Santiago Ramón y Cajal (1852-1934)

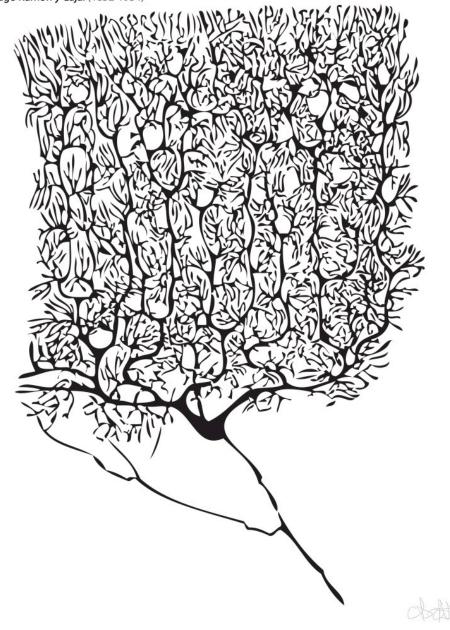
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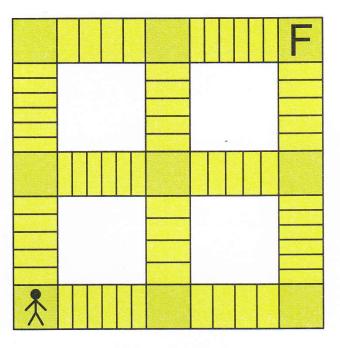
York University
Winter 2020
BPHS 4080 Lecture 10



Reference/Acknowledgement:

- TF Weiss (Cellular Biophysics)
- D Freeman





What is the smallest possible number of steps needed to reach floor F? Note that the square platforms are not counted as steps.

Resting Potential -40 $V_m(t)$ $V_m(t)$ $V_m(t) \; (\mathrm{mV})$ 50 Bath -60Bath composition Test 2 Normal Normal 0 solution solution solution Time (s) $(V_m)_m (t)_m -100 - 0$ -60 Figure 1.3 NaCl Q-HCl 20 $V_m(t) - V_m^o \text{ (mV)}$ 20 40 60 Time (s) Figure 1.2 HCl Sucrose $\int V_m(t)$ $V_m(t) \; (\mathrm{mV})$ x(t)0 Ó 100 100 $(m\pi)$ (t)x = 0 (t)x = 1.5Time (s) Figure 1.6 $5~\mu\mathrm{m}$

100

50

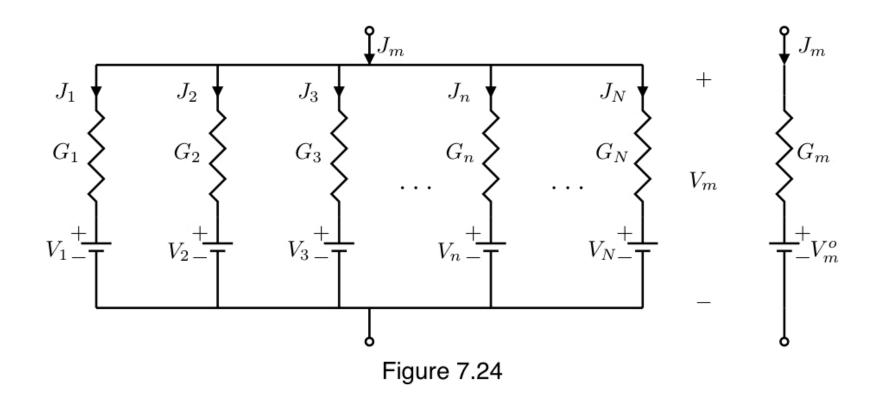
Time (ms)

0

Figure 1.5

→ What is the basis for such a resting potential?

→ What if different ions are able to diffuse?



K⁺ contribuition?

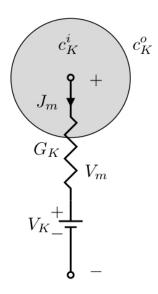
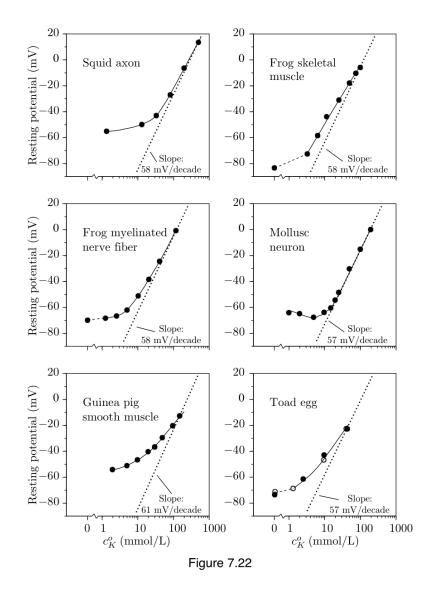


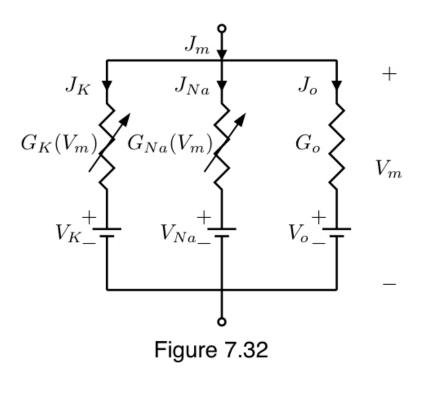
Figure 7.18

$$V_m^o = \frac{RT}{F} \left(\frac{G_K}{G_m} \right) \ln \left(\frac{c_K^o}{c_K^i} \right) + \sum_{n \neq K} \frac{G_n}{G_m} V_n$$

