

One Universal Common Endpoint in Mouse Models of Amyotrophic Lateral Sclerosis

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Abstract

There is no consensus among research laboratories around the world on the criteria that define endpoint in studies involving rodent models of amyotrophic lateral sclerosis (ALS). Data from 4 nutrition intervention studies using 162 G93A mice, a model of ALS, were analyzed to determine if differences exist between the following endpoint criteria: CS 4 (functional paralysis of both hindlimbs), CS 4+ (CS 4 in addition to the earliest age of body weight loss, body condition deterioration or righting reflex), and CS 5 (CS 4 plus righting reflex >20 s). The age (d; mean \pm SD) at which mice reached endpoint was recorded as the unit of measurement. Mice reached CS 4 at 123.9 \pm 10.3 d, CS 4+ at 126.6 \pm 9.8 d and CS 5 at 127.6 \pm 9.8 d, all significantly different from each other ($P < 0.001$). There was a significant positive correlation between CS 4 and CS 5 ($r = 0.95$, $P < 0.001$), CS 4 and CS 4+ ($r = 0.96$, $P < 0.001$), and CS 4+ and CS 5 ($r = 0.98$, $P < 0.001$), with the Bland-Altman plot showing an acceptable bias between all endpoints. Logrank tests showed that mice reached CS 4 24% and 34% faster than CS 4+ ($P = 0.046$) and CS 5 ($P = 0.006$), respectively. Adopting CS 4 as endpoint would spare a mouse an average of 4 days ($P < 0.001$) from further neuromuscular disability and poor quality of life compared to CS 5. Alternatively, CS 5 provides information regarding proprioception and severe motor neuron death, both could be important parameters in establishing the efficacy of specific treatments. Converging ethics and discovery, would adopting CS 4 as endpoint compromise the acquisition of insight about the effects of interventions in animal models of ALS?

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Introduction

The endpoints used in research studies involving mouse models of amyotrophic lateral sclerosis (ALS) vary widely between laboratories around the world [1–97]. Not all endpoints are in perfect agreement with each other; that is, depending on the endpoint, the age at which mice are euthanized (time point at which age is used to establish lifespan) may differ independently of the intervention treatment used in research studies (Table 1). Because specific endpoints inherently occur early or later in disease progression, researchers may not be able to directly compare the effectiveness of treatments on disease severity and progression, as well as functional outcomes, in animal models of ALS conducted in one research laboratory with similar treatments conducted in different laboratories. Identifying the proper endpoint is important, because it impacts lifespan and marks the end of data collection. A solution would be a universal endpoint which would allow researchers to follow disease progression and identify the effectiveness of an intervention treatment, while still meeting stringent ethical standards. Of relevance, this impacts the adoption of results from animal-based research for human randomized clinical trials, as well as the implications of adopting new recommendations, nutrition or pharmaceutical, for people with ALS.

ALS is a devastating neuromuscular disease characterized by death of motor neurons in the brain [98] and spinal cord [99].

Symptoms of ALS begin with muscle weakness, ultimately leading to paralysis and death [100]. The first mouse model used to study ALS was created by Gurney et al in 1994 [100] who discovered that a glycine to alanine substitution on the 93rd position in the human Cu,Zn superoxide dismutase (Cu/Zn-SOD) gene produced the phenotype of ALS. Mice testing positive for this mutation begin to overtly exhibit signs of motor degeneration through a change in gait between 85–110 d of life [15,82,83]. As the disease progresses, the hindlimbs become paralyzed, paw grip strength and endurance deteriorate, bony structures become palpable due to severe muscle and tissue loss, mobility is limited, and an inability to groom and scavenge for food and water become apparent [20,49,54,73,82]. Researchers conducting intervention studies in mouse models of ALS monitor the above changes to track the effectiveness of their intervention, however at what point is it no longer ethical to keep these mice alive?

Research ethics committees and animal care organizations/agencies serve to maintain standards for the care and use of animals used in research, including transgenic mice used in models of ALS [101]. Standards of care include the selection of “endpoint” which is the point at which an experimental animal is killed humanely to terminate pain, distress and/or suffering [101]. Hence, research ethics committees and animal care organizations/agencies must consult with ALS researchers to decide on a case-by-case basis which endpoint is suitable for a specific intervention study, while

Table 1. Raw data and summary of endpoint criteria for 162 B6SJL-TgN-(SOD1-G93A)1Gur autosomal hemizygous female (F) and male (M) mice.

Endpoint Criteria	N Total (F, M)	% of Mice Meeting Criteria (F, M)	Mean Age (d) (F, M)
CS 4	162 (100, 62)	100.0% (100.0%, 100.0%)	123.9±10.3* (125.8±10.7, 120.8±8.8)
20%CS2	47 (28, 19)	29.0% (28.0%, 30.6%)	128.1±10.7 (129.5±12.1, 125.9±8.5)
20%Peak	69 (40, 29)	42.6% (40.0%, 46.8%)	126.6±10.8 (128.9±11.6, 123.6±8.7)
BC<2	42 (16, 26)	25.9% (16.0%, 41.9%)	121.6±11.5 (127.8±8.3, 117.7±11.7)
CS 4+	162 (100, 62)	100.0% (100.0%, 100.0%)	126.6±9.8* (129.1±9.9, 122.5±8.3)
CS 5	162 (100, 62)	100.0% (100.0%, 100.0%)	127.6±9.8* (129.7±10.0, 124.1±8.5)

CS 4, clinical score of 4 = functional paralysis of both hindlimbs; 20%CS2, weight loss \geq 20% vs. body weight immediately prior to a clinical score of 2; 20%Peak, weight loss \geq 20% vs. peak body weight; BC<2, body condition score <2; CS 5, clinical score of 5 = CS 4 plus a righting reflex >20 s; CS4+, clinical score of 4+ = CS 4 in addition to the earliest of 20%CS2, 20%Peak, BC<2 or a righting reflex of >20 s.

*Significantly different from each other ($P < 0.001$). Data for Mean Age (d) are presented as means \pm SD.

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maintaining compatibility with the objectives and the integrity of the research project. Different laboratories using mouse models of ALS have chosen varying endpoints, some reflecting more advanced stages of the disease, including the righting reflex (mice are placed on their sides and are euthanized if they cannot right themselves to sternum in 3–30 s, time chosen depends on the laboratory), an inability to splay the hindlimbs due to paralysis, a percentage decrease in motor performance or grip strength from initial values, an inability to obtain food or water, a defined percentage of body weight loss from peak weight, serious eye infection, an inability to self-groom, no spontaneous breathing or movement for a predetermined time with no response to pain, complete hindlimb paralysis, or combinations of two or more of these criteria. The most commonly used endpoint is a righting reflex of at least 3 s [1,4,5,7,10–15,17–23,25,27,28,30–32,36,38–47,49–51,53,57–59,61,62,64,65,68,70–72,74,77,79–83,84,86–89,91,94–97], however some studies did not specify the length of time used as the cutoff for the righting reflex [26,54,60,73,78,85,90,93]. The popularity of the righting reflex is possibly due to its relative simplicity, value as an indicator of proprioception deterioration [102,103], and/or history as the first endpoint used in an intervention study in this particular disease model [79].

To date, a universal endpoint has not been established among researchers using rodent models of ALS. An ideal endpoint would meet strict ethical standards, could be adopted by all research laboratories, and would allow researchers to properly study the progression of ALS and the effectiveness of treatments tested. A consistent endpoint across research laboratories would reduce inter-laboratory variability that may be attributed at least partially to the selection of endpoint. Thus, our objective was to determine whether an earlier endpoint could replace the righting reflex, sparing mice undue suffering, while preserving the integrity of research in rodent models of ALS. To do this, we used the G93A transgenic mouse model of ALS to validate if functional paralysis in both hindlimbs (CS 4) could replace other later endpoints, including the righting reflex (CS 5).

Materials and Methods

Ethics Statement

The experimental protocols of all 4 studies followed the guidelines of the Canadian Council of Animal Care and were approved by the McMaster University Animal Research Ethics Board. All necessary steps were taken to minimize suffering and distress to the mice in the studies.

