**Week One**)

Visualizing the Photosynthetic Apparatus (**Week Two**)

Absorbing Light (**Week Three**)

Fluorescence and Reaction Centers

Determining Excitation and Emission Spectra (**Week Four**)

Delayed Fluorescence (**Week Four**)

Oxygen Electrode: Pathways of Photosynthetic Electron Transport

P/O ratios (**Week Five**)

Carbon dioxide coupling in chloroplasts and algae (**Week Six**)

Chloroplast Molecular Biology

Isolation of chloroplast DNA(**Week Seven**)

Chloroplast genome mapping (**Week Eight**)

Presentations: Pathways of Carbon Dioxide Fixation (**Week Nine**)

The laboratory exercises provided a detailed introduction to the scope of experimental techniques used to explore the primary processes in photosynthesis: From the light reactions, synthesis of chemical energy, and integration with carbon dioxide fixation, to, finally, the genomic identity of the chloroplast. The techniques included spectrometry (reflectance, absorbance and fluorescence), oxygen electrode, and genome mapping using restriction endonucleases.

Laboratory exercises are tightly integrated with lecture content (and, in fact, the material is synchronized with lecture coverage). Knowledge alone is insufficient. To work with one's hands, to experience how experiments are performed is fundamental to the Learning Objective of understanding and experiencing photosynthesis.

Course Director

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Textbook

Lawlor, D.W. (2001) Photosynthesis (3d edition). Springer-Verlag

1. Introduction to the photosynthetic process
2. Light-the driving force of photosynthesis
3. Light harvesting and energy capture in photosynthesis
4. Architecture of the photosynthetic apparatus
5. Electron and proton transport
6. Synthesis of ATP: photophosphorylation
7. The chemistry of photosynthesis
8. Metabolism of photosynthetic products
9. C₄ photosynthesis and crassulacean acid metabolism
10. Molecular biology of the photosynthetic system
11. Carbon dioxide supply for photosynthesis
12. Photosynthesis by leaves
13. Photosynthesis, plant production and environment

Assignments and Grading

Lecture Component: 70%

Two Term Tests and a Final Exam (60%): The lowest score is worth 15%, the middle is worth 20% and the highest score is worth 25%.

One Assignment (10%): Focused in methods in photosynthesis based on a research paper.

Laboratory Component: 30%

Two lab reports on selected lab exercises, the lowest weighted 10%, the highest weighted 15%. Participation (5%).

In the event of a documented absence from a term test, 15% will be carried over to the final and other term test (lowest score worth 27.5%, highest worth 32.5%)