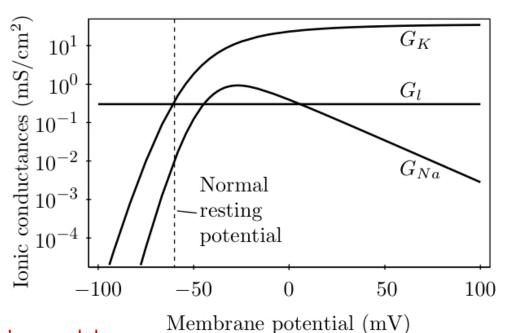


→ Chief issue is deviations at lower K⁺ concentrations (i.e., saturation in resting potential)

What if conductances were voltage-dependent?



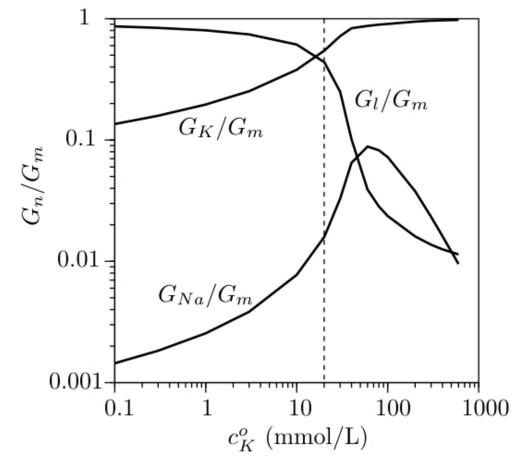
i.e., voltage-gated ion channels (more detail in vol.2 ch.6)



Hodgkin-Huxley model conductances

Figure 7.28

$$V_m^o = \frac{RT}{F} \left(\frac{G_K}{G_m} \right) \ln \left(\frac{c_K^o}{c_K^i} \right) + \sum_{n \neq K} \frac{G_n}{G_m} V_n$$



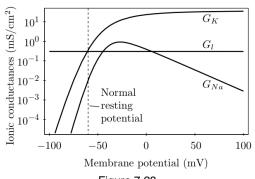
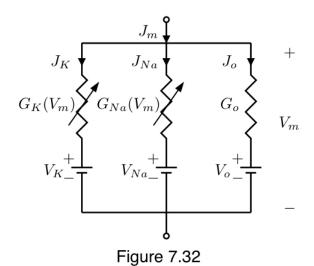


Figure 7.28

Figure 7.30

→ For physiological K+ concentrations, potassium is a dominant ion



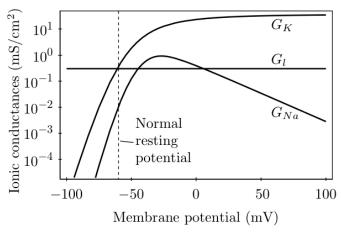
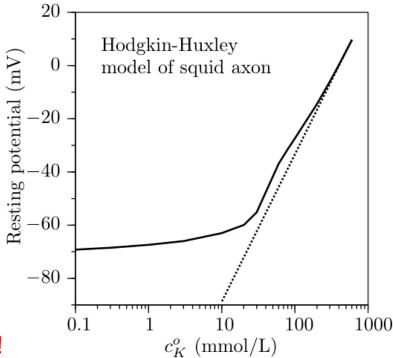


Figure 7.28

$$\sum_{n} G_n(V_m^o - V_n) = 0$$



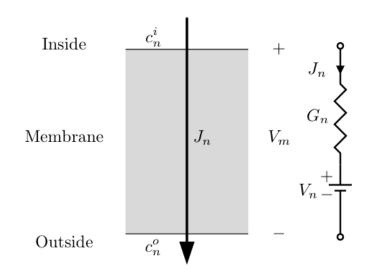
→ This looks like it might work!

Figure 7.29

<u>Is a purely passive model adequate?</u>

Inside cell: high [K⁺], low [Na⁺] Outside cell: low [K⁺], high [Na⁺]

Ion	$G_n (\mathrm{S/cm}^2)$	G_n/G_m	c_n^o/c_n^i	$V_n \text{ (mV)}$
K^+	3.7×10^{-4}	0.55	0.05	-72
Na ⁺	1×10^{-5}	0.016	9.8	+55
leakage	3.0×10^{-4}	0.44		-49



$$J_K = G_K(V_m^o - V_K) = 0.37 \times 10^{-3} (-60 + 72) \times 10^{-3} \approx +4 \,\mu\text{A/cm}^2$$

K⁺ efflux Na⁺ influx

$$J_{Na} = G_{Na}(V_m^o - V_{Na}) = 1 \times 10^{-5}(-60 - 55) \times 10^{-3} \approx -1 \,\mu\text{A/cm}^2$$

→ Passive case unsustainable! (eventually leads to equilibrium)

$$\frac{RT}{z_1F}\ln\left(\frac{c_1^o}{c_1^i}\right) = \frac{RT}{z_2F}\ln\left(\frac{c_2^o}{c_2^i}\right) = \dots = \frac{RT}{z_nF}\ln\left(\frac{c_n^o}{c_n^i}\right)$$