Animals

Raw data for clinical score (CS), body condition and body weight were compiled from 4 previously published [15,82,83,104,105] nutrition intervention studies using a total of 162 B6SJL-TgN-(SOD1-G93A)1Gur autosomal hemizygous mice (100 females, 62 males) that reached endpoint at a clinical score of 5 (CS 5). All mice expressed the phenotype of ALS due to the G93A mutation in the SOD1 (Cu/Zn-SOD) gene. Raw data were used to determine the following endpoint criteria, with age (d) at which mice reached endpoint as the unit of measurement:

- 1) CS 4 = both hindlimbs are functionally paralyzed
- 2) CS 4+ = CS 4 plus the earliest time mice attained one of the following:
 - a) weight loss \geq 20% vs. body weight immediately prior to a clinical score of 2 (CS 2 is considered disease onset) = 20%CS2
 - b) weight loss \geq 20% vs. peak body weight = 20%Peak
 - c) body condition score <2 = BC<2
 - d) righting reflex >20 s (clinical score of 5) = CS 5
- 3) CS 5 = CS 4 plus a righting reflex >20 s (considered as the endpoint in the previous 4 studies)

Body Weight and Body Condition

Body weights of mice in the 4 intervention studies were measured starting at age 35–40 d until mice reached CS 5. Body condition was assessed following a 5-point scale: 5 = obese mice, 4 = overconditioned mice (spine is a continuous column and the vertebrae are palpable only with firm pressure), 3 = well-conditioned mice (the vertebrae and dorsal pelvis are not prominent and are palpable with slight pressure), 2 = underconditioned mice (the segmentation of the vertebral column is evident and the dorsal pelvic bones are easily palpable), and 1 = emaciated mice (the skeletal structure is extremely prominent and the vertebrae are distinctly segmented). Body condition was recorded starting at age 43–79 d until mice reached CS 5.

Clinical Score

Using an 8-point scale, clinical score measurements for mice in the 4 intervention studies started at age 50–81 d until mice reached CS 5. The clinical score was based on signs exhibited by the mice to identify the severity of the disease: 0 = no evidence of

disease, 1 = shaking or splaying of the hindlimbs when suspended by the tail (an indication of weakness in the hindlimbs), 1.5 = weakness in one hindlimb (compensation for footdrop), 2 = change in gait (used as disease onset when attained on two consecutive days), 2.5 = extreme weakness in one hindlimb (inability to dorsiflex), 3 = extreme weakness in both hindlimbs, 3.5 = functional paralysis in one hindlimb, 4 = functional paralysis in both hindlimbs but can right themselves in less than 20 s after being placed on their side, and 5 = cannot right themselves to sternum within 20 s after being placed on their sides (endpoint).

Statistical Analysis

Data for all 162 mice were submitted to a one-way repeated measures ANOVA to determine significant differences between CS4, CS4+ and CS5. When ANOVA indicated significance, a Tukey's HSD post hoc was used to determine the source of difference. A Pearson product-moment correlation coefficient (r) was determined to establish the relationship between the different endpoints. A Bland-Altman plot was used to analyze the agreement between the different endpoints. A logrank test was used to determine whether there was a difference in the rate at which mice reached CS4, CS4+ and CS 5. For all logrank tests, CS 4 was used as the reference when comparing CS 4 vs. CS 4+ and CS 4 vs. CS 5, whereas CS 4+ was used as the reference when comparing CS 4+ vs. CS 5. All ANOVA, linear regression and logrank test comparisons were planned. All statistical analyses were completed using GraphPad Prism (version 4.0, GraphPad Software, La Jolla, CA). Significance was established at $P \leq 0.05$. Data are presented as means \pm SD, unless otherwise indicated.

Results

Mice reached CS 4 at 123.9 ± 10.3 d, CS 4+ at 126.6 ± 9.8 d and CS 5 at 127.6 ± 9.8 d (Table 1). There was a significant main effect between endpoints ($P < 0.001$), all being significantly different from each other ($P < 0.001$ for all).

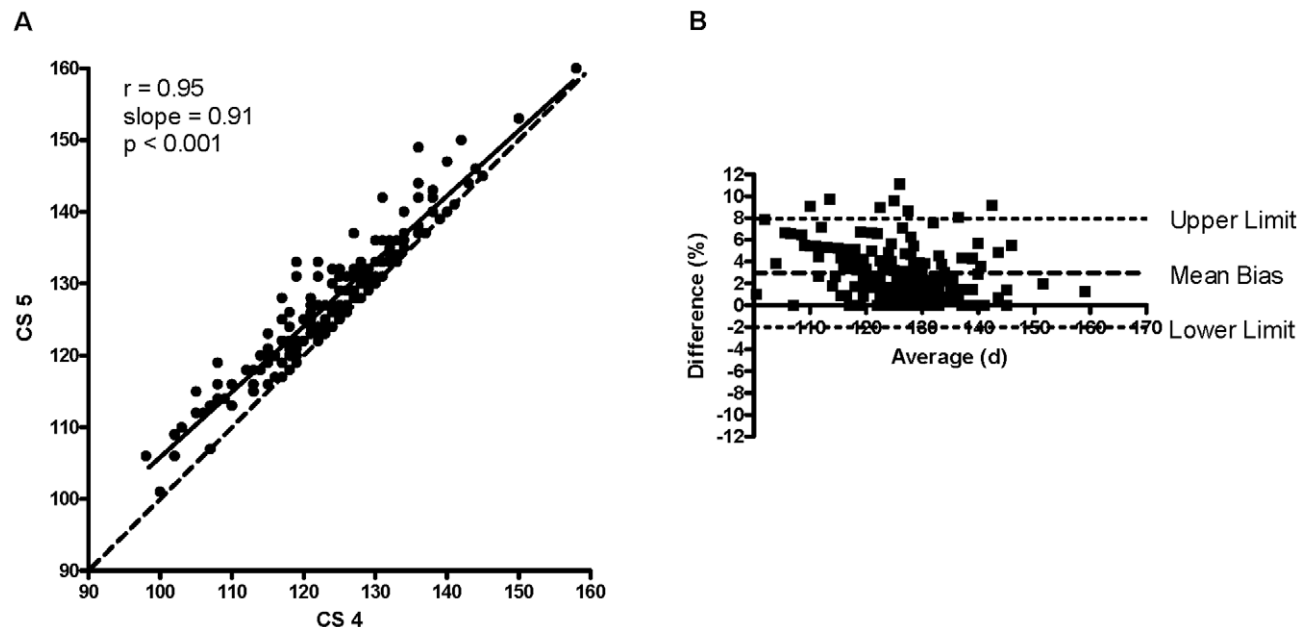


Figure 1. Correlation between CS 4 and CS 5 and the Bland-Altman plot for CS 4 vs. CS 5. (A) Correlation between CS 4 (clinical score of 4 = functional paralysis of both hindlimbs) and CS 5 (clinical score of 5 = CS 4 plus a righting reflex > 20 s). There was a strong positive relationship between CS 4 and CS 5 ($r = 0.95$, slope = 0.91, $P < 0.001$). $CS\ 5\ (d) = (14.80 \pm 2.83) + [(0.91 \pm 0.02) \times (CS\ 4\ in\ d)]$, mean \pm SEM. Dashed line indicates line of identity. (B) A Bland-Altman plot comparing CS 4 to CS 5. Mean bias \pm SD = $3.0 \pm 2.5\%$, lower limit = -2.0% , upper limit = 7.9% . doi:10.1371/journal.pone.0020582.g001

There was a strong positive correlation between CS 4 and CS 5 ($r = 0.95$; slope = 0.91; $P < 0.001$; Figure 1A), CS 4 and CS 4+ ($r = 0.96$; slope = 0.92; $P < 0.001$; Figure 2A), and CS 4+ and CS 5 ($r = 0.98$; slope = 0.98; $P < 0.001$; Figure 3A). The Bland-Altman plot revealed acceptable bias between CS 4 and CS 5 ($3.0 \pm 2.5\%$; lower limit = -2.0% , upper limit = 7.9% ; Figure 1B), between CS 4 and CS 4+ ($2.2 \pm 2.3\%$; lower limit = -2.4% , upper limit = 6.7% ; Figure 2B), and between CS 4+ and CS 5 ($0.8 \pm 1.7\%$; lower limit = -2.5% , upper limit = 4.1% ; Figure 3B).

