

Angio-adaptation in unloaded skeletal muscle: new insights into an early and muscle type-specific dynamic process

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With a remarkable plasticity, skeletal muscle adapts to an altered functional demand. Muscle angio-adaptation can either involve the growth or the regression of capillaries as respectively observed in response to endurance training or muscle unloading. Whereas the molecular mechanisms that regulate exercise-induced muscle angiogenesis have been extensively studied, understanding how muscle unloading can in contrast lead to capillary regression has received very little attention. Here we have investigated the consequences of a 9 day time course hindlimb unloading on both capillarization and expression of angio-adaptive molecules in two different rat skeletal muscles. Both soleus and plantaris muscles were atrophied similarly. In contrast, our results have shown different angio-adaptive patterns between these two muscles. Capillary regression occurred only in the soleus, a slow-twitch and oxidative postural muscle. Conversely, the level of capillarization was preserved in the plantaris, a fast-twitch and glycolytic muscle. We have also measured the time course protein expression of key pro- and anti-angiogenic signals (VEGF-A, VEGF-B, VEGF-R2, TSP-1). Our results have revealed that the angio-adaptive response to unloading was muscle-type specific, and that an integrated balance between pro- and anti-angiogenic signals plays a determinant role in regulating this process. In conclusion, we have brought new evidence that measuring the ratio between pro- and anti-angiogenic signals in order to evaluate muscle angio-adaptation was a more accurate approach than analysing the expression of molecular factors taken individually.

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Abbreviations CD, capillary density; C/F, capillary-to-fibre ratio; FCSA, fibre cross-sectional area; HU, hindlimb unloading; Mdm2, murine double minute-2; PVD, peripheral vascular diseases; TSP-1, thrombospondin-1; VEGF-A, vascular endothelial growth factor-A; VEGF-B, vascular endothelial growth factor-B; VEGF-R2, VEGF receptor-2.

Introduction

In response to various physiological or pathological conditions, skeletal muscle responds to an altered functional demand with a remarkable plasticity (Booth & Thomason, 1991). This includes changes in muscle size, in myofibres' contractile and metabolic phenotype, and also in the vasculature (Booth & Thomason, 1991; Hudlicka *et al.* 1992). Muscle angio-adaptation can either involve angiogenesis or capillary regression as respectively observed in response to exercise training or muscle unloading (Hudlicka *et al.* 1992; Egginton, 2009). Whereas the molecular mechanisms that regulate exercise-induced muscle angiogenesis have been extensively studied

(Hudlicka *et al.* 1992; Prior *et al.* 2004; Breen, 2007; Olfert *et al.* 2009; Malek & Olfert, 2009; Hudlicka & Brown, 2009; Egginton, 2009), understanding how muscle unloading can in contrast lead to capillary regression has received very little attention.

Hindlimb unloading (HU) is a well-established rodent model of hypokinesia and hypodynamia, often used to study the effect of weightlessness on skeletal muscle (Morey-Holton & Globus, 2002). HU results in a strong reduction in skeletal muscle contractile activity and leads to myofibre atrophy, reduced muscle blood flow, and capillary regression (Desplanches *et al.* 1987; Morey-Holton & Globus, 2002; Machida & Booth, 2004).

Previous studies have focused mostly on the soleus, a weight-bearing skeletal muscle. After 5 weeks of HU, this muscle was atrophied by 50% and its level of capillarization was decreased by 40% (Desplanches *et al.* 1987). The process of capillary regression, however, started as early as the first week of HU (Kano *et al.* 2000; Däpp *et al.* 2004). Interestingly, soleus from Wistar rats and C57/bl6 mice underwent a similar 20% capillary regression after 1 week of HU despite strain-specific differences in their fibre-type composition (Kano *et al.* 2000; Däpp *et al.* 2004). In addition, Kano and collaborators (2000) have shown no additional capillary regression between 1 and 3 weeks of HU.

Muscle angio-adaptation is tightly regulated by a balance between pro- and anti-angiogenic signals. To date, the pro-angiogenic vascular endothelial growth factor-A (VEGF-A) and the anti-angiogenic thrombospondin-1 (TSP-1) are the most well characterized regulators of skeletal muscle angio-adaptation (Tang *et al.* 2004; Olfert *et al.* 2006, 2009; Malek & Olfert, 2009; Malek *et al.* 2010). Their functional importance has been well demonstrated in the context of exercise-induced muscle angiogenesis (Olfert *et al.* 2009; Malek & Olfert, 2009). Surprisingly, the molecular factors and mechanisms responsible for the loss of capillaries in response to HU remain largely unknown.

It was recently reported that VEGF-A mRNA expression was unaffected after 10 days of HU in mouse gastrocnemius muscle whereas VEGF receptor mRNA levels were decreased (Wagatsuma, 2008). Unfortunately, neither VEGF protein levels nor muscle capillarization were measured in this study. Moreover, mRNA levels were measured at one single time point only, where it is known that about 20% of capillary regression has already occurred (Kano *et al.* 2000; Däpp *et al.* 2004). Muscle angio-adaptation is a very dynamic biological process and the expression of angio-adaptive factors and their receptors can be very rapidly affected in skeletal muscle in response to physiological changes such as alterations in blood flow or contractile activity (Egginton, 2009; Hudlicka & Brown, 2009). No study has ever investigated the time course effect of HU simultaneously on both muscle capillarization and expression of angio-adaptive factors. Therefore, there is an important need to conduct dynamic studies in order to better understand the molecular mechanisms responsible for HU-induced capillary regression.

Here, we propose to revisit the concept of muscle capillary regression through a kinetic study in two rat skeletal muscles: the oxidative and slow-twitch soleus *versus* the glycolytic and fast-twitch plantaris. We have compared the time course changes in both capillarization and protein expression for key angio-adaptive molecules in response to HU. While both muscles atrophied similarly, our results have shown different angio-adaptive

patterns between soleus and plantaris leading to capillary regression in the soleus only.

Methods

Animal care

The animal care and use were approved by the Institutional Animal Care Committee of Université Lyon 1 and was conducted accordingly to the directives of the *Declaration of Helsinki* for the use of laboratory animals.