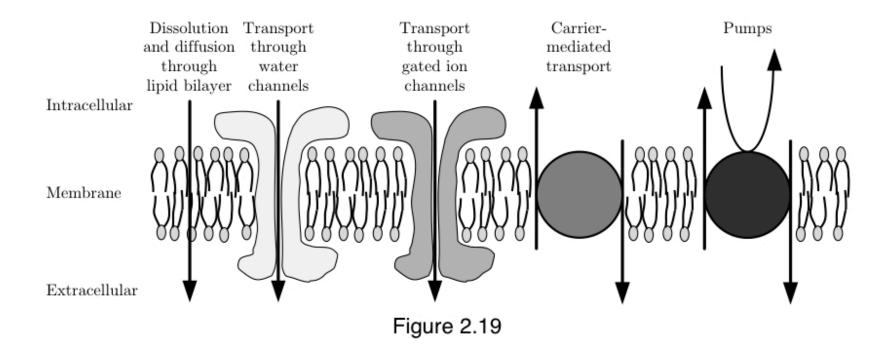
[osmotic effects also important here]

Table 7.4 Net flux of ions across the membranes of nerve axons during a propagated action potential (Cohen and De Weer, 1977). The ion fluxes are given per action potential.

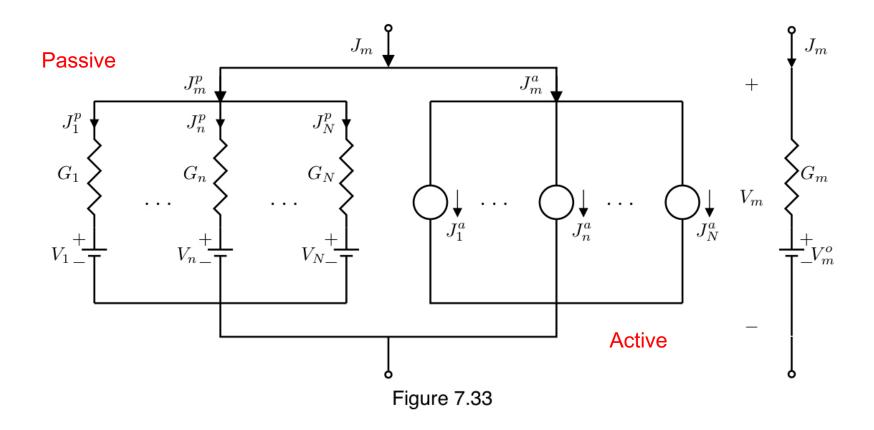
Preparation	K ⁺ efflux	Na ⁺ influx	
	(pmol/cm ²)		
Loligo forbesi axon	3.0	3.5	
Loligo pealei axon	3.7		
Sepia officinalis axon	3.6	3.8	
Homarus nerve	4.1	5.2	
Carcinus nerve	1.7-20	_	
Rabbit vagus nerve	1	_	

→ Action potentials make it even worse!

Ion Pumps



→ Need an 'active' (i.e., metabolically-dependent) mechanism to maintain normal physiological charge separation (e.g., Na⁺/K⁺ *pump*)



→ 'Active' implies energy is used to pump (i.e., create a current) against the electrochemical gradient

[see Weiss sec.6.4.3]

[Modify SS4 model] Suppose the two disassociation constants differ

$$\mathfrak{N}_{ES}^i = \frac{c_S^i}{c_S^i + K_i} \mathfrak{N}_{ET}$$
, and $\mathfrak{N}_{ES}^o = \frac{c_S^o}{c_S^o + K_o} \mathfrak{N}_{ET}$

$$\phi_S = (\phi_S)_{max} \left(\frac{c_S^i}{c_S^i + K_i} - \frac{c_S^o}{c_S^o + K_o} \right)$$

$$\phi_{S} = (\phi_{S})_{max} \frac{K_{o}c_{S}^{i} - K_{i}c_{S}^{o}}{(c_{S}^{i} + K_{i})(c_{S}^{o} + K_{o})}$$

Now suppose $c_S^i > c_S^o$, but $K_o c_S^i < K_i c_S^o$: then $\phi_S < 0$

→ Why though would such (i.e., transport up the gradient) "require" energy?