A logrank test showed a significant difference in the rate at which endpoint was reached between CS 4, CS 4+ and CS 5 ($P = 0.021$; Figure 4). Mice reached CS 4 at a rate 34% faster *vs.* CS 5 (HR = 1.34; 95% CI 1.10, 1.74; $P = 0.006$) and 24% faster *vs.* CS 4+ (HR = 1.24; 95% CI 1.00, 1.59; $P = 0.046$). Mice reached CS 4+ at a non-significant rate of 9% faster *vs.* CS 5 (HR = 1.09; 95% CI 0.88, 1.38; $P = 0.410$).

Statistical analyses were conducted for the same 3 endpoints within each sex. Differences between endpoints within each sex were similar as above.

Discussion

Our objective was to determine whether an earlier endpoint could replace the most commonly used righting reflex in a transgenic mouse model of ALS. This was done to validate the use of an endpoint that would meet the strict standards set by research ethics boards to decrease suffering and distress in mice, as well as to allow researchers from different laboratories the use of a uniform and consistent endpoint to directly compare the effectiveness of treatments in this particular animal model. We found strong positive correlations between all endpoints with an acceptable mean bias as measured by a Bland-Altman plot. Using CS 4+ and CS 5 would prolong life span by 2% and 3%, respectively, as compared to CS 4. Additionally, mice reached CS 4 at a rate 24% faster compared to CS 4+ and 34% faster compared to CS 5.

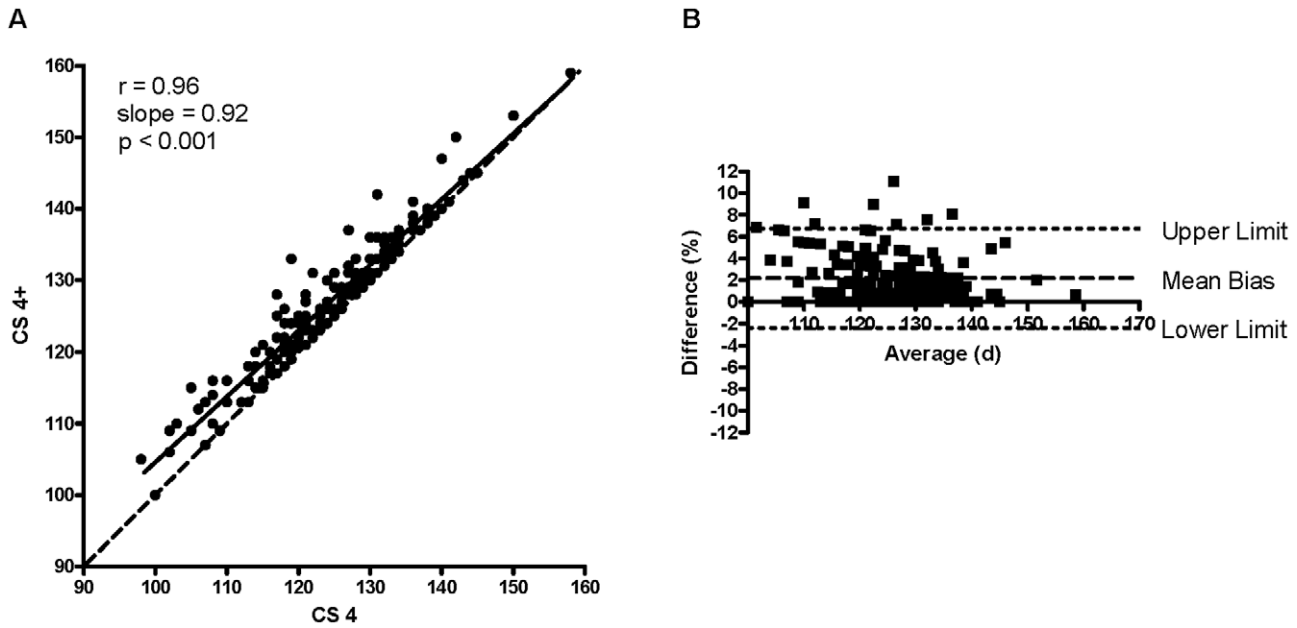


Figure 2. Correlation between CS 4 and CS 4+ and the Bland-Altman plot for CS 4 and CS 4+. (A) Correlation between CS 4 (clinical score of 4 = functional paralysis of both hindlimbs) and CS 4+ [CS 4 plus the earliest of a) weight loss $\geq 20\%$ vs. body weight immediately prior to a clinical score of 2, b) weight loss $\geq 20\%$ vs. peak body weight, c) body condition score < 2 , or d) a righting reflex > 20 s (CS 5)]. There was a strong positive relationship between CS 4 and CS 4+ ($r = 0.96$, slope = 0.92, $P < 0.001$). CS 4+ (d) = $(12.79 \pm 2.56) + [(0.92 \pm 0.02) \times (\text{CS 4 in d})]$, mean \pm SEM. Dashed line indicates line of identity. (B) A Bland-Altman plot comparing CS 4 to CS 4+. Mean bias \pm SD = $2.2 \pm 2.3\%$, lower limit = -2.4% , upper limit = 6.7% . doi:10.1371/journal.pone.0020582.g002

Once mice reach CS 4, they must rely on the strength of their forelimbs to obtain food and water which may place them at risk of starvation and dehydration [43]. Some studies have used the inability to scrounge for food and water as endpoint [11,13,28,

43,63,67,73,106], however establishing this is time consuming and indicates an advanced disease state possibly well beyond CS 5. As well, research ethics committees may institute policy requiring mice to have access to food and water-based gels at cage floor level

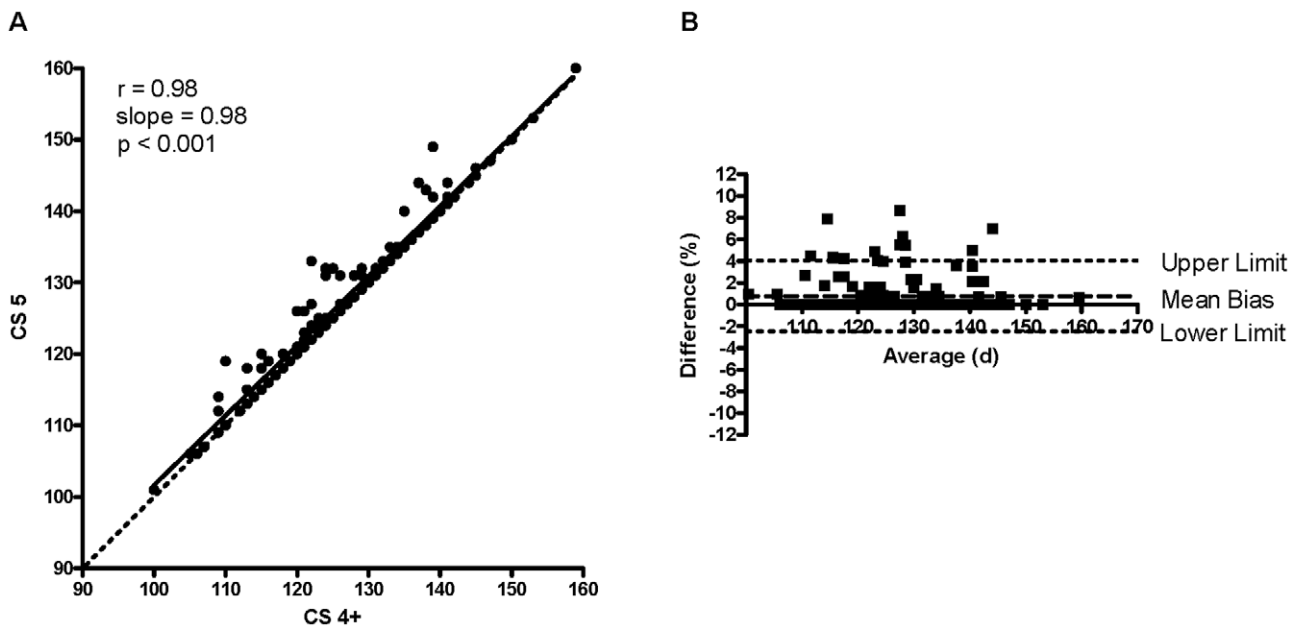


Figure 3. Correlation between CS 5 and CS 4+ and the Bland-Altman plot for CS 5 and CS 4+. (A) Correlation between CS 5 (clinical score of 5 = CS 4 and righting reflex > 20 s) and CS 4+ [CS 4 plus the earliest of a) weight loss $\geq 20\%$ vs. body weight immediately prior to a clinical score of 2, b) weight loss $\geq 20\%$ loss vs. peak body weight, c) body condition score < 2 , or d) a righting reflex > 20 s (CS 5)]. There was a strong positive relationship between CS 5 and CS 4+ ($r = 0.98$, slope = 0.98, $P < 0.001$). CS 5 (d) = $(3.93 \pm 2.15) + [(0.98 \pm 0.02) \times (\text{CS 4+ in d})]$, mean \pm SEM. Dashed line indicates line of identity. (B) A Bland-Altman plot comparing CS 5 to CS 4+. Mean bias \pm SD = $0.8 \pm 1.7\%$, lower limit = -2.5% , upper limit = 4.1% . doi:10.1371/journal.pone.0020582.g003