Hindlimb unloading model

Forty-two female Wistar rats (mean body weight 200 g) were purchased from Charles River (L'Arbresles, France) and housed on a 12:12 h light–dark cycle, with water and food access *ad libitum*. After 1 week of acclimatization, animals were divided into seven groups corresponding to different durations of hindlimb unloading: 0, 0.5, 1, 3, 5, 7 and 9 days ($n = 6/\text{group}$). Hindlimb unloading was performed by tail suspension as previously described (Desplanches *et al.* 1987; Morey-Holton & Globus, 2002; Flück *et al.* 2005).

Killing and tissue processing

Rats were weighed and then anaesthetized by ketamine (80 mg kg^{-1})–xylazine (10 mg kg^{-1}) intraperitoneal injection. Soleus and plantaris muscles were harvested, weighed and fast-frozen in liquid nitrogen. Killing was performed by exsanguinations and heart removal.

Immunohistochemistry

Frozen $8 \mu\text{m}$ transverse cross-sections were cut in the mid-belly of the muscle. Capillaries were visualized after a brief fixation in cold acetone followed by staining for alkaline phosphatase activity by incubation with 5-bromo-4-chloro-3-indolyl phosphate/nitro blue tetrazolium (FAST BCIP/NBT, Sigma-Aldrich, Oakville, ON, Canada) for 45 min at 37°C (Roudier *et al.* 2009). Pictures were acquired using an Axio-Imager (Zeiss, Jena, Germany) equipped with an AxioCam camera (Zeiss). Capillary density (CD) and capillary-to-fibre ratio (C/F) were determined after counting capillaries and myofibres on three to five cryosections from each muscle and in quintuplet on each of these cryosections. Mean fibre cross-sectional area (FCSA) was calculated from the total number of fibres divided by the total muscle area. As the soleus and the plantaris muscles are not homogeneous in their fibre-type composition, counting areas were randomly chosen all over the muscle cross-section

Table 1. Morphometric values in rats subjected to a 9 days time course hindlimb unloading

	Hindlimb unloading duration (days)					
	0	1	3	5	7	9
Mean body weight (g)	197 ± 4	203 ± 3	190 ± 6	190 ± 5	197 ± 4	195 ± 2
Soleus weight (mg)/ body weight (g)	0.40 ± 0.01	0.41 ± 0.01	0.38 ± 0.02	0.36 ± 0.02	0.31 ± 0.01***, ¶¶¶, ††	0.31 ± 0.02***, ¶¶¶, ††
Soleus mean FCSA (µm ²)	2532 ± 75	2437 ± 122	2397 ± 142	1833 ± 68***, ¶¶, ††	1621 ± 86***, ¶¶¶, †††	1879 ± 105***, ¶¶, ††
Plantaris weight (mg)/ body weight (g)	0.98 ± 0.03	0.85 ± 0.04**	0.87 ± 0.02**	0.85 ± 0.03*	0.84 ± 0.02**	0.78 ± 0.02***
Plantaris mean FCSA (µm ²)	2027 ± 86	2169 ± 236	1933 ± 71	1518 ± 50**, ¶, †	1578 ± 118**, ¶, †	1496 ± 58**, ¶, †

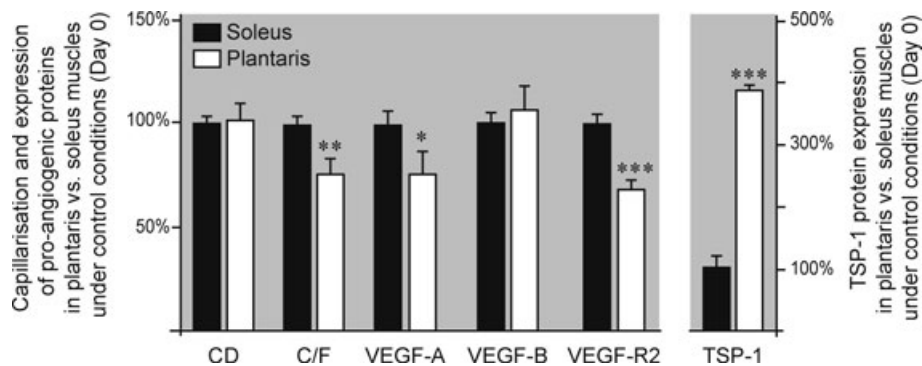
Both soleus and plantaris muscles atrophied to a similar extent after 9 days of hindlimb unloading. Based on relative muscle weight, the plantaris muscle was atrophied as early as day 1 ($P \leq 0.01$). Atrophy was delayed in soleus, significantly occurring only at day 7 ($P \leq 0.001$). Based on the mean fibre cross-sectional area (FCSA), atrophy presented similar kinetics and amplitude in both muscles. Significantly different from day 0: * $P \leq 0.05$; ** $P \leq 0.01$; *** $P \leq 0.001$. Significantly different from day 1: ¶ $P \leq 0.05$; ¶¶ $P \leq 0.01$; ¶¶¶ $P \leq 0.001$. Significantly different from day 3: † $P \leq 0.05$; †† $P \leq 0.01$; ††† $P \leq 0.001$. Data are expressed as means \pm s.e.m. ($n = 6$ rats/group).

in order to provide a global estimation of muscle C/F, CD and FCSA by muscle-type. An average of 120–180 fibres and 140–350 capillaries were counted on each counting area.