Simple, Symmetric Four-State Model

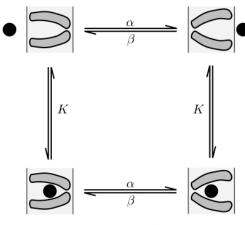


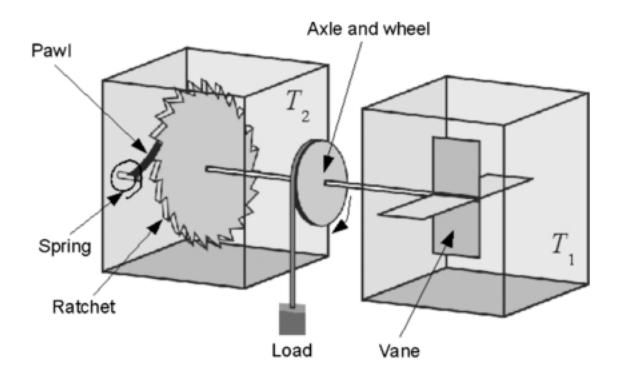
Figure 6.20

$$\begin{split} \mathfrak{N}_{ES}^{i} &= \left(\begin{array}{c} \beta \\ \overline{\alpha + \beta} \end{array} \right) \left(\frac{c_{S}^{i}}{c_{S}^{i} + K} \right) \mathfrak{N}_{ET} \\ \mathfrak{N}_{E}^{i} &= \left(\frac{\beta}{\alpha + \beta} \right) \left(\frac{K}{c_{S}^{i} + K} \right) \mathfrak{N}_{ET} \\ \mathfrak{N}_{ES}^{o} &= \left(\frac{\alpha}{\alpha + \beta} \right) \left(\frac{c_{S}^{o}}{c_{S}^{o} + K} \right) \mathfrak{N}_{ET} \\ \mathfrak{N}_{E}^{o} &= \left(\frac{\alpha}{\alpha + \beta} \right) \left(\frac{K}{c_{S}^{o} + K} \right) \mathfrak{N}_{ET} \end{split}$$

$$\phi_S = \left(\frac{\alpha\beta}{\alpha + \beta}\right) \mathfrak{N}_{ET} \left(\frac{c_S^i}{c_S^i + K} - \frac{c_S^o}{c_S^o + K}\right)$$

Why "active"?

Brownian ratchet (Feynman-Smoluchowski ratchet)



- Version of "Maxwell's demon"
- ➤ Violation of 2nd Law of Thermodynamics?

No (becuase it doesn't work)

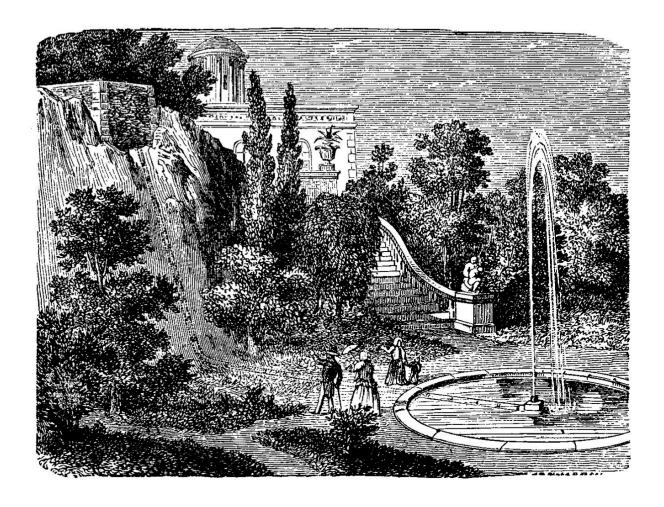


Figure 11.3: (Metaphor.) If water is continuously pumped to the upper reservoir, the fountain will come to a nonequilibrium steady state. If not, it will come to a quasisteady state, which lasts until the reservoir is empty.

Why "active"?

Here is a mechanical analogy: Suppose you visit your friend and see in his garden a fountain. The fountain could be supplied by a high tank of water (Figure 11.3). In that case it flows, converting the gravitational potential energy of the water in the tank to kinetic energy (and ultimately heat), until the tank is empty; that is, it drives to equilibrium. But if you watch the fountain for many hours and it never stops, you may begin to suspect that your friend instead recirculates the water with a *pump*, using some external source of energy. In that case the fountain is in a steady, but nonequilibrium, state.

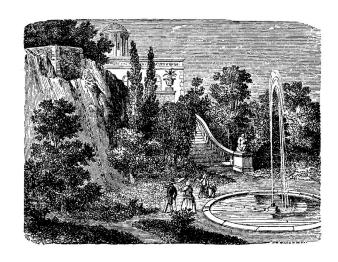


Figure 11.3: (Metaphor.) If water is continuously pumped to the upper reservoir, the fountain will come to a nonequilibrium steady state. If not, it will come to a quasisteady state, which lasts until the reservoir is empty.

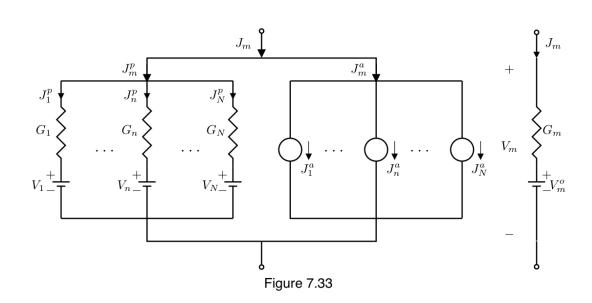
Inside cell: high [K⁺], low [Na⁺] Outside cell: low [K⁺], high [Na⁺]

$$\frac{RT}{z_1 F} \ln \left(\frac{c_1^o}{c_1^i} \right) = \frac{RT}{z_2 F} \ln \left(\frac{c_2^o}{c_2^i} \right) = \dots = \frac{RT}{z_n F} \ln \left(\frac{c_n^o}{c_n^i} \right)$$