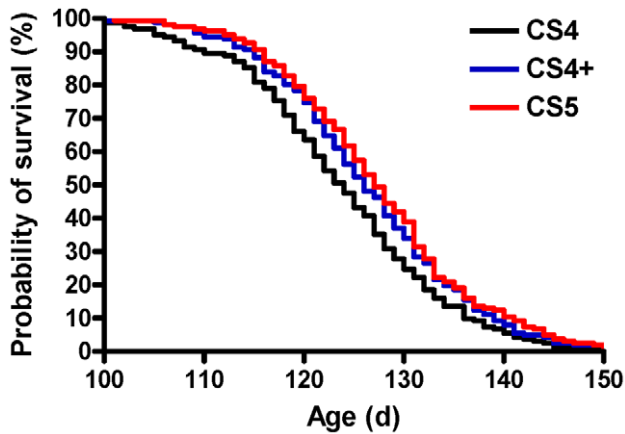


Figure 4. Probability of survival for CS 4, CS 4+ and CS 5. Probability of survival for the 3 different endpoints (CS 4, black line; CS 4+, blue line; CS 5, red line). For all logrank tests, CS 4 was used as the reference when comparing CS 4 vs. CS 4+ and CS 4 vs. CS 5, whereas CS 4+ was used as the reference when comparing CS 4+ vs. CS 5. The rate of reaching endpoint is significantly different ($P=0.021$) between CS 4 (clinical score of 4 = functional paralysis of both hindlimbs), CS 4+ [CS 4 plus the earliest of a) weight loss $\geq 20\%$ vs. body weight immediately prior to a clinical score of 2, b) weight loss $\geq 20\%$ vs. peak body weight, c) body condition score < 2 , or d) a righting reflex > 20 s (CS 5)], and CS 5 (clinical score of 5 = CS 4 and righting reflex > 20 s). Mice reached CS 4 at a rate of 34% faster vs. CS 5 (HR = 1.34; 95% CI 1.10, 1.74; $P=0.006$) and 24% faster vs. CS 4+ (HR = 1.24; 95% CI 1.00, 1.59; $P=0.046$). Mice reached CS 4+ at a non-significant rate of 9% faster vs. CS 5 (HR = 1.09; 95% CI 0.88, 1.38; $P=0.410$). doi:10.1371/journal.pone.0020582.g004

when mice reach a pre-defined disease severity, which actually prolongs disease exposure of mice due to easier access to nutrients.

Paw grip endurance and motor performance scores will have decreased precipitously by the time mice reached CS 4, as compared to scores prior to disease onset, due to hindlimb paralysis and weakness in the forelimbs [15,82,83]. A decrease in motor performance and/or paw grip strength has been previously used as endpoint in mouse models of ALS [3,20]. Adoption of such endpoints requires expensive, specialized equipment such as the rotarod apparatus [20] and commercial grip strength meters [3]. Also, there is no standardization for the percent decrease in paw grip strength among laboratories using this as a criterion for endpoint [3,20].

All mice in our analyses met the criteria for CS 4+, however when each additional criterion was assessed in isolation from CS 4, that is, as standalone criterion for endpoint (data collection ended at CS 5), only 29% of mice lost greater than 20% body weight

versus their weight immediately prior to disease onset (20%CS2), 43% lost greater than 20% body weight versus peak weight (20%Peak), 26% had a body condition score of less than 2 ($BC < 2$), while 100% met the criteria for CS 5 (Table 1). These results suggest that studies using any one of 20%CS2, 20%Peak, or $BC < 2$ as a standalone criterion to establish endpoint would be keeping at least 57% of their mice alive past CS 5, prolonging disease exposure beyond what is considered humane. Alternatively, another interpretation of these results is that mice could die past CS 5 without meeting the standalone criteria 20%CS2, 20%Peak, or $BC < 2$. Past CS 5, motor neuron degeneration is so far advanced that mice can no longer right themselves to scrounge for food and water and would be at a pronounced risk of starvation and dehydration. Our analyses also reveal that fewer than 9% of all mice met the standalone criteria 20%CS2, 20%Peak, or $BC < 2$ prior to reaching CS 4 (Table 2). Hence, we conclude that CS 4 does not prolong disease exposure compared to 20%CS2, 20%Peak, or $BC < 2$ in a mouse model of ALS. More male mice met 20%CS2, 20%Peak, or $BC < 2$ prior to reaching CS 4 compared to females, however this result is expected since male mice have greater muscle mass than females and muscle atrophy is a result of disease progression [107].

The righting reflex, either as a standalone criterion or used in conjunction with other parameters, has long been used to establish endpoint in a mouse model of ALS [1,4,5,7,10–15,17–23,25–28,30–32,36,38–47,49–51,53,54,57–62,64,65,68,70–74,77–91,93–97]. The righting reflex has its advantages. Failure to right within a pre-defined period of time (at least > 3 s) demonstrates severe muscle weakness, an indication of advanced motor neuron degeneration, as mice must use their strength to right themselves when placed on their side. The righting reflex may also be a measure of declining proprioception. Evidence suggests the dorsal root [102], dorsal root ganglia [102,108–110], dorsal funiculus [102], Clarke’s nuclei [103,109,111,112] and spinocerebellar tract [103,109,111–113], the regions of the spinal cord responsible for processing proprioception, may be affected in humans with ALS [103,108–112] and animal models of ALS [102,113], however some researchers failed to ascertain this association [113]. It is important to note that the magnitude of diminished proprioception and muscle loss may be different depending on the time used as the cutoff for the righting reflex, with greater atrophy of motor neurons occurring when longer cutoffs are used [113]. Although the righting reflex may provide insight into muscle wasting and proprioception deficits, no studies have used the righting reflex to quantify proprioception and motor neuron loss. Rather, the righting reflex is simply used to identify endpoint. Moreover, the time used to establish the righting reflex is not standardized (at least 3 s), introducing a confounding within the righting reflex methodology. Some studies did not specify the length

Table 2. Raw data and summary of 162 B6SJL-TgN-(SOD1-G93A)1Gur autosomal hemizygous female (F) and male (M) mice meeting the additional endpoint criteria prior to reaching CS 4.

Endpoint Criteria	N Prior to CS 4 (F, M)	% of Mice Meeting Criteria Prior to CS 4 (F, M)	Mean Age (d) (F, M)
20%CS2	1 (0, 1)	0.6% (0%, 1.6%)	106.0 (NA, 106.0)
20%Peak	7 (3, 4)	4.3% (3%, 6.5%)	119.9 ± 11.0 (123.7 ± 15.2, 117.0 ± 8.0)
$BC < 2$	9 (1, 8)	5.6% (1%, 12.9%)	110.6 ± 15.7 (108.0, 110.9 ± 16.8)
Any of 20%CS2, 20%Peak or $BC < 2$ *	14 (3, 11)	8.6% (3%, 17.7%)	115.6 ± 15.0 (123.0 ± 16.1, 113.6 ± 14.8)

20%CS2, weight loss $\geq 20\%$ vs. body weight immediately prior to a clinical score of 2; 20%Peak, weight loss $\geq 20\%$ vs. peak body weight; $BC < 2$, body condition score < 2 .