Western blotting

Proteins were extracted from 20–40 mg of frozen muscles using a RIPA lysis buffer containing 1 mg ml⁻¹ PMSF, 1 mM Na₃VO₄, 1 mM NaF (Sigma-Aldrich), 1 \times protease inhibitors cocktail (Complete Mini and PhosStop, Roche Diagnostics, Laval, QC, Canada). Muscle lysates were performed in a Retsch MM400 tissue lyser (Retsch GmbH, Haan, Germany). Denatured samples were separated on SDS-PAGE and blotted onto either a PVDF membrane (Immun-Blot, Bio-Rad, Mississauga, ON, Canada) or 0.45 µm nitrocellulose membrane (Whatman Protran BA95, Sigma-Aldrich). After blocking with 5% fat-free milk, membranes were probed using

antibodies against VEGF-A (1:500, sc-507, Santa-Cruz Biotechnology, Santa-Cruz, CA, USA), VEGF-B (1:500, H70, sc-13083, Santa-Cruz Biotechnology), TSP-1 (1:500, clone A6.1, no. 399300, Invitrogen, Burlington, ON, Canada), Flk-1 (1:500, C1158, sc-504, Cell Signaling, Pickering, ON, Canada), phospho-p53 (1:500, Ser15, no. 9284S, Cell Signaling), p53 (1:200, clone DO.1, sc-126, Santa-Cruz Biotechnology), Mdm2 (1:75, Clone 2A10, kindly provided by Dr Mary E. Perry, NCI-NIH, Frederick, MD, USA), β -tubulin (1:750, no. 2148, Cell Signaling), HRP-conjugated anti-mouse (1:1000, p0260, Dako, Mississauga, ON, Canada; or NA-931, GE Healthcare, Piscataway, NJ, USA) and HRP-conjugated anti-rabbit (1:1000, p0217, Dako). Proteins were visualized using an enhanced chemiluminescence procedure (Super Signal West Pico, Fisher Scientific, Markham, ON, Canada) on X-Ray film or using a Kodak Imaging station 4000MM Pro. Equal protein loading was verified before

**Figure 1. Muscle-type differences in capillarization and angio-adaptive proteins**

Relative levels of capillarization and angio-adaptive proteins in the plantaris muscle compared to the soleus muscle, under control conditions (i.e. day 0). Values have been normalized to soleus levels. Data are expressed as means \pm s.e.m. ($n = 6$ rats/group). Significant difference between soleus and plantaris muscles: * $P \leq 0.05$; ** $P \leq 0.01$; *** $P \leq 0.001$.

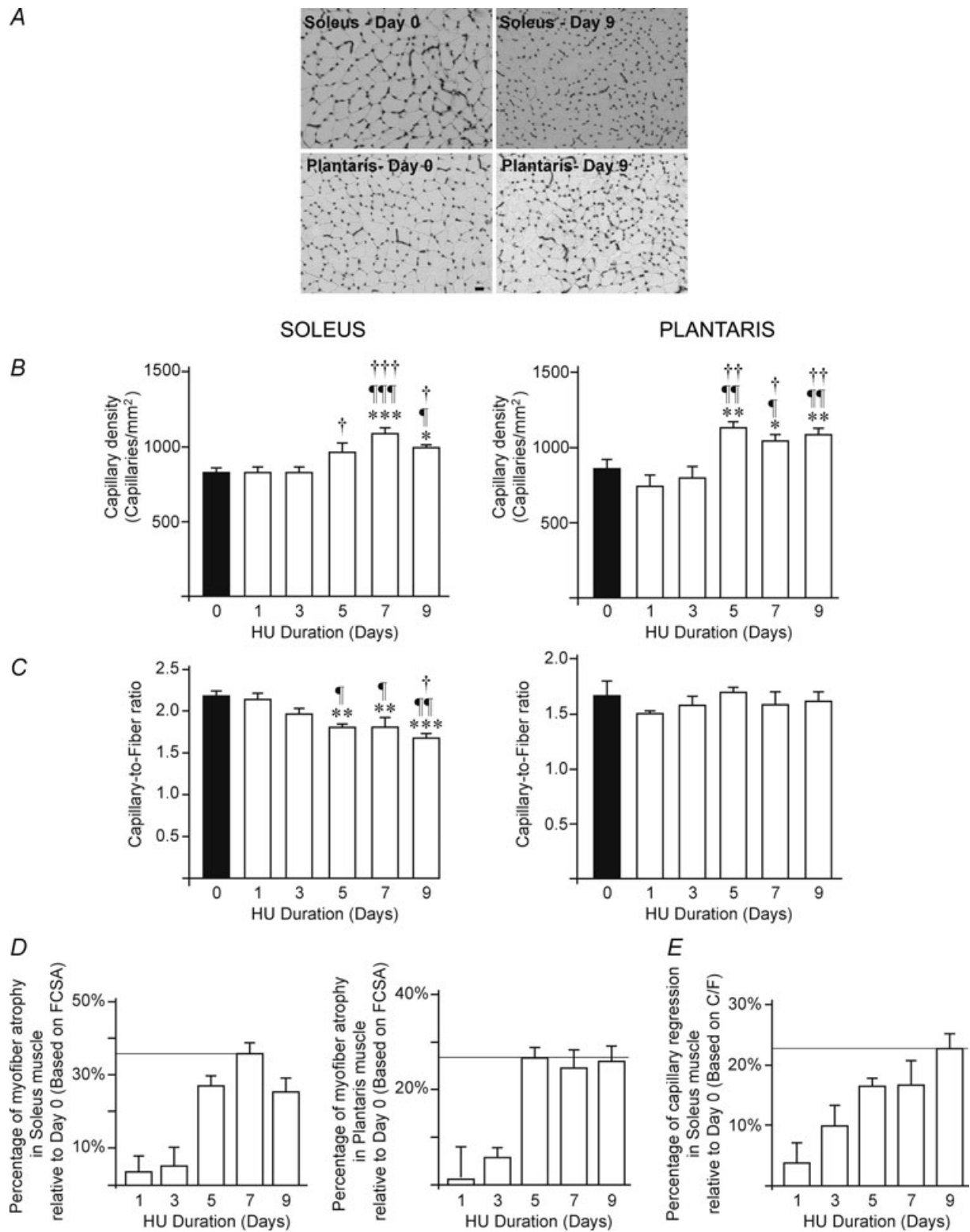


Figure 2. Hindlimb unloading and skeletal muscle capillarization

A, representative images of capillaries staining in soleus (left) and plantaris (right) muscles from control animals (day 0, top panel) and after 9 days of hindlimb unloading (day 9, bottom panel). Scale bar, 50 μ m. B–C, quantitative measurement of capillary density (B) and capillary-to-fibre ratio (C) in soleus (left panel) and plantaris (right panel) muscles following time course hindlimb unloading (HU, from days 0 to 9). Data are expressed as means \pm S.E.M. ($n = 6$ rats/group). Significantly different from day 0: * $P \leq 0.05$; ** $P \leq 0.01$; *** $P \leq 0.001$. Significantly different

immunodetection by Ponceau S staining of the membrane and by immunodetection of β -tubulin as our loading control. All western blots were run in duplicate. Quantifications of protein expression levels were carried out using NIH Image 1.62 Software and expressed as densitometric arbitrary units (DAIs). The levels of expression for our proteins of interest (VEGF-A, VEGF-B, Flk-1, TSP-1, Mdm2, phospho-p53, and p53) were individually normalized to β -tubulin expression level, and ratios were compared between the different HU time points (day 0 to day 9).