Idea: Cell is not strictly in equilibrium

Ion Pumps

 Active pumps allow for cellular quasi-equilibrium



$$J_n^p + J_n^a = 0$$

Implies concentration on *n* does not change

(J^a is the ion pump term)

uniform cell at rest:

$$J_m = J_m^p + J_m^a = 0$$

$$\sum_{n} G_n (V_m^o - V_n) + \sum_{n} J_n^a = 0$$

$$V_m^o = \sum_n \frac{G_n}{G_m} V_n - \frac{1}{G_m} \sum_n J_n^a$$

$$J_m = \sum_n G_n(V_m - V_n) + \sum_n J_n^a = G_m \left(V_m - \left(\sum_n \frac{G_n}{G_m} V_n - \frac{1}{G_m} \sum_n J_n^a \right) \right)$$

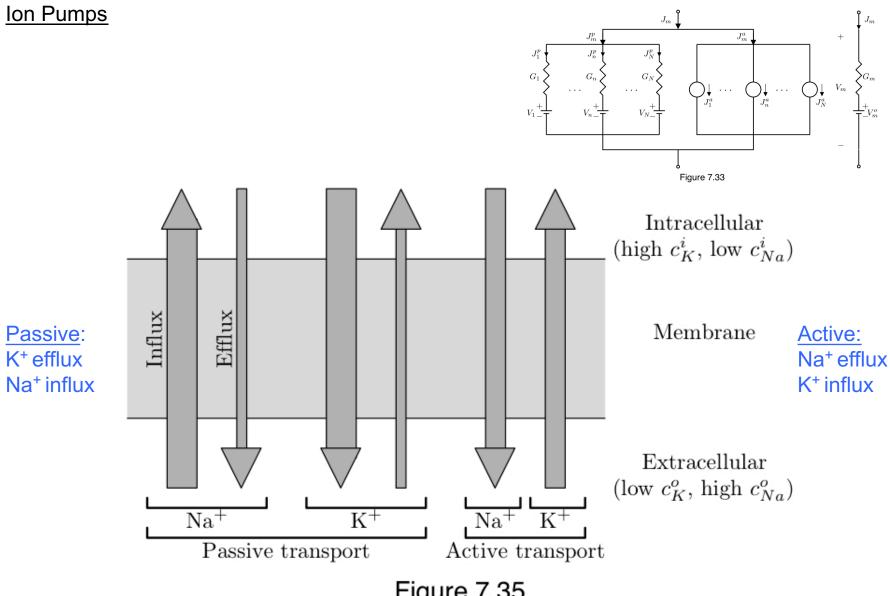
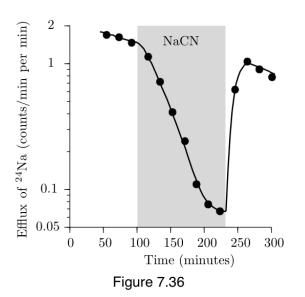


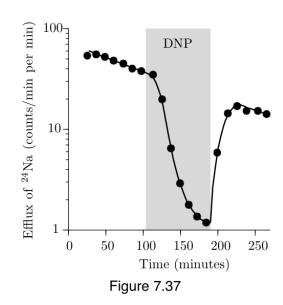
Figure 7.35

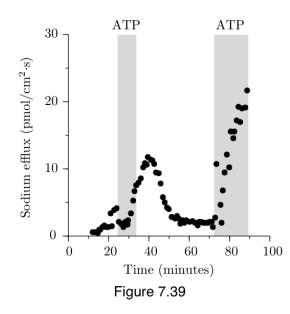
→ Empirical means to differentiate: Examine Na+ efflux

Na+ efflux is metabolically dependent

- Axons pre-loaded w/ radioactive ²⁴Na⁺ (left, center) or perfused w/ dialysis tubing (right)
- Light grey bars indicate addition of metabolic poisons (left, center) or ATP (right) ("the molecular unit of currency of intracellular energy transfer", Wikipedia)
- Plots use a proxy measure for Na⁺ efflux ("The change in count rate with time was proportional to efflux of radioactive Na")
- Refs: Hodgkin & Keynes (1955), Mullins & Brinley (1967)

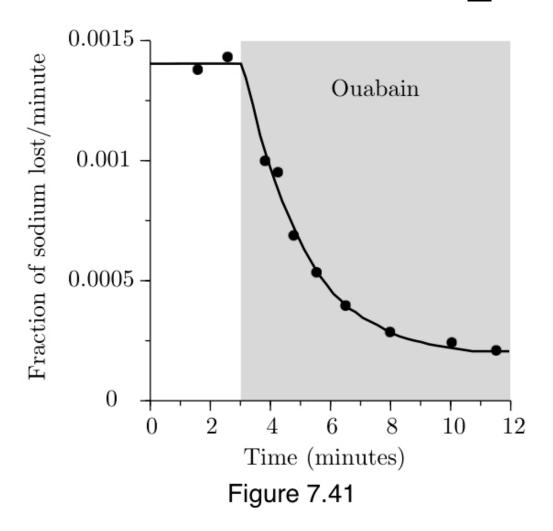






- Note decreasing in efflux before poisons (due to limited amount of tagged Na)
- Note distinction between "reversible" and irreversible" effects
 - → Taken together, data are suggestive that efflux is metabolically-dependent

Ref: Baker & Willis (1972)