*Earliest age (d) of 20%CS2, 20%Peak and $BC < 2$ was used to calculate mean age. Data for Mean Age (d) are presented as means ± SD.

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of time used as the cutoff for the righting reflex [26,54,60,73, 78,85,90,93].

The criteria for the ideal endpoint would meet strict ethical standards, be easily adopted by research laboratories, and ensure researchers are able to gather information regarding the progression of ALS and the effectiveness of treatments in intervention studies. Also, it would represent a point in the progression of the disease beyond which additional insight into the nature of disease or into the effectiveness of an intervention is absent, or at least nominal. CS 4, representing functional paralysis in both hindlimbs, occurs in all mice used in mouse models of ALS. CS 4 is reached earlier than both CS 4+ and CS 5, satisfying standards set by research ethics committees by shortening the time of disease exposure. A more difficult challenge arises when addressing the final criterion required to establish an ideal endpoint, that is, does CS 4 permit investigators the acquisition of sufficient data relating to disease progression? In rodent models of ALS, functional paralysis marks the beginning of the end in disease progression. Once paralysis is established in the hindlimbs, it will spread to the diaphragm, ultimately resulting in death due to respiratory failure. Between disease onset and hindlimb paralysis, changes in gait, paw grip strength and endurance, and motor performance deteriorate measurably allowing scientists to track these changes throughout the course of the disease [15,82,83]. These changes continue to occur past CS 4, but in severely disabled mice with compromised quality of life. Our analysis has yielded an equation that will allow researchers to predict the age at which CS 4+ and/or CS 5 are attained, on average, from CS 4. Animal models of multiple sclerosis (experimental autoimmune encephalomyelitis; EAE) [114–116] follow a similar disease progression, including hindlimb

weakness and paralysis, and use similar endpoint criteria as mouse models of ALS, suggesting our findings may be used in mouse models of EAE.

Is CS 4 the “ideal” endpoint? The righting reflex may provide information regarding muscle loss and proprioception deficits beyond that of CS 4. For those specific studies whereby severe muscle loss and compromised proprioception are inherent outcome measures reflecting the effectiveness of a specific intervention, CS 5 should be adopted as an endpoint. Alternatively, we have shown that CS 4, occurring on average 4 days sooner than CS 5, can predict the age at CS 4+ or CS 5. These 4 days will lessen the suffering and distress experienced by mice used in mouse models of ALS. Adopting CS 4 as endpoint negotiates an acceptable agreement between scientific discovery and ethics, a partnership that serves to protect scientific integrity and ethical standards in the humane treatment of research animals. At the forefront is the strength of the data extrapolated from animal-based research to serve as the background for potential recommendations adopted for people with ALS.

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Author Contributions

Conceived and designed the experiments: MJH. Performed the experiments: MJH. Analyzed the data: JAS MJH. Contributed reagents/materials/analysis tools: MAT MJH. Wrote the paper: JAS MJH.

References

- Mahoney DJ, Rodriguez C, Devries M, Yasuda N, Tarnopolsky MA (2004) Effects of high-intensity endurance exercise training in the G93A mouse model of amyotrophic lateral sclerosis. *Muscle Nerve* 29: 656–662.
- Lee J, Ryu H, Kowall NW (2009) Motor neuronal protection by L-arginine prolongs survival of mutant SOD1 (G93A) ALS mice. *Biochem Biophys Res Commun* 384: 524–529.
- Liebetanz D, Hagemann K, von Lewinski F, Kahler E, Paulus W (2004) Extensive exercise is not harmful in amyotrophic lateral sclerosis. *Eur J Neurosci* 20: 3115–20.
- Martinez JA, Francis GJ, Liu WQ, Pradzinsky N, Fine J, et al. (2008) Intranasal delivery of insulin and a nitric oxide synthase inhibitor in an experimental model of amyotrophic lateral sclerosis. *Neuroscience* 157: 908–925.
- Matthews RT, Yang L, Browne S, Baik M, Beal MF (1998) Coenzyme Q10 administration increases brain mitochondrial concentrations and exerts neuroprotective effects. *Proc Natl Acad Sci U S A* 95: 8892–8897.
- Mattson MP, Cutler RG, Camandola S (2007) Energy intake and amyotrophic lateral sclerosis. *Neuromolecular Med* 9: 17–20.
- Moges H, Vasconcelos OM, Campbell WW, Borke RC, McCoy JA, et al. (2009) Light therapy and supplementary Riboflavin in the SOD1 transgenic mouse model of familial amyotrophic lateral sclerosis (FALS). *Lasers Surg Med* 41: 52–59.
- Nagano S, Fujii Y, Yamamoto T, Taniyama M, Fukada K, et al. (2003) The efficacy of trientine or ascorbate alone compared to that of the combined treatment with these two agents in familial amyotrophic lateral sclerosis model mice. *Exp Neurol* 179: 176–180.
- Nagano S, Ogawa Y, Yanagihara T, Sakoda S (1999) Benefit of a combined treatment with trientine and ascorbate in familial amyotrophic lateral sclerosis model mice. *Neurosci Lett* 265: 159–162.
- Neymotin A, Petri S, Calingasan NY, Wille E, Schafer P, et al. (2009) Lenalidomide (Revlimid) administration at symptom onset is neuroprotective in a mouse model of amyotrophic lateral sclerosis. *Exp Neurol* 220: 191–197.
- Ohnishi S, Ito H, Suzuki Y, Adachi Y, Wate R, et al. (2009) Intra-bone marrow-bone marrow transplantation slows disease progression and prolongs survival in G93A mutant SOD1 transgenic mice, an animal model mouse for amyotrophic lateral sclerosis. *Brain Res* 1296: 216–224.
- Ohta Y, Kamiya T, Nagai M, Nagata T, Morimoto N, et al. (2008) Therapeutic benefits of intrathecal protein therapy in a mouse model of amyotrophic lateral sclerosis. *J Neurosci Res* 86: 3028–3037.
- Pamphlett R, Todd E, Vink R, McQuilty R, Cheema SS (2003) Magnesium supplementation does not delay disease onset or increase survival in a mouse model of familial ALS. *J Neurol Sci* 216: 95–98.
- Park JH, Hong YH, Kim HJ, Kim SM, Kim MJ, et al. (2007) Pyruvate slows disease progression in a G93A SOD1 mutant transgenic mouse model. *Neurosci Lett* 413: 265–269.
- Patel BP, Safdar A, Raha S, Tarnopolsky MA, Hamadeh MJ (2010) Caloric restriction shortens lifespan through an increase in lipid peroxidation, inflammation and apoptosis in the G93A mouse, an animal model of ALS. *PLoS One* Feb 24;5(2): e9386.
- Pedersen WA, Mattson MP (1999) No benefit of dietary restriction on disease onset or progression in amyotrophic lateral sclerosis Cu/Zn-superoxide dismutase mutant mice. *Brain Res* 833: 117–120.
- Petri S, Kiaei M, Wille E, Calingasan NY, Flint Beal M (2006) Loss of Fas ligand-function improves survival in G93A-transgenic ALS mice. *J Neurol Sci* 251: 44–49.
- Petri S, Calingasan NY, Alsaied OA, Wille E, Kiaei M, et al. (2007) The lipophilic metal chelators DP-109 and DP-460 are neuroprotective in a transgenic mouse model of amyotrophic lateral sclerosis. *J Neurochem* 102: 991–1000.
- Petri S, Kiaei M, Kipiani K, Chen J, Calingasan NY, et al. (2006) Additive neuroprotective effects of a histone deacetylase inhibitor and a catalytic antioxidant in a transgenic mouse model of amyotrophic lateral sclerosis. *Neurobiol Dis* 22: 40–49.
- Pitzer C, Kruger C, Plaas C, Kirsch F, Dittgen T, et al. (2008) Granulocyte-colony stimulating factor improves outcome in a mouse model of amyotrophic lateral sclerosis. *Brain* 131: 3335–3347.
- Pizzasegola C, Caron I, Daleno C, Ronchi A, Minoia C, et al. (2009) Treatment with lithium carbonate does not improve disease progression in two different strains of SOD1 mutant mice. *Amyotroph Lateral Scler* 10: 221–228.
- Poduslo JF, Whelan SL, Curran GL, Wengenack TM (2000) Therapeutic benefit of polyamine-modified catalase as a scavenger of hydrogen peroxide and nitric oxide in familial amyotrophic lateral sclerosis transgenics. *Ann Neurol* 48: 943–947.
- Reinholz MM, Merkle CM, Poduslo JF (1999) Therapeutic benefits of putrescine-modified catalase in a transgenic mouse model of familial amyotrophic lateral sclerosis. *Exp Neurol* 159: 204–216.
- Rembach A, Turner BJ, Bruce S, Cheah IK, Scott RL, et al. (2004) Antisense peptide nucleic acid targeting GluR3 delays disease onset and progression in the SOD1 G93A mouse model of familial ALS. *J Neurosci Res* 77: 573–582.
- Ryu H, Smith K, Camelo SI, Carreras I, Lee J, et al. (2005) Sodium phenylbutyrate prolongs survival and regulates expression of anti-apoptotic genes in transgenic amyotrophic lateral sclerosis mice. [Erratum appears in *J Neurochem*. 2006 Feb;96(3):908]. *J Neurochem* 93: 1087–1098.