Statistical analyses

Statistical analyses were performed with Prism5 for Mac OSX (v. 5.0, GraphPad Software Inc., La Jolla, CA, USA). One way ANOVA using a Newman-Keuls *post hoc* test was applied and results were considered to be statistically significant when $P \leq 0.05$.

Results

Muscle-type differences in capillarization and angio-adaptive proteins

Figure 1 illustrates the differences in the percentage of capillarization and expression of angio-adaptive proteins between control (i.e. day 0) soleus and plantaris muscles. Protein levels were determined by Western blotting. The capillary-to-fibre ratio (C/F) was 24% lower in plantaris muscle than in the soleus ($P \leq 0.01$). The plantaris muscle expressed less pro-angiogenic VEGF-A and VEGF-R2 signals and more anti-angiogenic TSP-1 than the soleus: -25% for VEGF-A ($P \leq 0.05$), -33% for VEGF-R2 ($P \leq 0.001$), and $+287\%$ for TSP-1 ($P \leq 0.001$).

Hindlimb unloading and skeletal muscle atrophy

Both soleus and plantaris muscles were atrophied to a similar extent in response to time course hindlimb unloading (HU, Table 1). The muscle weight relative to body weight was decreased at day 9 by respectively 22% and 20% in soleus and plantaris muscles when compared to day 0 (soleus: $0.31 \pm 0.02 \text{ mg g}^{-1}$ at day 9 vs. $0.40 \pm 0.01 \text{ mg g}^{-1}$ at day 0, $P \leq 0.001$; plantaris: $0.78 \pm 0.02 \text{ mg g}^{-1}$ at day 9 vs. $0.98 \pm 0.03 \text{ mg g}^{-1}$ at day 0, $P \leq 0.01$). The mean fibre cross-section area (FCSA) followed a similar pattern in soleus and plantaris muscles with a significant decrease starting at day 5

and reaching -26% at day 9 (soleus: $1833 \pm 68 \mu\text{m}^2$ at day 5 and $1879 \pm 105 \mu\text{m}^2$ at day 9 vs. $2532 \pm 75 \mu\text{m}^2$ at day 0, $P < 0.001$; plantaris: $1518 \pm 50 \mu\text{m}^2$ at day 5 and $1496 \pm 58 \mu\text{m}^2$ at day 9 vs. $2027 \pm 86 \mu\text{m}^2$ at day 0, $P \leq 0.05$). Unexpectedly, the plantaris relative weight at day 1 was already significantly lower than at day 0 (-13% , $0.85 \pm 0.04 \text{ mg g}^{-1}$ at day 1 vs. $0.98 \pm 0.03 \text{ mg g}^{-1}$ at day 0, $P \leq 0.01$).

Skeletal muscle capillarization in response to hindlimb unloading

Figure 2A shows representative pictures of soleus (left panel) and plantaris (right panel) muscle cross-sections after alkaline phosphatase staining. The capillary density (CD) and the capillary-to-fibre ratio (C/F) were measured in both soleus and plantaris muscles (Fig. 2B–C). In response to HU-induced myofibre atrophy, CD was significantly increased after 5 days of HU in both muscles reaching at day 9 respectively $+18\%$ in soleus and $+27\%$ in plantaris (Fig. 2B; soleus: $994 \pm 22 \text{ capillaries (cap.) mm}^{-2}$ at day 9 vs. $836 \pm 28 \text{ cap. mm}^{-2}$ at day 0, $P < 0.05$; plantaris: $1087 \pm 39 \text{ cap. mm}^{-2}$ at day 9 vs. $852 \pm 63 \text{ cap. mm}^{-2}$ at day 0, $P < 0.05$).

However, capillary regression only occurred in the soleus muscle as reflected by a significant 17% decrease in C/F ratio after 5 days of HU, reaching -23% at day 9 (Fig. 2C, left panel: 1.81 ± 0.03 at day 5 and 1.68 ± 0.05 at day 9 vs. 2.17 ± 0.06 at day 0, respectively $P < 0.01$ and $P < 0.001$). Percentages of capillary regression (Fig. 2E) and myofibre atrophy (Fig. 2D) followed a similar profile in soleus muscle.

In contrast, the equipment in capillaries was preserved in plantaris muscle (Fig. 2C, right panel; C/F: 1.62 ± 0.08 at day 9 vs. 1.66 ± 0.14 at day 0).

Expression of pro-angiogenic proteins during hindlimb suspension

We investigated the time course expression of VEGF-A, VEGF-B and VEGF receptor 2 (VEGF-R2) proteins in response to muscle unloading.

In soleus muscle, VEGF-A protein expression was unaffected by HU (Fig. 3A, left panel). VEGF-B expression showed two peaks at respectively days 3 and 7 of HU where protein levels were increased in comparison to all other time point measurements (Fig. 3B, left panel: 1.01 ± 0.10 densitometry arbitrary units (DAI) at day

from day 1: ¶ $P \leq 0.05$; ¶¶ $P \leq 0.01$; ¶¶¶ $P \leq 0.001$. Significantly different from day 3: † $P \leq 0.05$; †† $P \leq 0.01$; ††† $P \leq 0.001$. Significantly different from day 5: # $P \leq 0.05$. Percentage of muscle atrophy (D) and capillary regression (E) based on mean FCSA and C/F values, respectively. Data are expressed as means \pm S.E.M. ($n = 6$ rats/group). FCSA, mean fibre cross-sectional area; C/F, capillary-to-fibre ratio.