→ Cardiac glycosides (e.g., ouabain) inhibit/block Na⁺/K⁺ pump

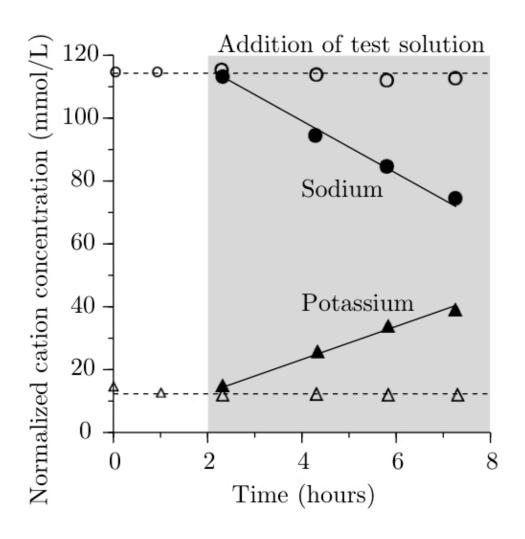
(though these drugs had long been used to treat heart failure, it wasn't until ~1950s that it was understood how)

Pump Dependence Upon Ion Concentration

Are pumping of Na⁺ and K⁺ linked?

- Human erythrocytes (i.e., red blood cells) preloaded w/ high [Na⁺] and low [K⁺]
- Circles represent [Na⁺] measurements
- Triangles represent [K⁺] measurements
- Grey bar indicates change from low [Na⁺]
- Filled symbols 'test solution' containing K⁺
- Open symbols 'control solution' with no K⁺
- Ref: Post & Jolly (1957)

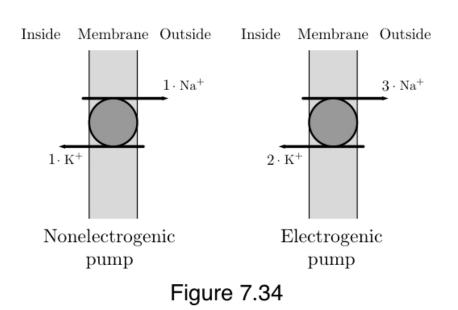
No external K⁺ means no Na⁺ efflux



→ Pump activity linked to concentration of both Na⁺ and K⁺

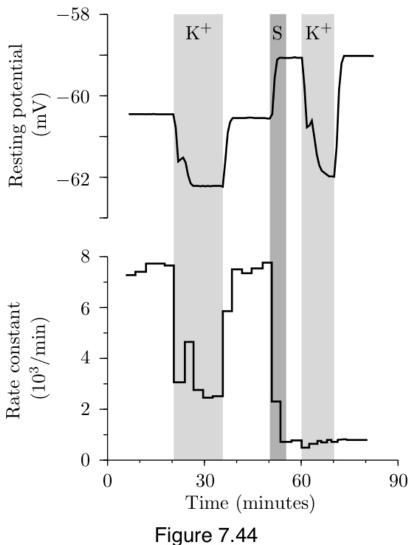
Figure 7.42

Ion Pumps



→ Does a pump contribute to the membrane potential?

(i.e., does active Na⁺ efflux exceed active K⁺ influx?)



De Weer & Geduldig (Science, 1973)

- Axon pre-loaded w/ radioactive ²²Na⁺
- Light grey bars indicate reduced external K⁺ concentration
- Dark grey bars indicate addition of strophanthidin (S) ["a cardiac glycoside which mechanism of action is similar to Digitalis, Ouabain and digitoxin. It specifically inhibits the membrane protein Na+/K+ ATPase", wikipedia]
- Top shows resting potential, bottom a proxy measure for Na⁺ efflux (via the radioactive rate count)
- Ref: De Weer & Geduldig (Science, 1973)
- → Pump inhibition (via strophanthidin or change in [K⁺]) indicates pump causes several mV of hyperpolarization
- → Need to be careful experimentally when potentially (pun!) affecting both active and passive transport mechanisms

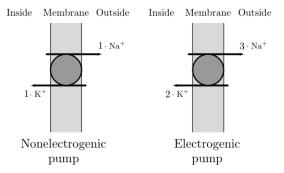


Figure 7.34

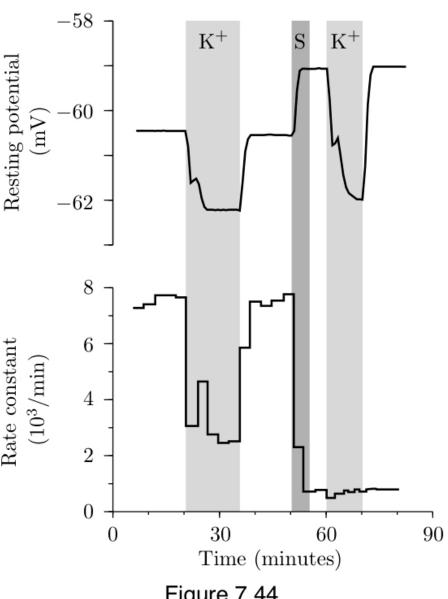


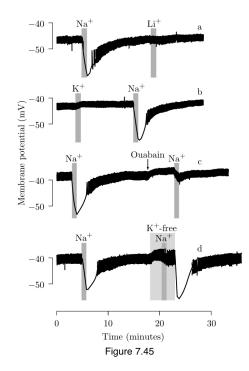
Figure 7.44

Electrogenicity of Na⁺/K⁺ pump?

Does active efflux of Na⁺ exceed active influx of K⁺?

$$V_m^o = \sum_n \frac{G_n}{G_m} V_n - \frac{1}{G_m} \sum_n J_n^a$$

Can stimulating pumps cause larger electrogenic effect?