26. Sekiya M, Ichihyanagi T, Ikeshiro Y, Yokozawa T (2009) The Chinese prescription Wen-Pi-Tang extract delays disease onset in amyotrophic lateral sclerosis model mice while attenuating the activation of glial cells in the spinal cord. *Biol Pharm Bull* 32: 382–388.
27. Shimojo Y, Kosaka K, Noda Y, Shimizu T, Shirasawa T (2010) Effect of rosmarinic acid in motor dysfunction and life span in a mouse model of familial amyotrophic lateral sclerosis. *J Neurosci Res* 88: 896–904.
28. Shoemaker JL, Seely KA, Reed RL, Crow JP, Prather PL (2007) The CB2 cannabinoid agonist AM-1241 prolongs survival in a transgenic mouse model of amyotrophic lateral sclerosis when initiated at symptom onset. *J Neurochem* 101: 87–98.
29. Snow RJ, Turnbull J, da Silva S, Jiang F, Tarnopolsky MA (2003) Creatine supplementation and riluzole treatment provide similar beneficial effects in copper, zinc superoxide dismutase (G93A) transgenic mice. *Neuroscience* 119: 661–667.
30. Suchy J, Lee S, Ahmed A, Shea TB (2010) Dietary supplementation with S-adenosyl methionine delays the onset of motor neuron pathology in a murine model of amyotrophic lateral sclerosis. *Neuromolecular Med* 12: 86–97.
31. Teng YD, Choi H, Huang W, Onario RC, Frontera WR, et al. (2006) Therapeutic effects of clenbuterol in a murine model of amyotrophic lateral sclerosis. *Neurosci Lett* 397: 155–158.
32. Tokuda E, Ono S, Ishige K, Watanabe S, Okawa E, et al. (2008) Ammonium tetrathiomolybdate delays onset, prolongs survival, and slows progression of disease in a mouse model for amyotrophic lateral sclerosis. *Exp Neurol* 213: 122–128.
33. Turner BJ, Parkinson NJ, Davies KE, Talbot K (2009) Survival motor neuron deficiency enhances progression in an amyotrophic lateral sclerosis mouse model. *Neurobiol Dis* 34: 511–517.
34. Turner BJ, Rembach A, Spark R, Lopes EC, Cheema SS (2003) Opposing effects of low and high-dose clonidine on survival of transgenic amyotrophic lateral sclerosis mice. *J Neurosci Res* 74: 605–613.
35. Turner BJ, Murray SS, Piccenna LG, Lopes EC, Kilpatrick TJ, et al. (2004) Effect of p75 neurotrophin receptor antagonist on disease progression in transgenic amyotrophic lateral sclerosis mice. *J Neurosci Res* 78: 193–199.
36. Van Damme P, Leyssen M, Callewaert G, Robberecht W, Van Den Bosch L (2003) The AMPA receptor antagonist NBQX prolongs survival in a transgenic mouse model of amyotrophic lateral sclerosis. *Neurosci Lett* 343: 81–84.
37. Veldink JH, Bar PR, Joosten EAJ, Otten M, Wokke JHJ, et al. (2003) Sexual differences in onset of disease and response to exercise in a transgenic model of ALS. *Neuromuscul Disord* 13: 737–743.
38. Vercelli A, Mereuta OM, Garbossa D, Muraca G, Mareschi K, et al. (2008) Human mesenchymal stem cell transplantation extends survival, improves motor performance and decreases neuroinflammation in mouse model of amyotrophic lateral sclerosis. *Neurobiol Dis* 31: 395–405.
39. Waibel S, Reuter A, Malessa S, Blaugrund E, Ludolph AC (2004) Rasagiline alone and in combination with riluzole prolongs survival in an ALS mouse model. *J Neurol* 251: 1080–1084.
40. Wang R, Zhang D (2005) Memantine prolongs survival in an amyotrophic lateral sclerosis mouse model. *Eur J Neurosci* 22: 2376–2380.
41. Weishaupt JH, Bartels C, Polking E, Dietrich J, Rohde G, et al. (2006) Reduced oxidative damage in ALS by high-dose enteral melatonin treatment. *J Pineal Res* 41: 313–323.
42. West M, Mhatre M, Ceballos A, Floyd RA, Grammas P, et al. (2004) The arachidonic acid 5-lipoxygenase inhibitor nordihydroguaiaretic acid inhibits tumor necrosis factor alpha activation of microglia and extends survival of G93A-SOD1 transgenic mice. *J Neurochem* 91: 133–143.
43. Wu AS, Kiaei M, Aguirre N, Crow JP, Calingasan NY, et al. (2003) Iron porphyrin treatment extends survival in a transgenic animal model of amyotrophic lateral sclerosis. *J Neurochem* 85: 142–150.
44. Xu Z, Chen S, Li X, Luo G, Li L, et al. (2006) Neuroprotective effects of (-)-epigallocatechin-3-gallate in a transgenic mouse model of amyotrophic lateral sclerosis. *Neurochem Res* 31: 1263–1269.
45. Zhang X, Chen S, Li L, Wang Q, Le W (2008) Folic acid protects motor neurons against the increased homocysteine, inflammation and apoptosis in SOD1 G93A transgenic mice. *Neuropharmacology* 54: 1112–1119.
46. Zhao Z, Lange DJ, Voustianiouk A, MacGrogan D, Ho L, et al. (2006) A ketogenic diet as a potential novel therapeutic intervention in amyotrophic lateral sclerosis. *BMC Neurosci* 7: 29.
47. Amante DJ, Kim J, Carreiro ST, Cooper AC, Jones SW, et al. (2010) Uridine ameliorates the pathological phenotype in transgenic G93A-ALS mice. *Amyotroph Lateral Scler* 2010 Jun 22 [Epub ahead of print].
48. Amodio R, Esposito E, De Ruvo C, Bellavia V, Amodio E, et al. (2006) Red wine extract prevents neuronal apoptosis in vitro and reduces mortality of transgenic mice. *Ann N Y Acad Sci* 1089: 88–97.
49. Andreassen OA, Dedeoglu A, Klivenyi P, Beal MF, Bush AI (2000) N-acetyl-L-cysteine improves survival and preserves motor performance in an animal model of familial amyotrophic lateral sclerosis. *Neuroreport* 11: 2491–2493.
50. Andreassen OA, Jenkins BG, Dedeoglu A, Ferrante KL, Bogdanov MB, et al. (2001) Increases in cortical glutamate concentrations in transgenic amyotrophic lateral sclerosis mice are attenuated by creatine supplementation. *J Neurochem* 77: 383–390.