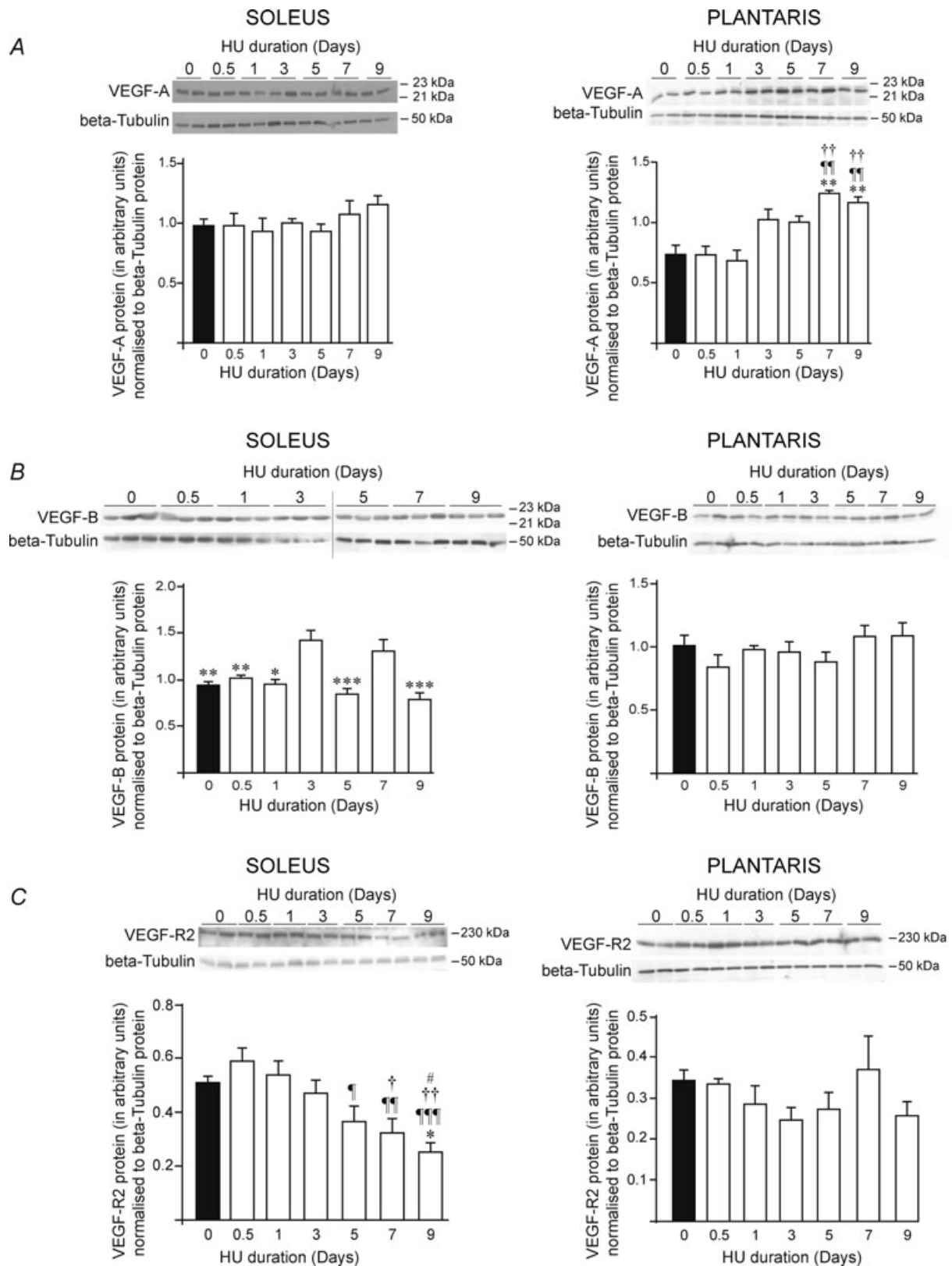


Figure 3. VEGF-A, VEGF-B and VEGF receptor-2 protein expression in response to hindlimb unloading
 Animals were subjected to a 9-day time course hindlimb unloading. Protein levels were measured by western blotting in soleus and plantaris muscles, left and right panels respectively. β -Tubulin protein was used as a loading control. Top-panels of A, B and C show images of VEGF-A, VEGF-B and VEGF-R2 protein expressions, respectively.

0 vs. 1.43 ± 0.12 DAI at day 3 (+50%), and vs. 1.30 ± 0.15 DAI at day 7 (+37%), $P < 0.01$). Interestingly, VEGF-R2 protein expression was progressively decreased by 50% over the 9-day HU time course period (Fig. 3C, left panel: 0.51 ± 0.03 DAI at day 0 vs. 0.25 ± 0.04 DAI at day 9, $P < 0.05$).

In plantaris muscle, VEGF-A protein expression was increased in response to HU by +68% and +57% after 7 and 9 days of HU, respectively (Fig. 3A, right panel: 1.24 ± 0.03 DAI at day 7 and 1.16 ± 0.06 DAI at day 9 vs. 0.74 ± 0.10 DAI at day 0, $P < 0.01$). In contrast, HU affected neither VEGF-B nor VEGF-R2 protein expression in plantaris muscle (Fig. 3B and C, right panels).

Thrombospondin-1, p53 and murine double minute-2 response to hindlimb unloading

In soleus muscle, thrombospondin-1 (TSP-1) protein expression was significantly increased by 137% after 5 days of unloading, reaching a maximum at day 7 (+258%), and remaining increased by 185% at day 9 (Fig. 4A, left panel. vs. day 0 (0.26 ± 0.05 densitometry arbitrary units, DAI): 0.61 ± 0.07 DAI at day 5, $P < 0.05$; 0.92 ± 0.14 DAI at day 7, $P < 0.001$; and 0.73 ± 0.11 DAI at day 9, $P < 0.01$). Expression of the transcription factor p53, which has been previously shown to regulate TSP-1 (Dameron *et al.* 1994), was increased in soleus in response to HU (Fig. 4B). An early and acute p53 response was observed 12 h after the onset of HU (+161%), and was then significantly increased between day 3 (+130%) and day 9 (+230%) (Fig. 4B: 0.49 ± 0.07 DAI at day 0 vs. 1.14 ± 0.08 DAI at days 5 and 7 ($P < 0.01$), and vs. 1.63 ± 0.16 DAI at day 9, $P < 0.001$). The expression of murine double minute-2 (Mdm2), a well-described p53 downstream target, followed the same pattern as TSP-1 (Fig. 4C). Mdm2 protein level was significantly increased by 97% after 5 days of HU, reaching +132% at day 9 (Fig. 4C: 0.38 ± 0.05 DAI at day 0 vs. 0.74 ± 0.04 DAI at day 5, and vs. 0.87 ± 0.05 DAI at day 9, $P < 0.01$).