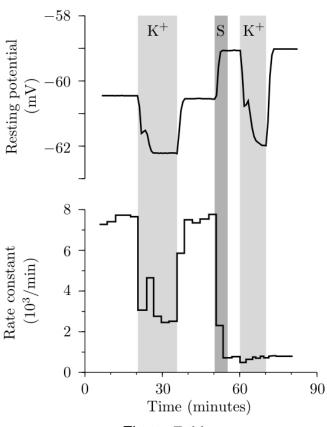


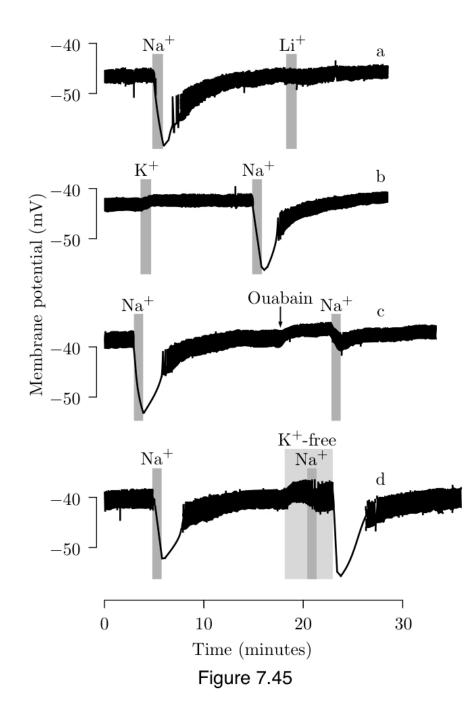
Figure 7.44

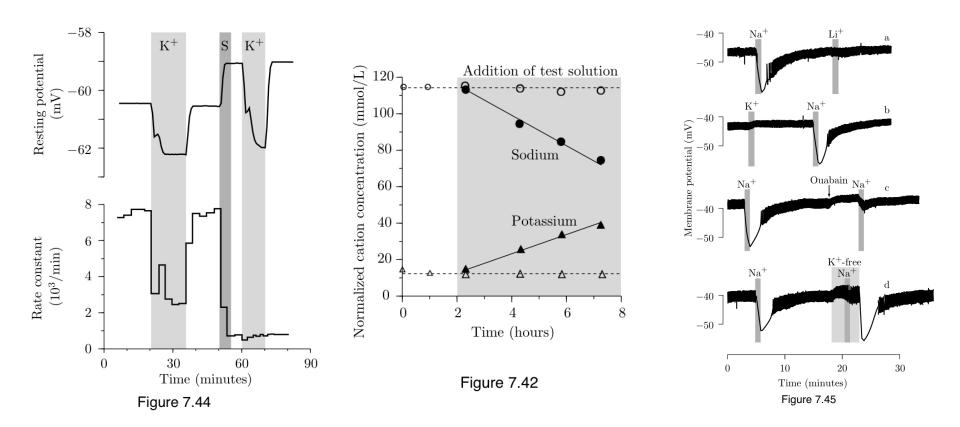
Electrogenicity of Na+/K+ pump?

- Snail neurons impaled by three pipettes: one to measure membrane potential and two to inject (intracellularly) a specific ion type
- Grey bars indicate ion injection
- Bottom traces also includes condition where external K⁺ is removed
- Ref: Thomas (1969)

→ Injecting Na+ (but not K+ or Li+) intracellularly hyperpolarizes

→ Blocking the pump (via ouabain) or removing K⁺ abolishes the effect





→ Evidence leading towards the notion of the Na⁺/K⁺ pump

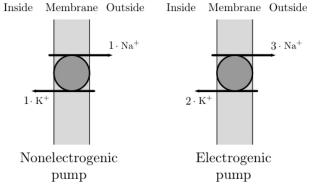


Figure 7.34

(Na⁺-K⁺)-ATPase

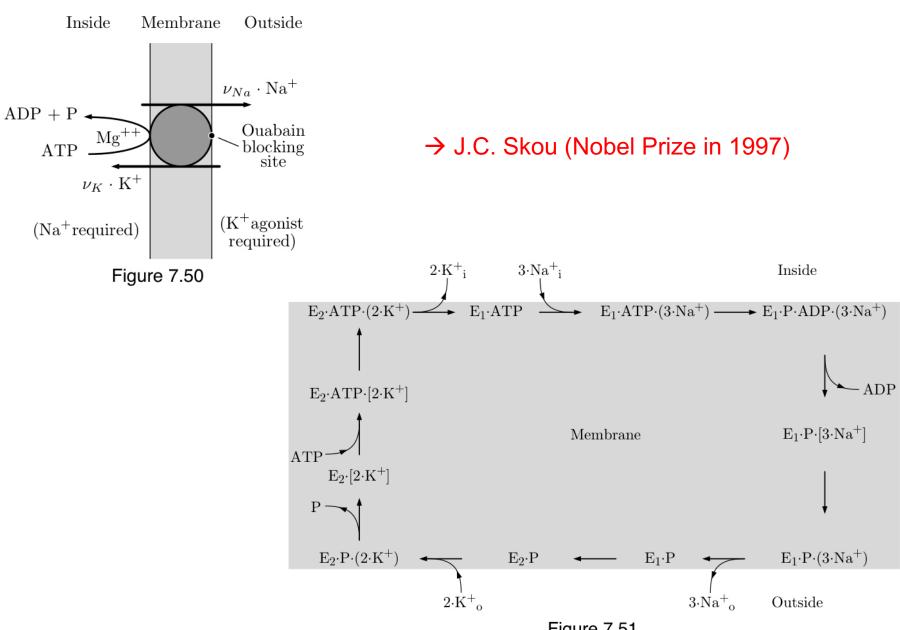
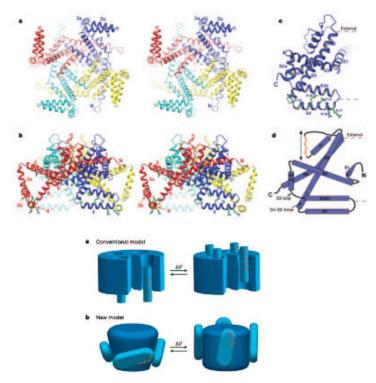