51. Andreassen OA, Dedeoglu A, Friedlich A, Ferrante KL, Hughes D, et al. (2001) Effects of an inhibitor of poly(ADP-ribose) polymerase, desmethylselegiline, trientine, and lipoic acid in transgenic ALS mice. *Exp Neurol* 168: 419–424.
52. Azari MF, Profyris C, Le Grande MR, Lopes EC, Hirst J, et al. (2005) Effects of intraperitoneal injection of Rofecoxib in a mouse model of ALS. *Eur J Neurol* 12: 357–364.
53. Azzouz M, Ralph GS, Storkebaum E, Walmsley LE, Mitrophanous KA, et al. (2004) VEGF delivery with retrogradely transported lentivector prolongs survival in a mouse ALS model. *Nature* 429: 413–417.
54. Barbeito AG, Martinez-Palma L, Vargas MR, Pehar M, Manay N, et al. (2010) Lead exposure stimulates VEGF expression in the spinal cord and extends survival in a mouse model of ALS. *Neurobiol Dis* 37: 574–580.
55. Bruce KM, Narayan K, Kong HC, Larmour I, Lopes EC, et al. (2004) Chemotherapy delays progression of motor neuron disease in the SOD1 G93A transgenic mouse. *Chemotherapy* 50: 138–142.
56. Bruestle DA, Cutler RG, Telljohann RS, Mattson MP (2009) Decline in daily running distance presages disease onset in a mouse model of ALS. *Neuromolecular Med* 11: 58–62.
57. Caraganis A, Benn S, Cudkowicz M, Brown Jr RH (2008) Thrombopoietin is ineffective in a mouse model of motor neuron disease. *Amyotroph Lateral Scler* 9: 354–358.
58. Chiba T, Yamada M, Sasabe J, Terashita K, Aiso S, et al. (2006) Colivelin prolongs survival of an ALS model mouse. *Biochem Biophys Res Commun* 343: 793–798.
59. Choi CI, Lee YD, Gwag BJ, Cho SI, Kim SS, et al. (2008) Effects of estrogen on lifespan and motor functions in female hSOD1 G93A transgenic mice. *J Neurol Sci* 268: 40–47.
60. Chritin M, Savasta M, Besson G (2006) Benefit of tianeptine and morphine in a transgenic model of familial amyotrophic lateral sclerosis. *Amyotroph Lateral Scler* 7: 32–37.
61. Ciriza J, Moreno-Igoa M, Calvo AC, Yague G, Palacios J, et al. (2008) A genetic fusion GDNF-C fragment of tetanus toxin prolongs survival in a symptomatic mouse ALS model. *Restor Neurol Neurosci* 26: 459–465.
62. Corti S, Locatelli F, Papadimitriou D, Del Bo R, Nizzardo M, et al. (2007) Neural stem cells LewisX+ CXCR4+ modify disease progression in an amyotrophic lateral sclerosis model. *Brain* 130: 1289–1305.
63. Crochemore C, Virgili M, Bonamassa B, Canistro D, Pena-Altamira E, et al. (2009) Long-term dietary administration of valproic acid does not affect, while retinoic acid decreases, the lifespan of G93A mice, a model for amyotrophic lateral sclerosis. *Muscle Nerve* 39: 548–552.
64. Crow JP, Calingasan NY, Chen J, Hill JL, Beal MF (2005) Manganese porphyrin given at symptom onset markedly extends survival of ALS mice. *Ann Neurol* 58: 258–265.
65. Del Signore SJ, Amante DJ, Kim J, Stack EC, Goodrich S, et al. (2009) Combined riluzole and sodium phenylbutyrate therapy in transgenic amyotrophic lateral sclerosis mice. *Amyotroph Lateral Scler* 10: 85–94.
66. Derave W, Van Den Bosch L, Lemmens G, Eijnde BO, Robberecht W, et al. (2003) Skeletal muscle properties in a transgenic mouse model for amyotrophic lateral sclerosis: effects of creatine treatment. *Neurobiol Dis* 13: 264–272.
67. Ende N, Weinstein F, Chen R, Ende M, Ende N (2000) Human umbilical cord blood effect on sod mice (amyotrophic lateral sclerosis). *Life Sci* 67: 53–59.
68. Ermilova IP, Ermilov VB, Levy M, Ho E, Pereira C, et al. (2005) Protection by dietary zinc in ALS mutant G93A SOD transgenic mice. *Neurosci Lett* 379: 42–46.
69. Esposito E, Rossi C, Amodio R, Di Castelnuovo A, Bendotti C, et al. (2000) Lyophilized red wine administration prolongs survival in an animal model of amyotrophic lateral sclerosis. *Ann Neurol* 48: 686–687.
70. Esposito E, Capasso M, di Tomasso N, Corona C, Pellegrini F, et al. (2007) Antioxidant strategies based on tomato-enriched food or pyruvate do not affect disease onset and survival in an animal model of amyotrophic lateral sclerosis. *Brain Res Sep 7*;1168: 90–6 Epub 2007 Jul 31.
71. Ferrante RJ, Klein AM, Dedeoglu A, Beal MF (2001) Therapeutic efficacy of EGb761 (Ginkgo biloba extract) in a transgenic mouse model of amyotrophic lateral sclerosis. *J Mol Neurosci* 17: 89–96.
72. Fischer LR, Culver DG, Davis AA, Tennant P, Wang M, et al. (2005) The WldS gene modestly prolongs survival in the SOD1G93A ALS mouse. *Neurobiol Dis* 19: 293–300.
73. Ghodoussi F, Galloway MP, Jambekar A, Bame M, Needleman R, et al. (2010) Methionine sulfoximine, an inhibitor of glutamine synthetase, lowers brain glutamine and glutamate in a mouse model of ALS. *J Neurol Sci* 290: 41–47.
74. Gifondorwa DJ, Robinson MB, Hayes CD, Taylor AR, Prevette DM, et al. (2007) Exogenous delivery of heat shock protein 70 increases lifespan in a mouse model of amyotrophic lateral sclerosis. *J Neurosci* 27: 13173–13180.
75. Groeneveld GJ, de Leeuw van Weenen J, van Muiswinkel FL, Veldman H, Veldink JH, et al. (2003) Zinc amplifies mSOD1-mediated toxicity in a transgenic mouse model of amyotrophic lateral sclerosis. *Neurosci Lett* 352: 175–178.
76. Groeneveld GJ, Van Muiswinkel FL, Sturkenboom JM, Wokke JH, Bar PR, et al. (2004) Ovariectomy and 17beta-estradiol modulate disease progression of a mouse model of ALS. *Brain Res* 1021: 128–131.
77. Gros-Louis F, Soucy G, Lariviere R, Julien JP, Gros-Louis F, et al. (2010) Intracerebroventricular infusion of monoclonal antibody or its derived Fab fragment against misfolded forms of SOD1 mutant delays mortality in a mouse model of ALS. *J Neurochem* 113: 1188–1199.