In contrast, the protein level of TSP-1 remained unchanged in plantaris muscle at all HU time points (Fig. 4A, right panel).

Muscle type-specific regulation of the angioadaptive balance

We have calculated the ratio between the pro- (VEGF-A, VEGF-B, VEGF-2) and the anti-angiogenic (TSP-1)

proteins in order to determine how the angio-adaptive balance was altered by HU in both skeletal muscles. The VEGF-A/TSP-1 balance was affected by HU in an opposite manner between soleus and plantaris muscles (Fig. 5A).

In soleus muscle, a significant decrease in VEGF-A/TSP-1 ratio was detected from day 5 to day 9 of HU when compared to day 0 (Fig. 5A, left panel: 6.13 ± 2.14 at day 0 vs. 2.00 ± 0.50 at day 5 (−63%), 1.35 ± 0.32 at day 7 (−78%), and 1.62 ± 0.16 at day 9 (−73%), $P < 0.05$).

In plantaris muscle, VEGF-A/TSP-1 ratio increased progressively over the HU time course, reaching significance at day 7 only (Fig. 5A, right panel: 0.75 ± 0.07 at day 0 vs. 1.41 ± 0.16 at day 7 (+88%), $P < 0.05$). The temporal relationship between changes in capillarization and in the ratios of individual pro-angiogenic proteins (VEGF-R2, VEGF-A, VEGF-B), as well as their sum, relative to TSP-1 are shown in Fig. 5B. In the soleus, all ratios followed the same pattern with a decrease over time (Fig. 5B, left panel). This suggests that the observed decrease in TSP-1 plays a determinant role in the angio-adaptive response to HU. In contrast, the VEGF-A/TSP-1 ratio and the ratio between all pro-angiogenic proteins and TSP-1 were increased over time in plantaris muscle (Fig. 5B, right panel).

All together, these results indicate that in both muscles the balance between all proangiogenic protein and TSP-1 perfectly matched the level of capillarization in the tissue.

Discussion

Here, we investigated in a time course manner and in two different muscle types the unloading-induced changes in both muscle capillarization and angio-adaptive molecule expression. The soleus muscle is a slow-twitch and very oxidative weight-bearing muscle, considerably affected by HU. The plantaris muscle is a fast-twitch and glycolytic muscle. Choosing a time course approach rather than one single time point measurement was crucial to be certain to overlap the dynamic molecular and tissue events known to occur within the first week of unloading (Kano *et al.* 2000; Däpp *et al.* 2004). We have also measured the expression of angio-adaptive molecules at the protein level rather than mRNA level since proteins would be more likely to be the key actors initiating and driving the capillary regression process.

Bottom panels of A, B and C show densitometric analyses of VEGF-A, VEGF-B or VEGF-R2 proteins relative to β -tubulin. Densitometric analyses show means \pm s.e.m. ($n = 6$ rats/group). Significant differences: vs. day 0 ** $P \leq 0.01$; *** $P \leq 0.001$; vs. day 0.5: ¶ $P \leq 0.05$; ¶¶ $P \leq 0.01$; ¶¶¶ $P \leq 0.001$; vs. day 1: † $P \leq 0.05$; †† $P \leq 0.01$; ††† $P \leq 0.001$; vs. day 3 # $P \leq 0.05$; ## $P \leq 0.01$; ### $P \leq 0.001$; vs. day 7 ‡‡ $P \leq 0.01$.

Unloaded soleus and plantaris muscles undergo similar muscle atrophy but different angio-adaptation

Our results brought evidence that despite similar muscle atrophy, angio-adaptation was different between soleus and plantaris muscles. Soleus and plantaris muscles were both atrophied to a similar extent after 9 days of HU, consistent with previous studies focusing on 7 or 15 days of HU in rats (McDonald *et al.* 1992*a,b*; Kano *et al.* 2000; Fujino *et al.* 2005). Whereas relative muscle weight and FCSA both started to decrease significantly in soleus muscle after 5 days of HU, we observed an unexpected 13% decrease in the plantaris relative weight after day 1. However, this was not associated with any concomitant decrease in FCSA, which was decreased at day 5 as in the soleus muscle.

Capillary density (CD) was significantly increased to a similar level in soleus and plantaris muscles in response to HU, most likely as a result of myofibre atrophy. However, changes in CD do not strictly reflect any capillary loss or growth. Analysis of the capillary-to-fibre ratio (C/F) revealed that capillary regression occurred in soleus muscle after 5 days of HU, concomitantly with FCSA reduction and reaching a 23% decrease at day 9 consistent with the existing literature (Kano *et al.* 2000; Däpp *et al.* 2004; Fujino *et al.* 2005). Conversely, capillarization was preserved in plantaris muscle over the 9 days of HU despite similar myofibre atrophy as observed in the soleus.

Regulation of the angio-adaptive balance differs between soleus and plantaris muscles

Gene deletion of the pro-angiogenic vascular endothelial growth factor-A (VEGF-A) and of the anti-angiogenic thrombospondin-1 (TSP-1) respectively led to decreased and increased basal capillarization in mouse skeletal muscle (Tang *et al.* 2004; Olfert *et al.* 2009; Malek & Olfert, 2009). In consequence, animal exercise capacity was respectively decreased in VEGF-A deficient mice and increased in TSP-1 deficient ones (Olfert *et al.* 2009; Malek & Olfert, 2009). Here we showed that HU affected VEGF-A and TSP-1 protein expression in a muscle type-specific manner.