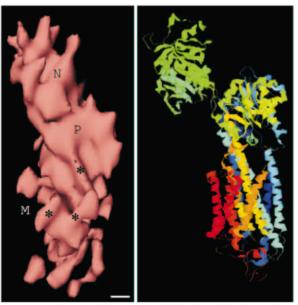
Figure 7.51

Family of related "channels"



Y. Jiang, A. Lee, J. Chen, V. Ruta, M. Cadene, B. Chalt, and R. MacKinnon (2003), Nature 423:33-41.

Family of related "pumps"



H. Hebert, P. Purhonen, H. Vorum, K. Thomsen, and A.BB. Maunsback (2001), J. Mol. Biol. 314:479-494.

→X-ray crystallography & electron microscopy can reveal structure of voltage-gated channels (i.e., *integral membrane proteins*)

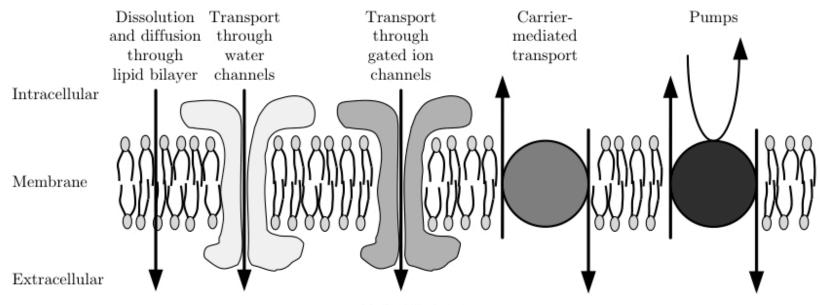
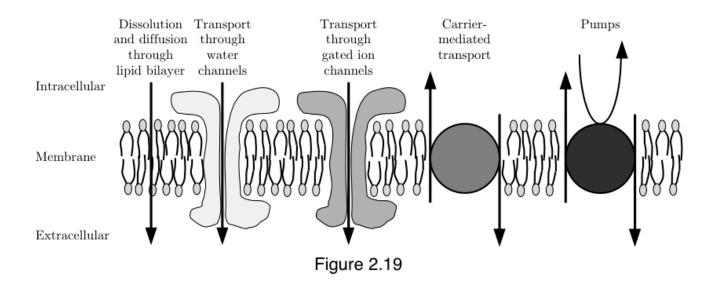
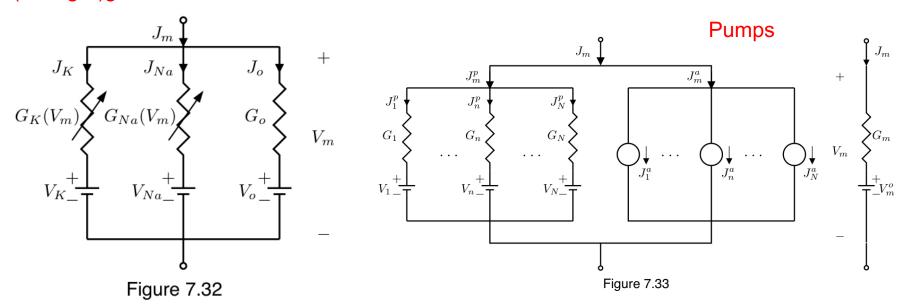


Figure 2.19



(Voltage-)gated ion channels



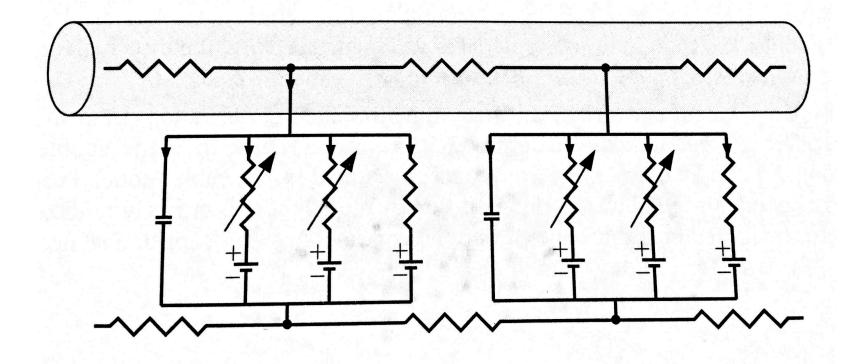


Fig.4.7 (vol.2)