78. Gurney ME, Fleck TJ, Himes CS, Hall ED (1998) Riluzole preserves motor function in a transgenic model of familial amyotrophic lateral sclerosis. *Neurology* 50: 62–66.
79. Gurney ME, Cutting FB, Zhai P, Doble A, Taylor CP, et al. (1996) Benefit of vitamin E, riluzole, and gabapentin in a transgenic model of familial amyotrophic lateral sclerosis. *Ann Neurol* 39: 147–157.
80. Habisch HJ, Schwalenstocker B, Danzeisen R, Neuhaus O, Hartung HP, et al. (2007) Limited effects of glatiramer acetate in the high-copy number hSOD1-G93A mouse model of ALS. *Exp Neurol* 206: 288–295.
81. Haenggeli C, Julien JP, Mosley RL, Perez N, Dhar A, et al. (2007) Therapeutic immunization with a glatiramer acetate derivative does not alter survival in G93A and G37R SOD1 mouse models of familial ALS. *Neurobiol Dis* 26: 146–152.
82. Hamadeh MJ, Tarnopolsky MA (2006) Transient caloric restriction in early adulthood hastens disease endpoint in male, but not female, Cu/Zn-SOD mutant G93A mice. *Muscle Nerve* 34: 709–719.
83. Hamadeh MJ, Rodriguez MC, Kaczor JJ, Tarnopolsky MA (2005) Caloric restriction transiently improves motor performance but hastens clinical onset of disease in the Cu/Zn-superoxide dismutase mutant G93A mouse. *Muscle Nerve* 31: 214–220.
84. Ito H, Wate R, Zhang J, Ohnishi S, Kaneko S, et al. (2008) Treatment with edaravone, initiated at symptom onset, slows motor decline and decreases SOD1 deposition in ALS mice. *Exp Neurol* 213: 448–455.
85. Jaarsma D, Guchelaar HJ, Haasdijk E, de Jong JM, Holstege JC (1998) The antioxidant N-acetylcysteine does not delay disease onset and death in a transgenic mouse model of amyotrophic lateral sclerosis. *Ann Neurol* 44: 293.
86. Jiang F, DeSilva S, Turnbull J (2000) Beneficial effect of ginseng root in SOD-1 (G93A) transgenic mice. *J Neurol Sci* 180: 52–54.
87. Joo IS, Hwang DH, Seok JJ, Shin SK, Kim SU (2007) Oral administration of memantine prolongs survival in a transgenic mouse model of amyotrophic lateral sclerosis. *J Clin Neurol* 3: 181–186.
88. Kalmar B, Novoselov S, Gray A, Cheetham ME, Margulis B, et al. (2008) Late stage treatment with arimocloamol delays disease progression and prevents protein aggregation in the SOD1 mouse model of ALS. *J Neurochem* 107: 339–350.
89. Kiaei M, Kipiani K, Chen J, Calingasan NY, Beal MF (2005) Peroxisome proliferator-activated receptor-gamma agonist extends survival in transgenic mouse model of amyotrophic lateral sclerosis. *Exp Neurol* 191: 331–336.
90. Kieran D, Kalmar B, Dick JR, Riddoch-Contreras J, Burnstock G, et al. (2004) Treatment with arimocloamol, a coinducer of heat shock proteins, delays disease progression in ALS mice. *Nat Med* 10: 402–405.
91. Kim K, Moore DH, Makriyannis A, Abood ME (2006) AM1241, a cannabinoid CB2 receptor selective compound, delays disease progression in a mouse model of amyotrophic lateral sclerosis. *Eur J Pharmacol* 542: 100–105.
92. Kira Y, Nishikawa M, Ochi A, Sato E, Inoue M (2006) L-carnitine suppresses the onset of neuromuscular degeneration and increases the life span of mice with familial amyotrophic lateral sclerosis. *Brain Res* 1070: 206–214.
93. Kirkinetzos IG, Hernandez D, Bradley WG, Moraes CT (2003) Regular exercise is beneficial to a mouse model of amyotrophic lateral sclerosis. *Ann Neurol* 53: 804–807.
94. Klivenyi P, Ferrante RJ, Matthews RT, Bogdanov MB, Klein AM, et al. (1999) Neuroprotective effects of creatine in a transgenic animal model of amyotrophic lateral sclerosis. *Nat Med* 5: 347–350.
95. Koh SH, Kim Y, Kim HY, Cho GW, Kim KS, et al. (2007) Recombinant human erythropoietin suppresses symptom onset and progression of G93A-SOD1 mouse model of ALS by preventing motor neuron death and inflammation. *Eur J Neurosci* 25: 1923–1930.
96. Koh SH, Kim Y, Kim HY, Hwang S, Lee CH, et al. (2007) Inhibition of glycogen synthase kinase-3 suppresses the onset of symptoms and disease progression of G93A-SOD1 mouse model of ALS. *Exp Neurol* 205: 336–346.
97. Koh SH, Lee SM, Kim HY, Lee KY, Lee YJ, et al. (2006) The effect of epigallocatechin gallate on suppressing disease progression of ALS model mice. *Neurosci Lett* 395: 103–107.
98. Shaw PJ (1999) Motor neurone disease. *BMJ* 318: 1118–1121.
99. Martin LJ (1999) Neuronal death in amyotrophic lateral sclerosis is apoptosis: possible contribution of a programmed cell death mechanism. *J Neuropathol Exp Neurol* 58: 459–471.
100. Gurney ME, Pu H, Chiu AY, Dal Canto MC, Polchow CY, et al. (1994) Motor neuron degeneration in mice that express a human Cu,Zn superoxide dismutase mutation. [Erratum appears in *Science* 1995 Jul 14;269(5221):149]. *Science* 264: 1772–1775.
101. Canadian Council on Animal Care (1998) CCAC guidelines on: choosing an appropriate endpoint in experiments using animals for research, teaching and testing.
102. Guo YS, Wu DX, Wu HR, Wu SY, Yang C, et al. (2009) Sensory involvement in the SOD1-G93A mouse model of amyotrophic lateral sclerosis. *Exp Mol Med* 41: 140–150.
103. Orrell RW, King AW, Hilton DA, Campbell MJ, Lane RJ, et al. (1995) Familial amyotrophic lateral sclerosis with a point mutation of SOD-1: intrafamilial heterogeneity of disease duration associated with neurofibrillary tangles. *J Neurol Neurosurg Psychiatry* 59: 266–270.
104. Seevaratnam R, Raha S, Tarnopolsky MA, Hamadeh MJ (2009) Coffee increases antioxidant enzyme capacity in the brain of male G93A mice, an animal model of amyotrophic lateral sclerosis (ALS). *FASEB J* 23: 109.6.
105. Seevaratnam R, Raha S, Tarnopolsky MA, Hamadeh MJ (2009) Caffeine reduces motor performance and antioxidant enzyme capacity in the brain of female G93A mice, an animal model of amyotrophic lateral sclerosis (ALS). *FASEB J* 23: 963.3.
106. Sugai F, Yamamoto Y, Miyaguchi K, Zhou Z, Sumi H, et al. (2004) Benefit of valproic acid in suppressing disease progression of ALS model mice. *Eur J Neurosci* 20: 3179–3183.
107. Holzbaur EL, Howland DS, Weber N, Wallace K, She Y, et al. (2006) Myostatin inhibition slows muscle atrophy in rodent models of amyotrophic lateral sclerosis. *Neurobiol Dis* 23: 697–707.
108. Kawamura Y, Dyck PJ, Shimono M, Okazaki H, Tateishi J, et al. (1981) Morphometric comparison of the vulnerability of peripheral motor and sensory neurons in amyotrophic lateral sclerosis. *J Neuropathol Exp Neurol* 40: 667–675.
109. Tateishi T, Hokonohara T, Yamasaki R, Miura S, Kikuchi H, et al. (2010) Multiple system degeneration with basophilic inclusions in Japanese ALS patients with FUS mutation. *Acta Neuropathol* 119: 355–364.
110. Sasaki S, Horie Y, Iwata M (2007) Mitochondrial alterations in dorsal root ganglion cells in sporadic amyotrophic lateral sclerosis. *Acta Neuropathol (Berl)* 114: 633–639.
111. Hirano A, Kurland LT, Sayre GP (1967) Familial amyotrophic lateral sclerosis. A subgroup characterized by posterior and spinocerebellar tract involvement and hyaline inclusions in the anterior horn cells. *Arch Neurol* 16: 232–243.
112. Suzuki M, Irie T, Watanabe T, Mikami H, Yamazaki T, et al. (2008) Familial amyotrophic lateral sclerosis with Gly93Ser mutation in Cu/Zn superoxide dismutase: a clinical and neuropathological study. *J Neurol Sci* 268: 140–144.
113. Dal Canto MC, Gurney ME (1994) Development of central nervous system pathology in a murine transgenic model of human amyotrophic lateral sclerosis. *Am J Pathol* 145: 1271–1279.
114. Butterfield RJ, Blankenhorn EP, Roper RJ, Zachary JF, Doerge RW, et al. (1999) Genetic Analysis of Disease Subtypes and Sexual Dimorphisms in Mouse Experimental Allergic Encephalomyelitis (EAE): Relapsing/Remitting and Monophasic Remitting/Nonrelapsing EAE Are Immunogenetically Distinct. *J Immunol* 162: 3096–3102.
115. Fleming KK, Bovaird JA, Mosier MC, Emerson MR, LeVine SM, et al. (2005) Statistical analysis of data from studies on experimental autoimmune encephalomyelitis. *J Neuroimmunol* 170: 71–84.
116. Thakker P, Leach MW, Kuang W, Benoit SE, Leonard JP, et al. (2007) IL-23 Is Critical in the Induction but Not in the Effector Phase of Experimental Autoimmune Encephalomyelitis. *J Immunol* 178: 2589–2598.