In the soleus, where capillaries regressed, the time course expression of VEGF-A protein was unaffected by HU, consistent with the study from Wagatsuma (2008), reporting no change in VEGF-A mRNA levels in gastrocnemius from CD1 mice subjected to 10 days of HU. In contrast, we found that VEGF-A protein expression was significantly increased after 7 and 9 days of HU in the plantaris muscle where capillarization was preserved.

Conversely to HU, muscle overload leads to myofibre hypertrophy and capillary growth. A 20% increase in C/F ratio was reported after 2 weeks of overload of the extensor digitorum longus muscle (Brown & Hudlicka,

2003; Williams *et al.* 2006*a,b*). Interestingly, treating the animals with a VEGF-A Trap prevented this angiogenic response (Williams *et al.* 2006*b*). In their study, Degens *et al.* (2003) have reported increased expression of VEGF-A mRNA and protein in rat plantaris muscle overloaded for 2 weeks. Recently, Parvaresh *et al.* (2010) have compared VEGF-A expression in overloaded soleus and plantaris muscles. VEGF-A expression was transiently increased in the plantaris muscle after 3 days of overload, returning to the level of unloaded muscle after 7 days. Surprisingly, VEGF-A expression in overloaded soleus followed the opposite pattern: a transient decrease at day 3 and a return to unloaded values at day 7 (Parvaresh *et al.* 2010).

Altogether, these results from overloading experiments and from our unloading study support the idea that increased VEGF-A levels might determine the maintenance or the growth of muscle capillaries. Interestingly, Parvaresh *et al.* (2010) suggest that regulation of VEGF-A expression by MyoD might explain the contrasting response between overloaded soleus and plantaris. VEGF-A promoter indeed possesses some MyoD-specific consensus sites (Bryan *et al.* 2008). Microarray analysis from 3-day overloaded soleus have shown no change for MyoD expression whereas an increase was found in plantaris muscle after 2 and 3 days of overload (Mozdziak *et al.* 1998; Carson *et al.* 2002; Hyatt *et al.* 2008). In the context of muscle unloading, MyoD expression seems to be more affected by HU in plantaris than in soleus muscle from young rats (Alway *et al.* 2001). Whereas MyoD mRNA and protein were respectively increased by 235% and 1600% in the plantaris, only MyoD protein was increased in soleus muscle by 85% (Alway *et al.* 2001). This is an interesting observation that could explain why our HU data have shown no change in VEGF-A expression in the soleus but an increase in plantaris muscle.

In the soleus, the maintenance of VEGF-A alone is not sufficient to prevent the capillary regression observed in response to unloading. Angio-adaptation is fine-tuned by a balance between pro- and anti-angiogenic signals. We therefore measured protein expression levels for VEGF-B and VEGF receptor-2 (VEGF-R2).

VEGF-R2 has been described as mediating most of VEGF-A pro-angiogenic signalling (Milkiewicz *et al.* 2006; Olfert *et al.* 2009; Ferrara, 2009). Consistent with the results from Wagatsuma (2008) at the mRNA levels, VEGF-R2 protein expression was significantly decreased in our soleus samples after 5 days of HU, possibly contributing to an angio-adaptive balance in favour of some vascular destabilization and capillary regression.

VEGF-B is another member of the VEGF family expressed in skeletal muscle (Olofsson *et al.* 1996*a,b*). Despite some contradictory results, most of the recent studies suggest that VEGF-B is more important for recovering angiogenesis in the ischaemic heart than in other tissues including ischaemic skeletal muscles

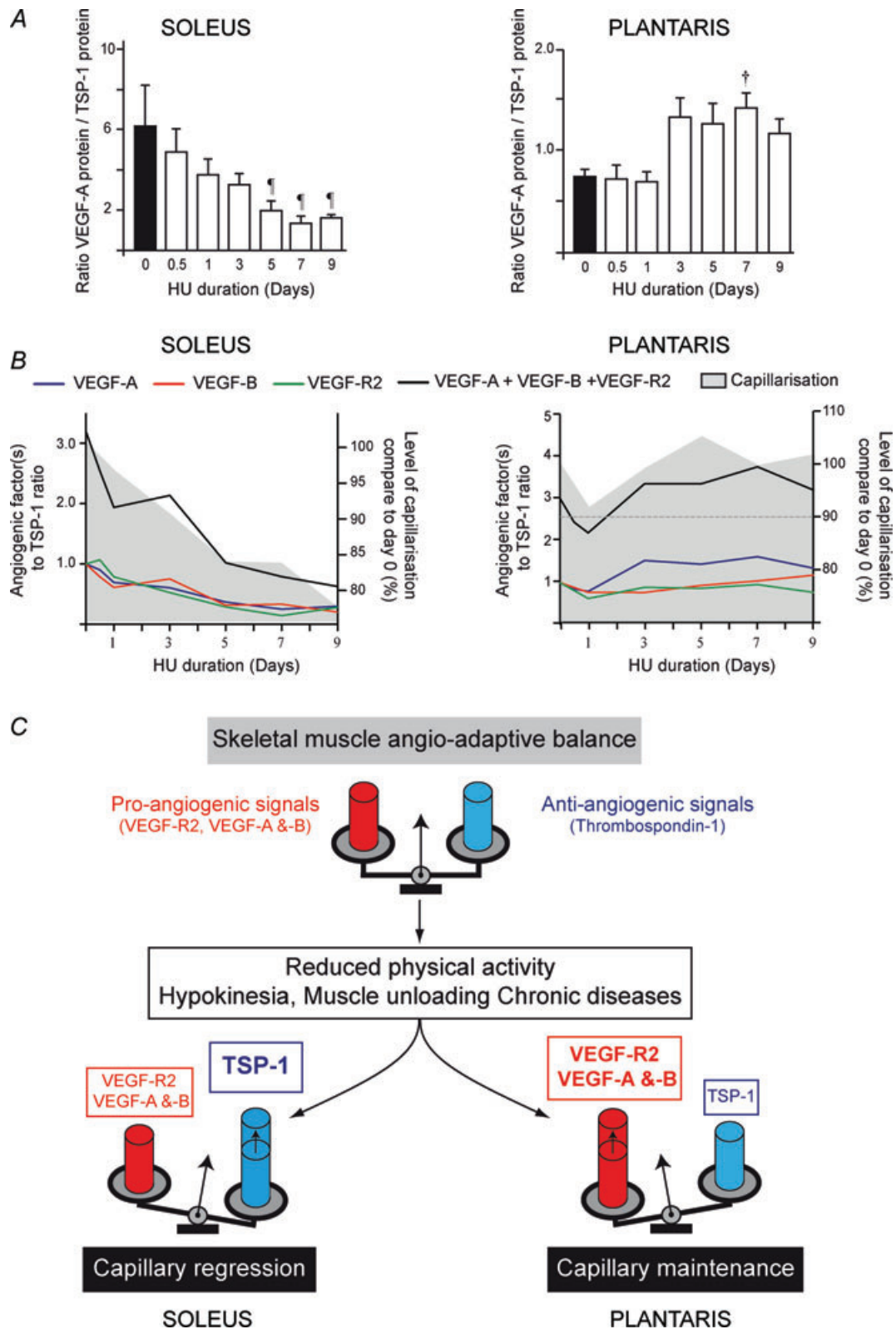


Figure 5. Hindlimb unloading and the angioadaptive balance in soleus and plantaris muscles
 The ratios between the relative expression of pro-angiogenic proteins (VEGF-A, VEGF-B and VEGF-R2) and the anti-angiogenic TSP-1 proteins were calculated during the time course of hindlimb unloading. A shows VEGF-A to TSP-1 ratio over the time of hindlimb unloading in soleus and plantaris muscles (left and right panels, respectively).

(Bellomo *et al.* 2000; Silvestre *et al.* 2003; Claesson-Welsh, 2008; Li *et al.* 2008; Li, 2010; Hagberg *et al.* 2010). However, through its binding to VEGF receptor-1, VEGF-B could limit VEGF-A trapping by this receptor, thus making VEGF-A more available for the pro-angiogenic VEGF-R2 (Claesson-Welsh, 2008). In addition, VEGF-B was recently shown to mediate some anti-apoptotic effects in endothelial cells (Li *et al.* 2008). The biological function of VEGF-B in skeletal muscle remains, then, debatable and poorly understood. Here, we observed two unexpected peaks of expression for VEGF-B protein after 3 and 7 days of HU in soleus. It is, however, very unlikely that such transient variation could exert any notable effect on preventing capillary regression. In plantaris muscle, neither VEGF-B nor VEGF-R2 protein expression were affected by HU.

Protein expression of the anti-angiogenic TSP-1 was strongly increased after 5 days of HU in the soleus concomitantly with capillary regression. In contrast, no change in TSP-1 level was observed in plantaris muscle. The tumour suppressor protein p53 has been shown to regulate TSP-1 promoter activity although data in that field still remain controversial (reviewed in Ren *et al.* 2006). TSP-1 promoter regulation by p53 seems to be indirect and to involve co-factors and/or co-repressors (Su *et al.* 2010). p53 has also been shown to impair VEGF-A expression, either directly via p53 binding to VEGF-A promoter (Qin *et al.* 1994; Pal *et al.* 2001) or indirectly through p53-mediated proteasomal degradation of HIF-1 α (An *et al.* 1998; Blagosklonny *et al.* 1998; Ravi *et al.* 2000). p53 expression was found to be increased in the cytoplasmic fraction of soleus muscle after 14 days of HU (Siu *et al.* 2006). Here, we observed a progressive increase in p53 protein expression in the soleus, starting at day 3 of HU and thus preceding TSP-1 up-regulation. We also measured murine double minute-2 (Mdm2) protein expression as a control p53 downstream target. p53 binds to Mdm2 promoter, enhances its activity, and leads to an increased Mdm2 protein expression (Vousden & Prives, 2009; Eischen & Lozano, 2009). We found that Mdm2 protein expression was increased in the soleus in response to HU with the same pattern as for TSP-1. All together, these results suggest that p53 could participate in the process of capillary regression in the soleus by up-regulating TSP-1 protein expression but does not

seem to play a role in down-regulating VEGF-A protein expression in response to HU.

Since angio-adaptation is tightly regulated by a balance between stimulatory and inhibitory signals, we have determined the ratio between pro- and anti-angiogenic proteins in our samples. In the soleus, all ratios were decreased, reflecting a balance in favour of some capillary destabilization and regression. The opposite observation was made in the plantaris where increased ratios were likely to be required for the maintenance of muscle capillarization. Interestingly, muscle capillarization followed the same time course pattern as for the different pro-/anti-angiogenic ratios in each unloaded muscle. This concept of an integrated angio-adaptive balance is illustrated in Fig. 5C.

In conclusion, our study has illustrated for the first time that the angio-adaptive response to unloading was muscle type specific and that an integrated balance between pro- and anti-angiogenic signals plays an important role in regulating this process. We have also brought new evidence that measuring the ratio between pro- and anti-angiogenic signals in order to evaluate muscle angio-adaptation was a more accurate approach than analysing the expression of molecular factors taken individually.

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Data are means \pm S.E.M. ($n = 6$ rats/group). Significant differences are vs. day 0.5, $\dagger P \leq 0.05$; and vs. day 1 $\ddagger P \leq 0.05$. *B* illustrates the time course of changes in muscle capillarization and in the ratios VEGF-A, VEGF-B, VEGF-R2, or the sum of all pro-angiogenic proteins measured (VEGF-A+VEGF-B+VEGF-R2), relative to TSP-1 in soleus and plantaris (left and right panels, respectively). Ratios at different HU time points were normalized to day 0 (equal to 1.0 arbitrary unit). Muscle capillarization at different HU time points is expressed as a percentage of capillarization at day 0. *C*, schematic illustration of how the angio-adaptive balance is differently regulated between soleus and plantaris muscles in response to hindlimb unloading.

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Author contributions

E.R. and O.B. contributed to the design of the study, performed experiments, analysed and interpreted data, and wrote the manuscript. C.G. performed animal conditioning, experiments, and analysed data. A.W. and K.D. performed Western blot analysis on Mdm2, p53, and VEGF-R2. D.D. contributed to animal conditioning. All authors contributed to the drafting and revision of the manuscript content and gave their final approval of the version to be published. Experiments were carried out at York University (Muscle Health Research Center) and Université Lyon 1 (UMR 5123).

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